



## TETRANORTRITERPENOID INSECT ANTIFEEDANTS FROM *SEVERINIA BUXIFOLIA*

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**Key Word Index**—*Severinia buxifolia*; Rutaceae; root bark; antifeeding effect; limonoids; severinolide; cycloseverinolide.

**Abstract**—Two new tetranortriterpenoids, severinolide and cycloseverinolide, together with four known compounds, were isolated and characterized from the root bark of *Severinia buxifolia*. Severinolide, atalantin and cycloepitalantin showed significant antifeeding effects against *Plutella xylostalla*. © 1997 Elsevier Science Ltd. All rights reserved

### INTRODUCTION

*Severinia buxifolia* (*Atalantia buxifolia*) is a Chinese folk medicine and has been used for treatment of chronic rheumatism, paralysis, snake-bite and malaria [1]. Essential oils, coumarins, acridone alkaloids, sesquiterpenoids and triterpenoids have been isolated from this plant [2–9]. The leaves of this plant show resistance to phytophagous insects and the ethanol extract of the root bark of *S. buxifolia* was found to show significant antifeedant activity. This led us to reinvestigate its constituents. Bioassay-directed fractionation of the plant extract led to isolation and characterization of severinolide (**1a**), atalantin (**3**) and cycloepitalantin (**6**) as the antifeedant principles of the chloroform soluble fraction. We now describe the structural elucidation of two new limonoids, severinolide (**1a**) and cycloseverinolide (**2a**) together with four known compounds (**3–6**) which were isolated from the root bark of *S. buxifolia* and their antifeeding activity.

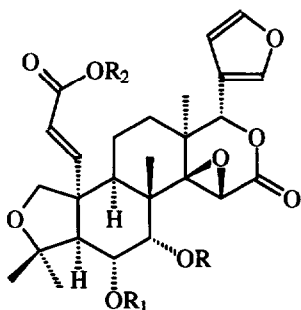
### RESULTS AND DISCUSSION

All of the limonoids except atalantolide (**5**) isolated in this study showed four C-Me resonances in their <sup>1</sup>H NMR spectra instead of five C-Me expected for an obacunone system. This fact suggested that these natural products belong to the limonin series and that C-19 has been oxidized.

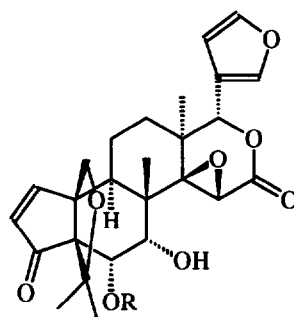
Severinolide (**1a**) was isolated as optically active

colourless plates and its elemental analysis indicated a molecular formula C<sub>31</sub>H<sub>38</sub>O<sub>11</sub>. The signals of **1a** at δ 165.4 (s), 167.3 (s), 170.3 (s) and 170.8 (s) in the <sup>13</sup>C NMR spectrum, together with the IR bands at 1750, 1735 and 1710 cm<sup>-1</sup>, revealed the presence of an α,β-unsaturated ester, δ-lactone and ester groups in the molecule. The <sup>1</sup>H NMR spectrum of **1a** showed typical signals [10, 11] for a β-substituted furan ring: H-17, H-15 an α,β-unsaturated ester system and an AB quartet for H-19 of a limonoid system as well as four C-Me resonances (Table 1). Two singlet acetyl signals appeared at δ 2.27 and 1.98 (each 3H). Severinolide (**1a**) was hydrolysed with sodium hydroxide and then acidified by hydrochloric acid to give **1b**. Methylation of **1b** with diazomethane afforded the diol derivative (**1c**). Oxidation of **1c** with Jones' reagent resulted in the formation of yellow crystals, which were identified as dehydroatalantin (**4**) by comparison of their spectral data and mixed melting points with an authentic specimen [12]. This result suggested that the two acetyl groups were located at C-6 and C-7. This arrangement was also supported by the fact that hydrolysis of **1a** to give diol (**1c**) results in a downfield shift of the H-15 resonance and an upfield shift of the H-5, H-6, H-7 and H-9 signals (Table 1). An AMX system [δ 2.90 (1H, *d*, *J* = 11 Hz), 5.11 (1H, *dd*, *J* = 3, 11 Hz) and 4.89 (1H, *d*, *J* = 3 Hz)] was attributed to H-5α (axial), H-6β (axial) and H-7β (equatorial). Acetylation of **1c** gave **1a** (21%) and **1d** (72%), consistent with the α (axial)-configuration of the 7-hydroxyl group. The complete structure and relative stereochemistry of **1a** was determined by single crystal X-ray analysis (Fig. 1). Thus severinolide has structure **1a**.

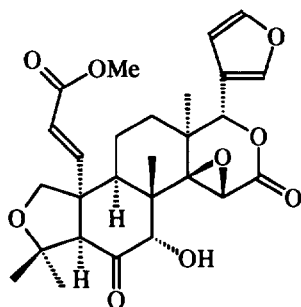
\* Author to whom correspondence should be addressed.



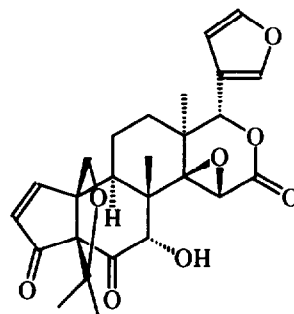
**1a** R=R<sub>1</sub>=Ac, R<sub>2</sub>=Me  
**1b** R=R<sub>1</sub>=R<sub>2</sub>=H  
**1c** R=R<sub>1</sub>=H, R<sub>2</sub>=Me  
**1d** R=H, R<sub>1</sub>=Ac, R<sub>2</sub>=Me



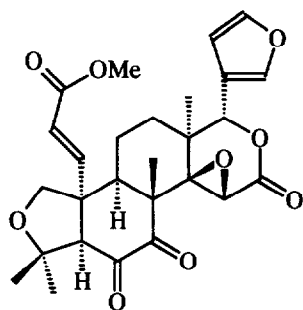
**2a** R=H  
**2b** R=Ac



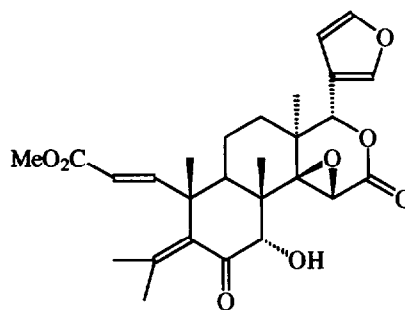
3



6



4



5

Cycloseverinolide (**2a**) was obtained as optically active colourless plates with the molecular formula C<sub>26</sub>H<sub>30</sub>O<sub>8</sub>. Its UV spectrum exhibited maxima at 217 and 319 nm characteristic of the presence of a cyclopentenone system [13]. The IR spectrum of **2a** showed absorption bands at 3530, 3300, 1725, 1662, 1030 and 895 cm<sup>-1</sup> indicating the presence of two hydroxyl groups, a  $\delta$ -lactone, an  $\alpha,\beta$ -unsaturated carbonyl system and a  $\beta$ -substituted furan ring in the molecule.

The <sup>1</sup>H NMR spectrum (Table 1) revealed typical H-15 and H-17 signals in a  $\beta$ -substituted furan ring, ring D epoxylactone, an  $\alpha,\beta$ -unsaturated ketone and two hydroxyl protons (exchanged with D<sub>2</sub>O). The appearance of four tertiary methyl signals and an AB quartet at  $\delta$  3.79 and 3.96 (each 1H, *d*, *J* = 9.5 Hz) suggested a carbon skeleton related to that of limonin with an ether bridge from C-19 to C-4. The above data suggested that cycloseverinolide (**2a**) was similar to

Table 1. <sup>1</sup>H NMR spectra of *Severinia* tetranortriterpenoids

	1a	1b*	1c	1d	2a	2a†	2b
H-1	6.15(1H, d, 13)	6.53(1H, d, 13)	6.52(1H, d, 13)	6.47(1H, d, 13)	7.69(1H, d, 5, 6)	7.76(1H, d, 6)	7.58(1H, d, 6)
H-2	5.85(1H, d, 13)	5.91(1H, d, 13)	5.88(1H, d, 13)	5.87(1H, d, 13)	6.14(1H, d, 5, 6)	6.06(1H, d, 6)	6.07(1H, d, 6)
H-5	2.90(1H, d, 11)	2.35(1H, d, 10)	2.24(1H, d, 10)	2.74(1H, d, 11)			
H-6	5.11(1H, dd, 3, 11)	3.82(1H, m)	3.84(1H, m)	5.08(1H, dd, 3, 11)	4.29(1H, dd, 3, 8)	4.16(1H, dd, 3, 10)	5.51(1H, d, 3)
6-OH			2.55(1H, d, 11)	2.09(3H, s)	6.29(1H, d, 8)	5.69(1H, d, 10)	
6-OAc	1.98(3H, s)			3.52(1H, dd, 3, 12)			2.18(3H, s)
H-7	4.98(1H, d, 3)	3.22(1H, m)	3.30(1H, dd, 3, 13)	4.74(1H, d, 12)	3.42(1H, t, 3)	3.28(1H, dd, 3, 4)	3.46(1H, dd, 2, 3)
7-OH			4.84(1H, d, 13)		2.50(1H, d, 3)	5.45(1H, d, 4)	2.31(1H, d, 2)
7-OAc	2.27(3H, s)						
H-9	3.21(1H, dd, 7, 13)	3.00(1H, m)	3.06(1H, dd, 6, 12)	3.05(1H, dd, 5, 12)	2.64(1H, dd, 7, 12)	2.56(1H, m)	2.78(1H, m)
H-15	3.62(1H, s)	3.95(1H, s)	4.09(1H, s)	3.98(1H, s)	3.83(1H, s)	3.86(1H, s)	3.78(1H, s)
H-17	5.57(1H, s)	5.60(1H, s)	5.62(1H, s)	5.57(1H, s)	5.58(1H, s)	5.55(1H, s)	5.58(1H, s)
H-19	3.90(1H, d, 10)	3.76(1H, d, 10)	3.71(1H, d, 9)	3.72(1H, d, 10)	3.79(1H, d, 9, 5)	3.76(1H, d, 10)	3.75(1H, d, 10)
	4.02(1H, d, 10)	3.96(1H, d, 10)	3.98(1H, d, 9)	4.06(1H, d, 10)	3.96(1H, d, 9, 5)	3.91(1H, d, 10)	4.03(1H, d, 10)
H-21,23	7.40(2H, d, 1)	7.52(2H, d, 2)	7.40(2H, d, 2)	7.35(2H, d, 2)	7.38(2H, d, 1, 5)	7.45(2H, d, 1)	7.41(2H, d, 1)
H-22	6.30(1H, d, 1)	6.43(1H, d, 2)	6.32(1H, d, 2)	6.27(1H, d, 2)	6.31(1H, d, 1, 5)	6.36(1H, d, 1)	6.33(1H, d, 1)
18-Me	1.12(3H, s)	1.13(6H, s)	1.08(3H, s)	1.08(3H, s)	1.11(3H, s)	1.10(3H, s)	1.12(6H, s)
28-Me	1.14(3H, s)	1.21(3H, s)	1.13(3H, s)	1.21(6H, s)	1.13(3H, s)	1.12(3H, s)	1.20(3H, s)
29-Me	1.25(3H, s)	1.38(3H, s)	1.25(3H, s)	1.24(3H, s)	1.19(3H, s)	1.15(3H, s)	1.27(3H, s)
30-Me	1.24(3H, s)		1.44(3H, s)		1.51(3H, s)		
CO <sub>2</sub> Me	3.67(3H, s)		3.77(3H, s)	3.75(3H, s)		1.47(3H, s)	

\* Run in (CD<sub>3</sub>)<sub>2</sub>CO.† Run in CDCl<sub>3</sub> + 10% DMSO-*d*<sub>6</sub>.

Figures in parentheses are coupling constants in Hz.

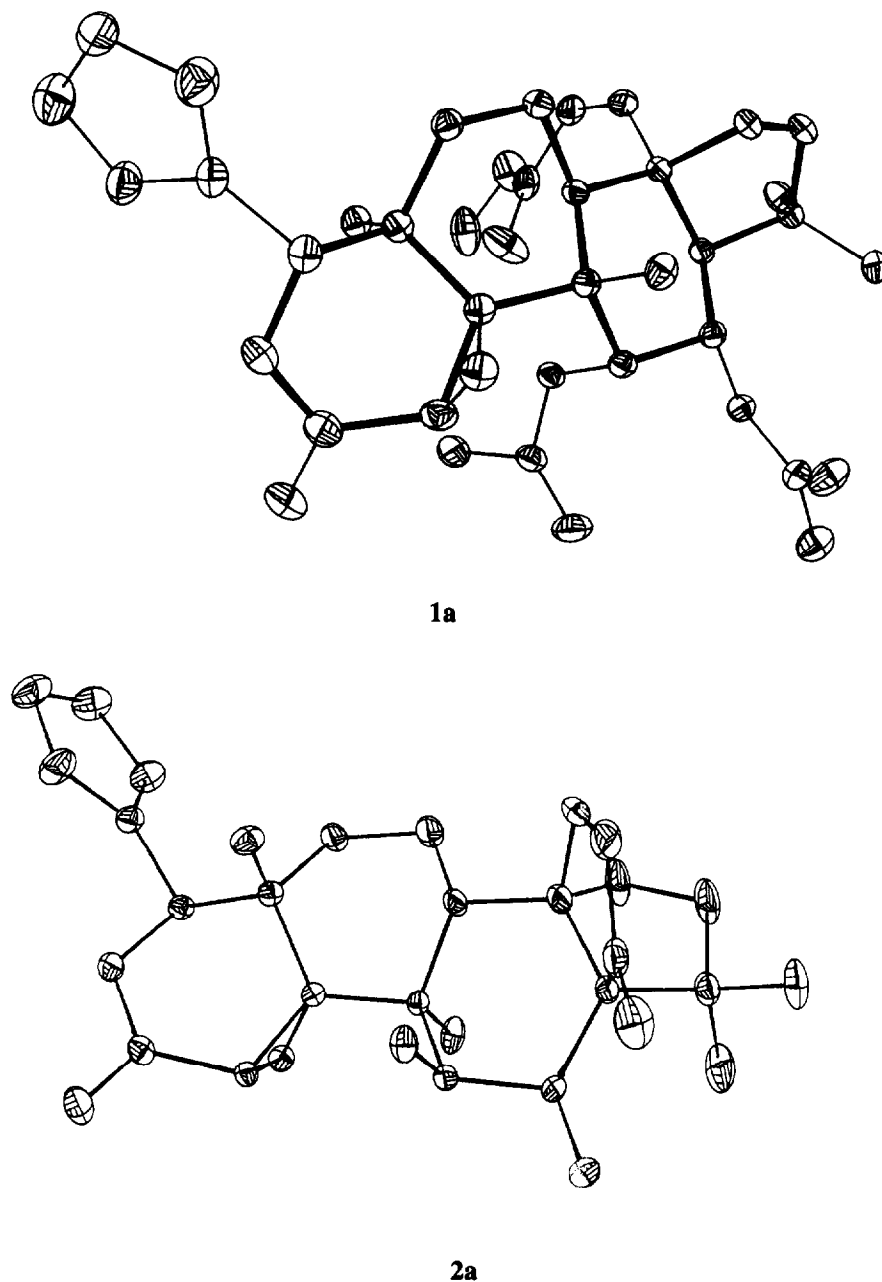


Fig. 1. Structure and solid-state conformation of compound **1a** and **2a**.

cycloepialantins (**6**) [14]. Oxidation of **2a** with Jones' reagent afforded colourless needles which had identical spectral data and TLC behaviour with an authentic sample of **6**. This indicated that the two hydroxyl groups were located at C-6 $\alpha$  and C-7 $\alpha$  [ $J_{6,7} = 3$  Hz]. The complete structure and relative stereochemistry of **2a** was determined by a single crystal X-ray analysis (Fig. 1). Thus cycloseverinolide has structure **2a**.

The known compounds, atalantins (**3**) [15], dehydro-atalantins (**4**) [12], atalantolide (**5**) [16] and cycloepialantins (**6**) [14] were also isolated and characterized by comparison of their spectroscopic data (UV, IR, NMR and mass spectrometry) with literature values.

The isolated compounds were subjected to anti-feedant activity evaluation [17, 18]. Severinolide (**1a**), atalantins (**3**) and cycloepialantins (**6**) showed strong anti-feedant activity against third instar larvae of the Diamondback moth (*Plutella xylostella*) with ED<sub>50</sub> at concentrations of 0.0625, 0.0625 and 0.25%, respectively (Table 2).

#### EXPERIMENTAL

Mps: uncorr. <sup>1</sup>HNMR (100, 200, 400 MHz) were recorded in CDCl<sub>3</sub>, except where noted. Chemical shift values are shown in ppm ( $\delta$ ) with TMS as an internal standard. MS were recorded using a direct inlet

Table 2. Antifeeding activity of tetranortriterpenoids from *Severinia buxifolia*

Compd	Method concn. (%)	II Time (day)								
		I Time (day)			1		2		5	
		1	2	5	C	T	C	T	C	T
<b>1a</b>	0.5	±	±	±	++++	±	++++	±	++++	+
	0.25	±	±	±	++	±	++++	±	++++	+
	0.125	±	±	±	+	±	++++	±	++++	±
	0.0625	+	+	++	+++	±	++++	++	++++	+++
	0.03125	+	+++	+++	+++	+	++++	+++	++++	++++
<b>2a</b>	0.5	++	+++	++++	+	++	++++	++++	++++	++++
	0.25	+	+++	+++	+	++	++++	++++	++++	++++
<b>3</b>	0.5	±	±	±	+++	±	++++	+	++++	++
	0.25	±	±	±	++	±	++++	+	++++	++
	0.125	±	±	+	++	±	++++	++	++++	+++
	0.0625	±	++	+++	+++	±	++++	++	++++	++++
	0.03125	+	+++	+++	+++	±	++++	+++	++++	++++
<b>4</b>	0.5	±	±	+	+++	±	++++	++	++++	+++
	0.25	++	++++	++++	+++	+	++++	++++	++++	++++
<b>5</b>	0.5	++	++++	++++	+++	+	++++	++++	++++	++++
	0.25	++	++++	++++	++++	+	++++	++++	++++	++++
<b>6</b>	0.5	±	±	±	+++	+	++++	+++	++++	+++
	0.25	±	±	±	++++	±	++++	++	++++	++
Galecron	0.5	±	+	+	+++	±	++++	±	++++	±
	0.25	±	+	++	++	±	++++	+++	++++	++
	0.125	±	++	+++	++	±	++++	++	++++	++
	0.0625	±	++	+++	++	+	++++	+++	++++	+++
	0.03125	+	+++	+++	+	±	++++	++	++++	+++
Control	—	++	++++	++++	++	/	++++	/	++++	/

C: no treatment with compound; T: treatment with compound.

Observation: Consumptions of the cabbage leaf disks were evaluated by the index of 6 grades as shown below at 1, 2 and 5 days after treatment.

Grades of leaf disk consumption: —: 0%; ±: 1–20%; +: 21–40%; ++: 41–60%; +++: 61–80%; ++++: 81–100%.

system. UV were determined in MeOH and IR were recorded in KBr disc.

**Plant material.** *Severinia buxifolia* (Pior.) Tenore was collected from Tainan, Taiwan and identified by Prof. C. S. Kuoh. A voucher specimen is deposited in the Herbarium of the National Cheng Kung University, Tainan, Taiwan, R.O.C.

**Extraction and separation.** The procedure of extraction and sepn was as related reference [6]. The benzene eluted fr. was rechromatographed on silica gel and eluted with *i*-Pr<sub>2</sub>O and Et<sub>2</sub>O to afford **1a** (5.1 g), **3** (0.61 g), **5** (0.37 g), **4** (0.03 g) and a mixt. This mixt. was chromatographed on silica gel using EtOAc–benzene (1:4) as eluent and 20 ml frs were collected and monitored by TLC. The frs giving an identical spot were combined together. Compound **6** (0.62 g) and **2a** (0.57 g) were obtained, respectively.

**Severinolide (1a).** Colourless plated (Me<sub>2</sub>CO), mp 219–221°. [ $\alpha$ ]<sub>D</sub> +53.08° (c 1.3, CHCl<sub>3</sub>). Anal. calcd for C<sub>31</sub>H<sub>38</sub>O<sub>11</sub>: found: C, 63.42; H, 6.55%, required: C, 63.48; H, 6.48%. UV  $\lambda_{\max}$  nm: 214. IR  $\nu_{\max}$  cm<sup>-1</sup>: 1750, 1735, 1710, 1640. EIMS *m/z* (rel. int.): 586[M]<sup>+</sup>, 511, 463, 451, 432, 403, 393, 361, 343, 311, 303, 285, 253, 225, 95(100), 43. <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  170.8(s), 170.3(s), 167.3(s), 165.4(s), 157.3(d), 143.0(d), 141.2(d), 120.4(s), 120.2(d), 109.8(d), 82.9(s), 78.0(d), 73.2(t), 70.6(d), 69.8(s), 69.2(d), 56.7(d), 54.6(s),

51.3(q), 51.1(d), 42.3(s), 38.8(s), 35.4(d), 30.3(q), 25.5(t), 23.9(q), 21.0(q), 20.7(q), 18.0(t), 18.0(q), 17.1(q).

**Hydrolysis of 1a.** Compound **1a** (0.5 g) was dissolved in MeOH (50 ml) containing NaOH (2.5 g) and stirred at room temp. for 7 hr. The reaction product was treated in the usual way to yield colourless needles of **1b** (310 mg) (Me<sub>2</sub>CO), C<sub>26</sub>H<sub>32</sub>O<sub>9</sub>, mp 245–247°. IR  $\nu_{\max}$  cm<sup>-1</sup>: 3340, 1732, 1705, 1625. EIMS *m/z*: 448[M]<sup>+</sup>, 452, 444, 426, 422(100%), 408, 394, 380, 366, 210, 95.

**Methylation of 1b.** Treatment of **1b** (300 mg) with excess CH<sub>2</sub>N<sub>2</sub> in the usual way afforded **1c** (290 mg) as colourless syrup, C<sub>27</sub>H<sub>34</sub>O<sub>9</sub>. IR  $\nu_{\max}$  cm<sup>-1</sup>: 3430, 1740, 1700, 1625. EIMS *m/z*: 469([M]<sup>+</sup>–43), 368, 361(100), 256, 249, 236, 223, 213, 185, 171, 129, 121, 111, 97, 95.

**Oxidation of 1c.** Jones' reagent was added dropwise to stirring soln of **1c** (0.07 g) in Me<sub>2</sub>CO. After standing for 20 min at room temp., excess of reagent was destroyed by using two drops of MeOH. Standard workup afforded yellowish crystals (0.053 g) (Me<sub>2</sub>CO), mp 204°, which were identified as dehydroatantian (**4**) by comparison of their spectral data, TLC and mixed mp with authentic sample [12].

**Acetylation of 1c.** Compound **1c** (0.15 g) was dissolved in C<sub>3</sub>H<sub>5</sub>N (2 ml) and Ac<sub>2</sub>O (3 ml) and the mixt.

allowed to stand overnight at room temp. Standard workup gave a residue which showed two spots on TLC (benzene–Me<sub>2</sub>CO, 4:1). The mixt. was sep'd by prep. TLC using the same solvent system as TLC. The front spot (32 mg) was identified with **1a** by direct comparison. The second spot, compound **1d** (107 mg), was recrystallized from Me<sub>2</sub>CO as colourless needles, C<sub>29</sub>H<sub>36</sub>O<sub>10</sub>, mp 238–240°. UV  $\lambda_{\max}$  nm: 215. IR  $\nu_{\max}$  cm<sup>-1</sup>: 3450, 1740, 1720, 1695, 1620. EIMS  $m/z$ : 544[M]<sup>+</sup>, 513, 511, 469, 451, 421, 403(100%), 361, 345, 343, 303, 285, 253, 225, 95.

*Crystal data of 1a.* M = 586, triclinic, space group P2  $a = 18.6659(73)$ ,  $b = 13.1231(35)$ ,  $c = 11.9066(28)$  Å,  $\alpha = \beta = \gamma = 92.909(3)^\circ$ ,  $U = 2912.84$  Å<sup>3</sup>,  $Z = 4$ ,  $D_c = 1.336$  mg m<sup>-3</sup>,  $\mu$  (MoK $\alpha$  radiation,  $\lambda = 0.70923$  Å) crystal dimensions:  $0.1 \times 0.2 \times 0.3$  mm. Intensity data ( $\pm h$ ,  $\pm k$ ,  $\pm l$ ,  $\theta_{\max} = 67^\circ$ ) were recorded on a Siemens R3m/V diffractometer. The crystal structure was solved by a direct method. Full-matrix least-squares refinement of atomic parameters (anisotropic C, O; isotropic H) converged at  $R = 5.10$  ( $R_w = 6.33$ ) over 5753 reflections with  $I > 3.0\sigma$  (1).

*Cycloseverinolide (2a).* Colourless plates (Me<sub>2</sub>CO), mp  $> 360^\circ$ .  $[\alpha]_D + 50.75^\circ$  ( $c$  0.4, CHCl<sub>3</sub>). Anal. calcd for C<sub>26</sub>H<sub>30</sub>O<sub>8</sub>: found: C, 66.29; H, 6.55%, required: C, 66.37; H, 6.43%. UV  $\lambda_{\max}$  nm: 217, 319. IR  $\nu_{\max}$  cm<sup>-1</sup>: 3530, 3300, 1725, 1662, 1590. EIMS  $m/z$ : 470[M]<sup>+</sup>, 455, 329(100%), 271, 123, 121, 107, 105, 95.

*Oxidation of 2a.* Compound **2a** (0.1 g) was oxidized as above to give a colourless needle (0.075 g)(Me<sub>2</sub>CO), mp 325–328°(dec.). Anal. calcd for C<sub>26</sub>H<sub>28</sub>O<sub>8</sub>: found: C, 66.29; H, 6.11%, required: C, 66.65; H, 6.02%. UV  $\lambda_{\max}$  nm: 215, 327. IR  $\nu_{\max}$  cm<sup>-1</sup>: 3480, 1725, 1715, 1690, 1590.  $[\alpha]_D - 77.02^\circ$  ( $c$  0.47, CHCl<sub>3</sub>). EIMS  $m/z$ : 468[M]<sup>+</sup>, 456, 422, 345, 327(100%), 287, 269, 121, 95, 91, identical with cyclopiatalantin (**6**) [13].

*Acetylation of 2a.* Compound **2a** (50 mg) was heated in Ac<sub>2</sub>O (10 ml) and pyridine (2 ml) at 105° for 7 hr. The usual workup gave rectangular prisms of **2b** (45 mg)(Me<sub>2</sub>CO), C<sub>28</sub>H<sub>32</sub>O<sub>9</sub>, mp 298–300°. UV  $\lambda_{\max}$  nm: 219, 326. IR  $\nu_{\max}$  cm<sup>-1</sup>: 3340, 1730, 1720, 1690, 1605. EIMS  $m/z$ : 512[M]<sup>+</sup>, 497, 389, 371, 329, 313, 271(100%), 95.

*Crystal data of 2a.* M = 470, triclinic space group P2,  $a = 11.3099(17)$ ,  $b = 11.7209(17)$ ,  $c = 16.9764(28)$  Å,  $\alpha = \beta = \gamma = 90(3)^\circ$ ,  $U = 2250.43$  Å<sup>3</sup>,  $Z = 4$ ,  $D_c = 1.39$  mg m<sup>-3</sup>,  $\mu$  (MoK $\alpha$  radiation,  $\lambda = 0.70923$  Å) crystal dimensions:  $0.1 \times 0.2 \times 0.3$  mm. Intensity data ( $\pm h$ ,  $\pm k$ ,  $\pm l$ ,  $\theta_{\max} = 70^\circ$ ) were recorded on a Siemens R3m/V diffractometer. The crystal structure was solved by a direct method. Full-matrix least-squares refinement of atomic parameters (anisotropic C, O; isotropic H) converged at  $R = 5.53$  ( $R_w = 4.07$ ) over 3490 reflections with  $I > 3.0\sigma$  (1).

*Antifeeding activity test.* The test insect was third

instar larvae of Diamondback moth (*Plutella xylostella*). (a) A filter paper moistened with 1 ml of H<sub>2</sub>O was placed in each Petri dish (2 cm(H)  $\times$  9 cm(D)). (b). Both sides of cabbage leaf disks (2 cm diameter) were treated with 20  $\mu$ l MeOH of chemicals and air dried, then these were placed on the filter paper. Method I: 4 treated disks were placed in a dish. Method II: 3 treated disks and 3 untreated disks were placed alternately. (c) In method I and II, two larvae per disk were released in a dish.

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#### REFERENCES

- Sasaki, S., *Khoyo Taiwan Minkan Yakuyo Shokubutsu Shi*. Khobunkan, Taipei, 1924, p. 36.
- Scora, R. W., *Phytochemistry*, 1966, **5**, 823.
- Tin-Wa, M., Scora, R. W. and Kumanoto, J., *Lloydia*, 1972, **35**, 183.
- Tin-Wa, M., Bonomo, S. and Scora, R. W., *Planta Medica*, 1979, **37**, 379.
- Dreyer, D. L., *Tetrahedron*, 1967, **23**, 4613.
- Wu, T. S., Kuoh, C. S. and Furukawa, H., *Phytochemistry*, 1982, **21**, 1771.
- Wu, T. S., Niwa, M. and Furukawa, H., *Phytochemistry*, 1984, **23**, 595.
- Gu, G. M., *Yaoxue Xuebao*, 1987, **22**, 886.
- Qin, D. K., *Yaoxue Xuebao*, 1986, **21**, 683.
- Dreyer, D. L., *Tetrahedron*, 1965, **21**, 75.
- Powell, J. W., *Journal of the Chemical Society*, 1969, **47**, 2849.
- Thakar, M. R. and Sabata, B. K., *Industrial Journal of Chemistry*, 1969, **7**, 870.
- Pasto, D. J. and Johnson, C. R., in *Organic Structure Determination*. Prentice-Hall, New Jersey, 1969, p. 96.
- Dreyer, D. L., Bennett, R. D. and Basa, S. C., *Tetrahedron*, 1976, **32**, 2376.
- Basu, D. and Basu, S. C., *Journal of Organic Chemistry*, 1972, **37**, 3035.
- Shringarpure, J. D. and Sabata, B. K., *Industrial Journal of Chemistry*, 1975, **13**, 24.
- Kubo, I. and Klocke, J. A., *Les Colloques de II N.R.A. No*, 1981, **7**, 117.
- Klocke, J. A. and Kubo, I., *Entomology, Experimental and Applied*, 1982, **32**, 299.