# STEROIDS AND TRITERPENOIDS FROM ROSA LAEVIGATA

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Key Word Index—Rosa laevigata; Rosaceae; aerial parts; ursolic acid derivatives; euscaphic acid derivatives; oleanolic acid derivatives; sitosteryl glucosides; stigmastadiol glucoside.

Abstract—An acetone extract of aerial parts of Rosa laevigata was found to contain 16 components, including derivatives of ursolic, euscaphic and oleanolic acids as well as glucosides of sterols. Among them,  $2\alpha$ -methoxyursolic acid,  $11\alpha$ -hydroxytormentic acid, tormentic acid 6-methoxy- $\beta$ -glucopyranosyl ester and stigmasta- $3\alpha$ ,  $5\alpha$ -diol 3-O- $\beta$ -D-glucopyranoside are new compounds. Their structures were established by chemical and spectral methods.

## INTRODUCTION

Rosa laevigata is a shrub, very common in thickets at low altitudes, from China, Japan and Taiwan [1]. This plant has been used as a traditional folk medicine [2], an extract of the root of R. multiflora has been shown to have hypolipidimic activity [3]. The chemical constituents of R. laevigata have not been investigated, but a survey of the literature shows that plants of the same genus are rich in triterpenoid acids, sitosterol and their glycosides [3–5]. We describe here constituents isolated from R. laevigata for the first time.

## **RESULTS AND DISCUSSION**

An acetone extract of air-dried aerial parts of R. laevigata was concentrated to give semi-solids. Chromatography of the semi-solids and combination of appropriate fractions (monitored by TLC analyses) led to four fractions A-D. Fraction A was found to contain ursolic acid (1) [6], oleanolic acid (10) [7] and hederagenin (11) [7]. Fractions B and C also consisted of related triterpenoid acids, but no methyl esters by analyses of <sup>1</sup>H NMR spectra, which showed no signal between  $\delta 3.5$ and 3.8 attributable to CO<sub>2</sub>Me. Fractions B and C were separately treated with diazomethane and subjected to chromatography to give methyl ursolate (1a) [8, 9], methyl  $2\alpha$ -hydroxyursolate (2a) [8–10], methyl  $2\alpha$ -methoxyursolate (3a), methyl tormentate (4a) [10], methyl 11α-hydroxytormentate (5a), methyl cuscaphate (8a) [11, 12], methyl oleanolate (10a) [9], and the methyl ester of hederagenin (11a) [9]. Fraction D was triturated with ethyl ether and the soluble components were composed of tormentic acid  $\beta$ -D-glucopyranosyl ester (6) [3], tormentic acid 6-methoxy- $\beta$ -D-glucopyranosyl ester (7), euscaphic acid  $\beta$ -D-glucopyranosyl ester (9) [12] and methyl  $\beta$ -D-glucopyranoside (16) [13]. The insoluble residue of fraction D, which showed no proton resonances for acetates MeCO<sub>2</sub>R at  $ca \delta 2.0$ , was subjected to peracetylation and separated by HPLC to give sitosteryl- $\beta$ -Dglucopyranoside tetraacetate (12Ac) [14, 15], 7-oxositosteryl-β-D-glucopyranoside tetraacetate (13Ac) [15], 7hydroxysitosteryl-3- $O-\beta$ -D-glucopyranoside tetraacetate

(14Ac) [15] and stigmasta- $3\alpha$ ,  $5\alpha$ -diol 3-O- $\beta$ -D-glucopyranoside tetraacetate (15Ac).

Identification of the known compounds was based on comparison of the physical and spectral properties (mp,  $[\alpha]$ , mass spectrum, IR, <sup>1</sup>H and <sup>13</sup>C NMR) with those reported in literature. Compounds 3, 5, 7 and 15 are new



compounds. The methyl ester  $3a (C_{32}H_{52}O_4)$  showed a parent peak at m/z 500 and a base peak at m/z 262 attributable to the fragment derived from retro-Diels-Alder reaction [16]. The H-3 resonance at  $\delta 3.02$  exhibited as a doublet with a large coupling constant 9.4 Hz, which indicated the trans-diaxial relationship between H-2 and H-3. Compound 3a was further proved to be a methylation product of 2a (2 equiv NaH, 1 equiv MeI). Complete assignment of the proton and carbon resonances for the methyl ester 5a (Tables 1 and 2) was obtained with the assistance of the  ${}^{1}H{-}^{1}H$  and  ${}^{1}H{-}^{13}C$ correlation spectra. The 11-hydroxy group was inferred to be on the  $\alpha$ -face (equatorial position) because the axial H-11 at  $\delta$  4.84 had a large coupling constant (8.6 Hz) with H-9. The allylic alcohol 5a  $(C_{31}H_{50}O_6)$  was prone to autoxidation on standing to give the corresponding enone  $(C_{31}H_{48}O_6)$  which showed the exact mass for  $[M]^+$  at m/z 516.3427 (calcd 516.3452).

In addition to the 30 carbons for triterpenes of the ursolic acid type, compound 6 displayed another six signals at  $\delta 62.2, 71.1, 73.9, 78.8, 79.2$  and 95.7 attributable to the carbons of glucopyranose. The FAB mass spectrum of 6 showed negative ions at m/z 649 for  $[M-1]^-$  and at m/z 487 owing to cleavage at the glycoside bond. Com-

pound 6, namely rosamultin, has been reported once as a constituent of roots of *R. multiflora* [3]. The FAB mass spectrum of 7 showed a  $[M-1]^-$  ion at m/z 663 and an intense peak at m/z 487 for the aglycone. The structure of 7 was assigned by comparison of the <sup>13</sup>C NMR spectrum with that of 6 (Table 2). The values of <sup>13</sup>C resonance of the two compounds were similar except for 7, which showed an additional signal for the methoxy group at  $\delta$  57.0 and the C-6 resonance occurring at a lower field of  $\delta$  72.6. The proton of the anomeric carbon (C-1') appearing at  $\delta$  6.21 (d, J = 7.8 Hz) was in agreement with an axial  $\alpha$ -orientation.

Compounds 13 and 14 are the first report in nature although synthetic samples have already been used in pharmacological studies [15]. Compound 14Ac actually consisted of the  $7\alpha$  and  $7\beta$  epimers in nearly equal amounts as indicated by the H-7 signals at  $\delta 3.78$  (m) and 3.82 (m). Unlike the sitosterol derivatives 12Ac-14Ac, compound 15Ac has no C=C double bond by analyses of its <sup>1</sup>H and <sup>13</sup>C NMR spectra (Table 3). The parent peak at m/z 762 supported the assigned structure of 15Ac. The H-3 $\beta$  (equatorial) of 15Ac occurred as a broad singlet at  $\delta 3.91$ , whereas the axial H-3 $\alpha$  for 12Ac-14Ac showed as multiplets at higher fields ( $\delta 3.44$ -3.57). A resonance at

Table 1. Pertinent spectral data for compounds 3a, 5a, 7, 13Ac and 15Ac

	m/z (rel. int.)	$v_{\rm max}^{\rm KBr} {\rm cm}^{-1}$	δ <sub>H</sub> (200 MHz)
<b>3</b> a	70 eV: 500 [M] <sup>+</sup> (7), 440 (5), 262 (100), 237 (12), 203 (65), 189 (17), 187 (10), 133 (31). HRMS, C <sub>32</sub> H <sub>52</sub> O <sub>4</sub> requires 500.3867. Found: 500.3866.	3472, 2926, 1720.	CDC1 <sub>3</sub> : 0.72 (3H, s), 0.81 (3H, s), 0.82 (3H, d, $J = 6.0$ Hz), 0.91 (3H, d, $J = 6.0$ Hz), 0.95 (3H, s), 1.02 (3H, s), 1.05 (3H, s), 2.15 (1H, d, $J = 11.2$ Hz, H-18), 3.02 (1H, d, $J = 9.4$ Hz, H-3), 3.20 (1H, ddd, $J = 9.7$ , 9.4, 4.2 Hz, H-2), 3.35 (3H, s, OMe), 3.58 (3H, s, CO <sub>2</sub> Me), 5.23 (1H, t, $J = 3.6$ , H-12)
5a	20 eV: 518 [M] <sup>+</sup> (8), 501 (17), 500 (15), 441 (20), 295 (3), 277 (4), 251 (100), 217 (3), 205 (15), 187 (15).	3416, 2936, 1707, 1151, 1049.	Pyridine- $d_5$ : 0.93 (3H, s), 1.02 (3H, d, $J = 6.3$ Hz), 1.07 (3H, s), 1.15 (3H, s), 1.23 (3H, s), 1.58 (3H, s), 1.78 (3H, s), 2.25 (1H, d, $J = 8.6$ Hz, H-9), 2.94 (1H, s, H-18), 2.97 (1H, dd, H-1 $\beta$ ), 3.33 (1H, d, $J = 9.3$ Hz, H-3), 3.70 (3H, s, CO <sub>2</sub> Me), 4.08 (1H, m, H-2), 4.84 (1H, dd, $J = 8.6$ , 3.9 Hz, H-11), 6.18 (1H, d, $J = 3.9$ Hz, H-12).
7	FAB-NI: 663 [M – 1] <sup>~</sup> , 487 (aglycone), 469.	3404, 2922, 1724, 1449, 1364, 1166, 1072, 1029.	Pyridine- $d_5$ : 1.02 (3H, $d$ , $J = 6.0$ Hz, Me-20), 1.06 (3H, $s$ ), 1.08 (3H, $s$ ), 1.17 (3H, $s$ ), 1.24 (3H, $s$ ), 1.36 (3H, $s$ ), 1.64 (3H, $s$ ), 2.91 (1H, $s$ , H-18), 3.37 (1H, $d$ , $J = 9.0$ Hz, H-3), 3.38 (3H, $s$ , OMe), 3.95–4.28 (6 H, $m$ ), 5.51 (1H, $br$ $s$ , H-12), 6.21 (1H, $d$ , $J = 7.8$ Hz).
13Ac	20 eV: 758 [M] <sup>+</sup> (30), 426 (5), 412 (48), 410 (65), 394 (73), 331 (100), 271 (22), 168 (15).	2953, 1747, 1669, 1458, 1363, 1221, 1039, 906.	CDCl <sub>3</sub> : 0.64 (3H, s, Me-18), 0.77 (3H, $d$ , $J = 6.6$ Hz, Me-26)*, 0.79 (3H, $d$ , $J = 6.6$ Hz, Me-27)*, 0.80 (3H, $t$ , $J = 7.3$ Hz, Me-29), 0.88 (3H, $d$ , $J = 6.2$ Hz, Me-21), 1.14 (3H, s, Me-19), 1.97 (3H, s, MeCO <sub>2</sub> ), 1.99 (3H, s, MeCO <sub>2</sub> ), 2.02 (3H, s, MeCO <sub>2</sub> ), 2.04 (3H, s, MeCO <sub>2</sub> ), 3.57 (1H, $m$ , H-3), 3.65 (1H, $m$ , H-5'), 4.08 (1H, $dd$ , $J = 12.2$ , 2.3 Hz, H-6'), 4.21 (1H, $dd$ , $J = 12.2$ , 4.7 Hz, H-6'), 4.56 (1H, $d$ , $J = 7.9$ Hz, H-1'), 4.93 (1H, $dd$ , $J = 9.5$ Hz, H-2'), 5.04 (1H, $s$ , H-6).
15Ac	20 eV: 762 [ <b>M</b> ] <sup>+</sup> (1), 761 (1), 743 (2), 413 (38), 396 (31), 331 (56), 271 (19), 253 (3), 169 (100), 109 (17).	3439, 2953, 1748, 1364, 1226, 1038.	CDCl <sub>3</sub> : 0.65 (3H, s, Me-18), 0.78 (3H, d, $J = 6.7$ Hz, Me-26)*, 0.80 (3H, d, $J = 6.7$ Hz, Me-27)*, 0.81 (3H, t, $J = 7.3$ Hz, Me-29), 0.88 (3H, d, $J = 6.2$ Hz, Me-21), 1.22 (3H, s, Me-19), 1.98 (3H, s, MeCO <sub>2</sub> ), 2.00 (3H, s, MeCO <sub>2</sub> ), 2.02 (3H, s, MeCO <sub>2</sub> ), 2.04 (3H, s, MeCO <sub>2</sub> ), 3.67 (1H, m, H-5'), 3.91 (1H, br s, H-3), 4.10 (1H, dd, $J = 12.2$ , 2.3 Hz, H-6'), 4.23 (1H, dd, $J = 12.2$ , 4.9 Hz, H-6'), 4.57 (1H, d, $J = 7.9$ Hz, H-1'), 4.94 (1 H, dd, $J = 9.5$ , 8.0 Hz, H-2'), 5.05 (1H, t, $J = 9.5$ Hz, H-4'), 5.17 (1H, t, $J = 9.5$ Hz, H-3').

\*These assignments are made according to ref. [20] but are not conclusive.

c	3a	42	5a	6	7	8 <b>a</b>
1	42.0	47.7	48.9	47.8	47.8	41.4
2	78.7	68.4	68.8	68.6	68.6	77.2
3	81.5	83.2	83.7	83.8	83.8	78.6
4	39.0	39.1	40.0	38.4	38.5	39.9
5	55.0	55.0	56.0	55.9	55.9	53.0
6	18.1	18.3	19.2	18.9	19.0	17.9
7	32.8	32.4	33.8	33.3	33.5	32.3
8	39.5	39.7	41.4	39.8	39.8	41.0
9	47.5	47.7	47.4	47.8	47.8	47.7
10	38.0	37.9	39.5	38.4	38.5	37.3
11	23.4	23.5	80.9	24.0	24.1	23.5
12	125.2	128.5	129.7	128.3	128.2	128.8
13	138.3	138.0	144.3	139.2	139.3	138.0
14	41.2	41.0	44.0	40.6	40.6	41.0
15	27. <b>9</b>	25.8	26.0	26.5	26.7	28.0
16	24.1	25.2	25.7	26.0	26.1	25.3
17	48.0	46.3	48.3	48.6	48.6	47.8
18	52.8	53.0	54.0	54.4	54.4	51.5
19	38.8	72.8	72.8	72.6	72.6	72.9
20	38.7	41.0	42.4	42.1	42.1	46.7
21	30.6	28.0	29.1	29.3	29.1	25.3
22	36.6	37.2	38.3	37.6	37.8	37.3
23	28.7	28.5	29.5	29.3	29.3	28.5
24	17.0	16.7	18.9	17.4	17.5	21.8
25	16.9	16.4	18.1	16.9	17.0	16.1
26	17.0	16.7	19.0	17.6	17.7	17.9
27	23.6	24.3	23.6	24.5	24.6	24.6
28	178.0	178.2	178.6	177.0	177.0	178.4
29	17.0	28.0	27.1	27.0	27.0	27.2
30	21.2	15.9	16.8	16.7	16.6	16.5
CO <sub>2</sub> Mc	51.4	51.4	51.8			66.2

Table 2. <sup>13</sup>C NMR spectral data of compounds **3a-8a** (75 MHz,  $\delta$ )

Compounds 3a, 4a and 8a were measured in chloroform-d, compounds 5a, 6 and 7 in pyridine-d<sub>5</sub>. The OMe-2 group of compound 3a showed a signal at  $\delta$  56.3. Signals for carbons of the glucosyl moiety in compound 6 occurred at  $\delta$  62.2 (C-6'), 71.1 (C-4'), 73.9 (C-2'), 78.8 (C-3'), 79.2 (C-5') and 95.7 (C-1'), while those of 7 showed at  $\delta$  57.0 (6'-OMe), 71.7 (C-4'), 72.6 (C-6'), 74.0 (C-2'), 77.9 (C-3'), 78.9 (C-5'), 95.7 (C-1').

 $\delta$  84.3 (s) was attributable to the tertiary carbon (C-5) with a substituent of the hydroxyl group. Changing the solvent from chloroform-d to pyridine-d<sub>5</sub> did not cause any apparent shift for either H-3 $\beta$  or Me-10. This experiment indicated that the C-5 hydroxyl group was on the  $\alpha$ -face, being away from the H-3 $\beta$  and the Me-10 $\beta$  group [17]. The sterols 12Ac, 13Ac and 15Ac showed negative optical rotations. It is known that the sitosterols in most vascular plants possess the 24 $\alpha$ R-configuration [18]. Meticulous examination of the <sup>13</sup>C NMR spectra of 12Ac-15Ac also revealed that they all have the 24R-configuration [19, 20].

### **EXPERIMENTAL**

Plant material. Aerial parts of R. laevigata Michx (1.4 kg) were collected from the seaside area of Nanliaou. A specimen of this plant is deposited in the herbarium of our university. Aerial parts without fruits were extd with  $3 \times 81$  Me<sub>2</sub>CO. The Me<sub>2</sub>CO extract was passed through a short column of activated charcoal. The filtrate was concd and the insol. oily solids collected. The semi-

solids (35 g) were subjected to CC on silica gel (340 g) and elution with gradients of EtOAc and n-hexane. The appropriate frs (monitored by TLC analyses) were combined to give 4 frs A (3 g), B (1.5 g), C (3 g) and D (3.2 g). The components of A were sepd by flash CC to give ursolic acid (1) (2 g) and oleanolic acid (10) (0.3 g) and hederagenin (11) (0.1 g). As the <sup>1</sup>H NMR spectrum of the portion B showed no signal for Me esters, portion B was treated with Et2O-CH2N2 to convert the acids into their corresponding Me esters, which were then separated by flash CC to give Me esters 1a (0.5 g), 10a (0.1 g) and 11a (30 mg). C was similarly treated with CH2N2 and the derivatives sepd to give 2ahydroxyursolic acid Me ester 2a (150 mg), 2a-methoxyursolic acid Me ester (3a) (34 mg), tormentic acid Me ester (4a) (300 mg), 11a-hydroxy tormentic acid Me ester (5a) (20 mg) and euscaphic acid Me ester (8a) (134 mg). D was triturated with Et<sub>2</sub>O and the ppts filtered. The mother liquor contained the glucosides 6 (58 mg), 7 (12 mg), 9 (42 mg) and 16 (0.8 g). The ppts, which showed no signal for acetates, were dissolved in pyridine and treated with excess Ac<sub>2</sub>O to give the corresponding peracetates. After purification by HPLC on a LiChrosorb Si 60 column, the tetraacetates of sitosteryl glucosides 12Ac (350 mg), 13Ac

с	12Ac	13Ac	14Ac	15Ac
1	37.2	36.7	38.4	34.2
2	29.7	28.5	29.4 <b>*</b>	28.2
3	80.0	78.6	79.7/79.5	75.7
4	38.9	38.4	39.5	39.7
5	140.3	164.3	143.0/145.0	84.3
6	122.1	126.3	124.2/125.9	28.2
7	31.9	202.2	73.3/64.5	33.5ª
8	31.8	29.0	29.1 <sup>a</sup>	36.1
9	50.1	49.9	48.3	46.1
10	36.7	36.2	36.6	39.8
11	21.0	21.1	21.0	21.2
12	39.7	38.6	40.9	39.7
13	42.3	42.8	42.2	42.9
14	56.7	55.9	55.9	56.1
15	24.3	25.9	24.3	24.1
16	28.2	28.5	28.5	29.1
17	56.0	54.6	55.4	55.7
18	11.8	11.8	11.8	12.0
19	19.3	19.0	19.1	19.0
20	36.1	36.0	36.1	36.1
21	18.8	17.2	18.8	18.2
22	33.9	33.9	34.0	33.9*
23	26.0	26.2	26.1	26.1
24	45.8	45.8	45.8	45.8
25	29.5	29.1	29.2*	30.3
26	19.0**	18.9**	19.0**	18.7 <sup>a</sup> *
27	19.8**	19.7**	19.8**	19.8**
28	23.0	23.0	23.1	23.0
29	11.9	11.9	12.0	12.1
1'	99.6	99.8	99.7	100.4
2'	71.7ª	71.8ª	71.7ª	71.7ª
3'	71.5 <sup>*</sup>	71.7ª	71.5ª	71.6ª
4'	68.5	68.4	68.5	68.5
5'	72.9	72.7	72.9	72.9
6'	62.1	61.9	62.1	62.1
MeCO,	20.6, 20.7, 169.2,	20.5, 20.6, 169.2,	20.6, 20.7, 169.2,	20.6, 20.7, 169.7,
2	169.3, 170.3, 170.5	169.3, 170.2, 170.6	169.3, 169.4, 170.3	169.8, 170.4, 170.7

Table 3. <sup>13</sup>C NMR spectral data of compounds 12Ac-15Ac (75 MHz, CDCl<sub>3</sub>,  $\delta$ )

<sup>a</sup>These assignments may be interchangeable.

\*These assignments are made according to ref. [20] but are not conclusive.

(51 mg), 14Ac (20 mg) and a stigmastadiol derivative 15Ac (40 mg) were obtained. All of these compounds are obtained as crystals from MeOH. Compound 1. Mp 290–292°,  $[\alpha]_{P}^{25} + 70.5^{\circ}$ (MeOH; c 1.4); 1a, mp 110-111°. Compound 2a, mp 202-204°,  $[\alpha]_{D}^{25} + 39.8^{\circ}$  (CHCl<sub>3</sub>; c 8.6). Compound **3a**, mp 84–86°,  $[\alpha]_{D}^{25}$ +16.5 (CHCl<sub>3</sub>; c2.25). Compound 4a, mp 146-148°,  $[\alpha]_{\rm P}^{25}$  $+30.2^{\circ}$  (CHCl<sub>3</sub>; c 1.4). Compound **5a**, mp 141–143°, [ $\alpha$ ] -11.6° (CHCl<sub>3</sub>; c1.35). Compound 6, mp 207-209°,  $[\alpha]_D^{25} + 15^\circ$ (MeOH; c 1.1). Compound 7, mp 195–198°,  $[\alpha]_{p}^{25} + 1.5^{\circ}$  (MeOH; c 0.2). Compound 8a, mp 122–124°,  $[\alpha]_D^{25}$  + 23.8° (CHCl<sub>3</sub>; c 8.9). Compound 9, mp 203–205°,  $[\alpha]_D^{25}$  +4.2° (MeOH; c0.3). Compound 10, mp 302-304°,  $[\alpha]_{D}^{25}$  + 79.2° (CHCl<sub>3</sub>; c 1.4). Compound 11, mp 328–331°,  $[\alpha]_{\rm B}^{25}$  + 75.4° (MeOH; c 1.6); 11a, mp 218–220°. Compound 12Ac, mp 165–167°,  $[\alpha]_{\rm B}^{25}$  – 29.1° (CHCl<sub>3</sub>; c 2.1). Compound 13Ac, mp 114–116°,  $[\alpha]_{\rm B}^{25}$  – 34.9° (CHCl<sub>3</sub>; c 3.37). Compound 14Ac (two epimers), mp 98-100°. Compound 15Ac, mp 99-101°,  $[\alpha]_D^{25}$ -14.0° (CHCl<sub>3</sub>; c0.6). Compound 16, mp 106–109°,  $[\alpha]_D^{25} - 30^\circ$  (Me<sub>2</sub>CO; c 1.2).

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