Detection of Vibrio anguillarum and Vibrio alginolyticus by Randomly Cloned DNA Fragments

Yu-Ling Hsieh(1) and San-San Tsay(1,2)

(Manuscript received 30 July, 1999; accepted 8 September, 1999)

ABSTRACT: Vibrio anguillarum and Vibrio alginolyticus were used as tested organisms. Restriction endonuclease HindIII and EcoRI digested DNA fragments from the above two bacteria were randomly selected and inserted into vector pUC19. Among 0.5-2.0 kb DNA fragments were recovered from agarose gel to prepare non-radioactive DIG-labeled probes. One out of 102 cloned fragments could hybridize only to V. alginolyticus. Three out of 94 cloned fragments could hybridize only to V. anguillarum and one fragment is specific for serotype C.

KEY WORDS: Vibrio anguillarum, Vibrio alginolyticus, Colony hybridization, Randomly cloned fragments of DNA, Serotype.

INTRODUCTION

Vibrio anguillarum and V. alginolyticus are the major pathogens for both marine (Brunn and Heiberg, 1932; Nybelin, 1935; Rucker et al., 1953; Smith, 1961; Anderson and Conroy. 1970; Sindermann, 1970) and fresh water fishes (Ross et al., 1968; Kou et al., 1976; Huang, 1977; Tung et al., 1985) as well as shellfishes (Bowser et al., 1980). The detection and identification of these pathogens are important aspects of both diagnostic and therapy (Aoki et al., 1990). In the diagnosis of etiological agents, pathogenic microorganisms required selective media or special cultural conditions for growth. Identification of pathogens has used combinations of biochemical and serological tests after growth on agar media. These processes are time consuming and costly. DNA probe hybridization technology, widely used now for the direct detection and identification of microorganisms, provides rapid and accurate diagnosis.

This report describs probes isolated from the random fragments of chromosomal DNA are specific for *V. anguillarum* and *V. alginolyticus*.

MATERIALS AND METHODS

Bacteria strains and vectors

Vibrio species used in this study are listed in Table 1. Transformation recipient Escherichia coli RR1 was a gift from Molecular Biology Laboratory, and E. coli TG1 and vector pUC19 were from Genetic Laboratory, Department of Botany, National Taiwan University. V. anguillarum NIE 275 and V. alginolyticus ATCC 17749 were selected for the preparation of detection probes.

^{1.} Department of Botany, National Taiwan University, Taipei 106, Taiwan, Republic of China.

^{2.} Corresponding author. E-mail: sstsay@ccms.ntu.edu.tw

Table 1. Bacterial strains used in this study.

Strain no.	Species	No. of	Source or strains designation(s)*					
	O(1). • (1) (1) (1) (1) (1) (1) (1) (1) (1) (1)	strains						
	Vibrio							
1 - 6	V. alginolyticus	6	ATCC 17749, NPUST					
7 - 15	V. anguillarum	9	ATCC 19264, NTU-Z, T. Aoki					
16 - 17	V. campbellii	2	ATCC 25920, NTU-AC					
18 - 19	V. carchariae	2	NTU-Z					
20 - 25	V. damsela	6	ATCC 33539, NTU-Z, NPUST					
26	V. fischeri	1	FIRDI					
27	V. fluvialis	1	NTU-AC					
28 - 29	V. harveyi	2	ATCC-14126, NPUST					
30 - 32	V. mediterarranei	3	NPUST, NTU-Z					
33	V. minicus	1	NTU-AC					
34	V. natriegens	1	NTU-Z					
35	V. ordallii	1	NPUST					
36 - 38	V. parahaemolyticus	3	ATCC-27519, ATCC-27969, NPUST					
39	V. percolans	1	NTU-AC					
40	V. Proteolyticus	1	NPUST					
41	V. salmonicida	1	ATCC 43839					
42	V. splendidus	1	ATCC 33125					
43 - 44	V. vulnificus	2	NPUST					
45 - 61	V. spp.	17	NTU-Z, NTU-AC, NPUST,					
	Other genera							
62	Bacillus subtilus	1	NTU-B					
63 - 65	Escherichia coli	3	NTU-B					
66	Klebsiella pneumonia	1	NTU-AC					
67	Micrococcus lutea	1	NTU-AC					
68	Salmonella typhi	1	NTU-AC					
69	S. typhimurium	1	NTU-AC					
70	Staphylococcus aureus	1	NTU-AC					

^{*:} FIRDI: Culture Collection and Research Center (CCRC), the Food Industry Research and Development Institute, Institute, Taiwan, R.O.C.; NTU-B: Department of Botany, National Taiwan University; NTU-Z: Department of Zoology, National Taiwan University, Taipei, Taiwan, R.O.C.; NTU-AC: Department of Agricultural Chemistry, National Taiwan University, Taipei, Taiwan, R.O.C.; NPUST: Department of Veterinary Medicine, National Pingtung University of Science and Technology, Pingtung, Taiwan, R.O.C.; ATCC: American Type Culture Collection; T. Aoki: Department of Biological Resources, Miyazaki University, Miyazaki, Japan.

Culture condition

Tryptic soy agar (TSA) supplemented with 2% NaCl was used to culture all the *Vibrio* spp. at 30°C. Other bacterial cultures were grown on Luria-Bertani (LB) medium at 37°C.

Preparation of DNA

After organisms were grown overnight, the extraction and purification of bacterial chromosomal DNA were performed according to the method of Ausubel *et al.* (1995). Fragments of *Vibrio* DNA were generated by digestion of appropriate amount of chromosomal DNA with 2-3 units of *HindIII* or *EcoRI* for every one μ g DNA at 37°C overnight and separated by agarose gel electrophoresis (Ausubel *et al.*, 1995). Vector DNA was purified and similarly digested with the same restriction enzyme and then ligated with the *Vibrio* fragments using T4 DNA ligase (Ausubel *et al.*, 1995).

E. coli competent cell was prepared following the methods of Ausubel et al. (1995), and transformed with recombinant plasmids in the ligation mixture.

Construction of the DNA probe specific for V. anguillarum and V. alginolyticus

Random fragments of chromosomal DNA from *V. anguillarum* NIE 275 and *V. alginolyticus* ATCC 17749 were cloned in pUC19. Recombinant plasmids were digested with *Eco*R1 and *Hin*dIII, and DNA fragments with a size between 0.5 and 2 kb were purified and recovered by electroelution onto DEAE membrane (Ausubel *et al.*, 1995). The probes were labeled with digoxigenen (DIG) using random oligonucleotide primers (Sambrook *et al.*, 1989) according to the specifications of the manufacturer (DIG labeling kit, Boehringer Mannheim).

Colony hybridization

Tested organisms listed in Table 1 were grown on TSA supplemented with 2% NaCl and transfered to Nitroplus membrane (Sumbrook *et al.*, 1989). Colonies on Nitroplus membranes were lysed and detected by DIG-labeled probe with DIG detection kit (Boehringer Mannheim).

RESULTS

Screening of randomly cloned fragments specific for V. alginolyticus

V. alginolyticus ATCC 17749 was used as type organism for the preparation of probes. Recombinant plasmids containing DNA fragments from this strain were constructed after digestion of the bacteria and vector pUC19 DNA with EcoR1 and HindIII. Recombinant plasmids containing V, alginolyticus DNA fragments with a size between 0.5 and 2.0 kb were selected to hybridize Vibrio species DNA. The fragments hybridized only with type organism but not with other Vibrio species were selected. Nine out of 102 fragments purified from recombinant plasmids were collected (Fig. 1). Hybridization of these selected fragments with the collected V. alginolyticus strains were shown in Table 2. Probe 8 could hybridize with all the strains tested. All the probes could hybridize with strain numbers 1, 5 and 6.

Screening of randomly cloned fragments specific for V. anguillarum

V. anguillarum NIE275 native strain (serotype C) was selected as type organism. Recombinant plasmids containing DNA fragments from this strain were constructed after digestion of the bacteria and vector pUC19 DNA with EcoR1 and HindIII. Recombinant plasmids containing V. anguillarum DNA fragments with a size between 0.5 and 2.0 kb were selected to hybridize Vibrio species DNA. The fragments hybridized only with V. anguillarum but not with other Vibrio species were selected. Fourteen out of 94 fragments purified from recombinant plasmids were collected (Fig. 2). Hybridization of these selected fragments with the collected V. anguillarum strains were shown in Table 3. Probes 13, 17 and 19 could hybridize with all the tested V. anguillarum. Probe 23 could only hybridize with serotype C (strain numbers 9 and 10), but not with other serotypes. Probes 14 and 18 could hybridize with serotype C (strain numbers 9 and 10) and D (strain number 11). Probes 10 and 16 could hybridize with all the tested organisms except serotype B (strain number 8). Probes 12 and 22 could hybridize with all the strains but not with serotype F and H respectively. Probe 24, originally selected for serotype A by Dr. Aoki, could only hybridize with strain number 7.

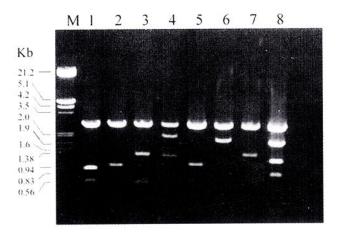


Fig. 1. Agarose gel electrophoresis of HindIII and EcoRI-digested DNA from pUC19 carrying V. alginolyticus fragment. M: λ digested by HindIII and EcoRI. Probe 1: second fragment of lane 1, about 0.83 kb; Probe 2: second fragment of lane 2; about 0.93 kb, Probe 3: second fragment of lane 3, about 1.38 kb; Probe 4: third fragment of lane 4, about 1.38 kb; Probe 5: second fragment of lane 5, about 1.0 kb; Probe 6: second fragment of lane 6, about 1.9 kb; Probe 7: second fragment of lane 7, about 1.38 kb; Probe 8: third fragment of lane 8, about 1.38 kb; Probe 9; forth fragment of lane 8, about 0.83 kb.

Table 2. Detection of V. alginolyticus by colony hybridization with randomly cloned DNA fragment.

Bacterial strain	Probe number ¹													
number	1	2	3	4	- 5	6	7	8	9					
1	+	+	+	+	+	+	+	+	+					
2		1-0	-	-	-	1071	-	+	-					
3	-	-	-	-	-	-	-	+	-					
4	+	-	+	+	+	+	+	+	+					
5	+	+	+	+	+	+	+	+	+					
6	+	+	+	+	+	+	+	+	+					

¹ The molecular weight of the probes is indicated in Fig. 1.

DISCUSSION

A probe (Macario and De Macario, 1990), used for the identification of microorganisms, can be a whole cell chromosomal DNA (Morotomi et al., 1988; Moncla et al., 1988; Roberts et al., 1987), specific genes (Moseley and Falkow, 1980; Moseley et al., 1982; Morris et al., 1987; Datta et al., 1987), ribosomal RNA (DeLong, et al., 1989; Romaniuk and Trust, 1987, Juha et al., 1994), plasmids (Totten et al., 1983; Horn et al., 1986), or oligonucleotides (Datta et al., 1988; Karch and Meyer, 1989; Miliotis et al., 1989; Lee et al., 1992). Even the randomly cloned fragments of chromosomal DNA were also used as the probe for the identification of Pasteurella piscida, V. anguillarum (Aoki et al., 1990), Campelobacter spp. (Totten et al., 1985), Leptospira interrogans (van Eys et al., 1988), Salmonella spp. (Fitts et al., 1983; Tompkins et al., 1986) and Bacteroides thetaitamicron (Salyers et al., 1986).

In this study, probe 8 was found to be specific for *V. alginolyticus* and probes 13, 17 as well as 19 for *V. anguillarum*. These probes did not hybridized with any other *Vibrio* species or with any other non-vibrio bacteria tested. Probe 23 was specific only for serotype C of *V. anguillarum*. During cross hybridization tests of the selected probes (data not shown), only probes 4 and 8 could hybridize with each other and showed similar hybridization patterns. The rest of the probes did not cross hybridize with any other probes selected. Since some of the tested organisms were gifts from other laboratories. In order to confirm our results, we had reidentified the tested organisms with Nissui ID test EB-20 (Nissui Pharmaceutical Co., Ltd., Tokyo, Japan). The identification results do match hybridization results (Hsieh, 1993). Thus, these optimally selected randomly cloned DNA fragments can be used as probes to identify those organisms whose specific gene probe is not available.

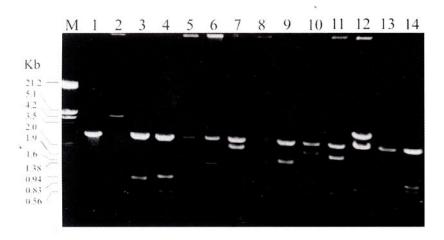


Fig. 2. Agarose gel electrophoresis of HindIII and EcoR1-digested DNA from pUC19 carrying V. anguillarum fragment. M: λ digested by HindIII and EcoR1. Probe 10: second fragment of lane 1, about 0.56 kb; Probe 11: second fragment of lane 2, about 0.56 kb; Probe 12: second fragment of lane 3, about 0.83 kb; Probe 13: second fragment of lane 4, about 0.94 kb; Probe 14: third fragment of lane 5, about 1.2 kb; Probe 15: second fragment of lane 6, about 1.38 kb; Probe 16: second fragment of lane 7, about 2.0 kb; Probe 17: second fragment of lane 8, about 1.38 kb; Probe 18: second fragment of lane 9, about 1.6 kb; Probe 19: second fragment of lane 10, about 2.0 kb; Probe 20: second fragment of lane 11, about 2.0 kb; Probe 21: second fragment of lane 12, about 0.94 kb; Probe 22: second fragment of lane 13, about 1.38 kb; Probe 23: second fragment of lane 14, about 1.2 kb.

Table 3. Detection of V. anguillarum by colony hybridization with randomly cloned DNA fragment.

Bacterial strain no.	Serotype	Probe number ¹														
		10	11	12	13	14	15	16	17	18	19	20	21	22	23	242
7	Α	+	+	+	+	-	+	+	+	-	+	-	141	+	-	+
8	В	2	-	+	+	-	-	_	+	-	+	+	-	+	_	-
9	C	+	+	+	+	+	+	+	+	+	+	+	+	+	+	2
10	C	+	+	+	+	+	+	+	+	+	+	+	+	+	+	~
11	D	+	-	+	+	+	-	+	+	+	+	+	+	+	_	-
12	E	+	_	+	+	_	+	+	+	-	+	+	+	+	-	2
13	F	+	+	-	+	_	+	+	+	-	+	+	-	+	- 1	2
14	Н	+	-	+	+	-	+	+	+	-	+	+	+	_	-	2
15	I	+	-	+	+	-	+	+	+	-	+	+	+	+	_	-

¹ The molecular weight of the probes is indicated in Fig. 2.

² probe number 24: 562 bp DNA fragment, gift from Dr. T. Aoki (Aoki et al., 1990).

ACKNOWLEDGMENT

This study was supported by a grant from the Council for Agricultural Planning and Development.

LITERATURE CITED

- Anderson, J. I. W. and D. A. Conroy. 1970. *Vibrio* disease in marine fishes. Am. Fish. Sco. Pub. **10**: 266-272.
- Aoki, T., I. Hirono, and J. Zhao. 1990. Identification of fish pathogenic bacteria using nucleic acid probes. Proceedings of ROC-Japan Symposium on Fish Diseases. pp. 53-58.
- Ausubel, F. M., R. Brend, R. E. Kingston, D. D. Moore, J. G. Seidman, J. A. Smith, and K. Struhl. (eds). 1995. Short Protocols in Molecular Biology. 3rd ed. John Wiley and Sons, Inc.
- Bowser, P. R., R. Rosemark, and C. R. Reiner. 1980. A preliminary report of vibriosis in cultured American lobsters, *Homoarus americanus*. J. Invertebr. Pathoi. 37: 80-85.
- Brunn, A. F. and B. Heiberg. 1932. The red disease of eels in Danish waters. Medd. Komm. Havunders Øg. Kbh. Ser. Fiskeri 9: 1-19.
- Datta, A. R., B. A. Wentz, and W. E. Hill. 1987. Detection of hemolytic *Listeria monocytogenes* by DNA colony hybridization. Appl. Environ. Microbiol. **53**: 2256-2259.
- DeLong, E. F., G. S. Wickham, and N. R. Pace. 1989. Phylogenic strains: ribosomal RNA-based probes for the identification of single cells. Science **243**: 1360-1363.
- Fitts, R., M. Diamond, C. Hamilton, and M. Nerik. 1983. DNA-DNA hybridization assay for detection of *Salmonella* spp. in foods. Appl. Environ. Microbiol. **46:** 1146-1151.
- Horn, J. E., M. L. Hammer, S. Falkow, and T. C. Quinn. 1986. Detection of *Chlamydia trachomatis* in tiss culture and cervical scrapings by *in situ* DNA hybridization. J. Infect. Dis. 153: 1155-1159.
- Hsieh, Y.-L. 1993. Detection of fish and shrimp pathogenic bacteria *Vibrio* spp. by colony hybridization. Master Thesis, National Taiwan University, Taipei, Taiwan, Republic of China
- Huang, Y.-H. 1977. Preliminary report of studies on bacterial disease of milkfish, *Chanos chanos* during winter. JCRR Fisheries Series **29**: 50-54.
- Juha, K., P. Jukka, and K. Marja-Leena. 1994. RFLP analysis of *Mycobacterium malmoense* strains using ribosomal RNA gene probes: an additional tool to examine intraspecies variations. J. Microbiol. Meth. **19**: 261-267.
- Karch, H. and T. Meyer. 1989. Evaluation of oligonuclotide for identification of shigh-like-toxin-producing *Escherichia coli*. J. Clin. Microbiol. **27**: 632-635.
- Kou, S.-C., H.-Y. Chung, and G.-H. Kou. 1976. *Vibrio anguillarum* isolated from a vibrio disease of fresh-water culture ayu, *Plecoglossus altivelis*. J. Fish Sco. Taiwan 4: 21-24.
- Lee, C., L.-H. Chen, M.-L. Liu, and Y-C. Su. 1992. Use of an oligonucleotide probe to detect *Vibrio parahaemolyticus* in artificially contaminated oysters. Appl. Environ. Microbiol. **58**: 3419-3422.
- Macario, A. J. L. and E. C. De Macario 1990. Gene probes for bacteria. Academic Press, San Diego.

- Miliotis, M. D., J. E. Galen, J. B. Kaper, and J. G. Morris, Jr. 1989. Development and testing of a synthetic oligonucleotide probe for the detection of pathogenic *Yersinia* strains. J. Clin. Microbiol. 27: 1667-1670.
- Moncla, B. J., L. Strockbine, P. Braham, J. Karlinsey, and M. C. Roberts. 1988. The of whole-cell DNA probes for the identification of *Bacteroides intermedius* isolates in a dot-blot assay. J. Dent. Res. 67: 1267-1270.
- Morotomi, M., T. Ohno, and M. Mutai. 1988. Rapid and correct identification of intestinal *Bacteroides* spp. with chromosomal DNA probes by whole-cell dot blot hybridization. Appl. Envrion. Microbiol. **54**: 1158-1162.
- Morris, J. G., A. C. Wright, D. M. Roberts, P. K. Wood, L. M. Simpson, and J. D. Oliver. 1987. Identification of environmental *Vibrio vulnificus* isolates with a DNA probe for the cytotoxin-hemolysin gene. Appl. Environ. Microbiol. **53**: 193-195.
- Moseley, S. L., I. Huq, A. R. M. Alim, M. So, M. Samadpour-Motalebi and S. Falkow. 1980. Detection of enterotoxigenic *Escherichia coli* by DNA colony hybridization. J. Infect. Dis. **142**: 892-898.
- Moseley, S. L. and S. Falkow. 1980. Nucleotide sequence homology between the heat-labile enterotoxin gene of *Escherichia coli* and *Vibrio cholerae* deoxyribonucleic acid. J. Bacteriol. **144**: 444-446.
- Moseley, S. L., P. Echeverria, J. Seriwatana, C. Tirapat, W. Chaicumpa, T. Sakuldaipeara and S. Falkow. 1982. Identification of enterotoxigenic *Escherichia coli* by colony hybridization using three enterotoxin gene probes. J. Infect. Dis. **145**: 863-869.
- Nybelin, O. 1935. Untersuchungen über den bei fishen Krankheiserregenden spaltpilz Vibrio anguillarum. Medd. Untersökn. Anst. Sötvattensfisk. Stoch. 8: 1-62.
- Roberts, M. C., C. McMillan and M. B. Coyle. 1987. Whole chromosomal DNA probes for rapid identification of *Mycobacterium tuberculosis* and *Mycobacterium avium* complex. J. Clin. Microbiol. **25**: 1239-1243.
- Romaniuk, P. J. and T. J. Trust. 1987. Identification of *Campylobacter* species by Southern hybridization of genomic DNA using an oligonucleotide probe for 16 S rRNA genes. FEMS Microbiol. Lett. **43**: 331-335.
- Ross. A. J., J. E. Martin, and V. Bress. 1968. *Vibrio anguillarum* from an epizootic in rainbow trout (*Salmon gairdneri*) in the U.S.A. Bull. Off. Int. Epiz. **69**: 1139-1148.
- Rucker, R. R., B. J. Earp, and E. J. Ordal. 1953. Infectious disease of Pacific salmon. Trans. Amer. Fish Sco. 83: 297-312.
- Salyers, A. A., S. P. Lynn, and J. F. Gardner. 1983. Use of randomly cloned DNA fragments for identification of *Bacteriodes thetaiotaomicron*. J. Bacteriol. **154**: 287-293.
- Sambrook, J., E. F. Fritsch, and T. Maniatis. 1989. Molecular cloning: a laboratory manual. 2nd ed. Cold Spring Harbor Laboratory, Cold Spring Harbor, New York.
- Sindermann, C. J. 1970. Principal disease of marine fish and shellfish. Academic Press, New York, London.
- Smith, I. W. 1961. A disease of finnock due to *Vibrio anguillarum*. J. Gen. Microbiol. **24**: 247-253.
- Totten, P. A., C. L. Fennell, F. C. Tenover, J. M. Wezenberg, P. L. Perine, W. E. Stamm, and K. K. Holmes. 1985. *Campylobacter cinaedi* (sp. nov.), *Campylobacter fennelliae* (sp. nov.): Two new *Campylbacter* species associated with enteric disease in homosexual men. J. Infect. Dis. **151**: 131-139.

- Tompkins, L. S., N. Troup, A. Labigne-Roussel, and M. L. Cohen. 1986. Cloned random chromosomal sequences as probes to identify *Salmonella* species. J. Infect. Dis. **154**: 156-162.
- Tung, M.-C., S.-S. Tsai, and S.-C. Chen. 1985. Study on *Vibrio anguillarum* infection in cultured milkfish (*Chanos chanos*) in Taiwan. COA Fisheries Series No. 10, Fish Disease Research (VII): 27-37.
- van Eys, G. J. J. M., J. Zaal, G. J. Schoone, and W. J. Terpstra. 1988. DNA hybridization with hardjobovis-specific recombinant probe as a method for type discrimination of *Leptospira interrogans* serovar *hardlo*. J. Gen. Microbiol. **134**: 567-574.

應用隨機基因探針偵測 Vibrio anguillarum 和 Vibrio alginolyticus

謝玉玲(1)、蔡珊珊(1,2)

(收稿日期:1999年7月30日;接受日期:1999年9月8日)

摘 要

以 Vibrio anguillarum 和 Vibrio alginolyticus 作為測試菌株。利用限制酶 HindIII 和 EcoRI 將細菌的基因體 DNA 切割後,選出介於 0.5-2.0 kb 之片段選殖到質體 pUC19上。利用非放射性探針標定篩選的片段,由 102 選殖的片段中找到一段只能與 V. alginolyticus 雜交;由 94 選殖的片段中找到三段只能與 V. anguillarum 雜交的片段,並找到一段只能與 V. anguillarum 血清型 C作用的片段。

關鍵詞: Vibrio anguillarum, Vibrio alginolyticus, 菌落雜交, 隨機選殖 DNA 片段, 血清型。

^{1.} 國立台灣大學植勿學系,台北市 106,台灣,中華民國。

^{2.} 通訊連絡員。E-mail: sstsay@ccms.ntu.edu.tw