

Effect of gestational smoke exposure on atopic dermatitis in the offspring

Wang I-J, Hsieh W-S, Wu K-Y, Guo YL, Hwang Y-H, Jee S-H, Chen P-C. Effect of gestational smoke exposure on atopic dermatitis in the offspring.

Pediatr Allergy Immunol 2008; 19: 580–586.

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The adverse impact of smoking on respiratory diseases and birth outcomes in children is well-known. However, the influence of smoke exposure including environmental tobacco smoke (ETS) and maternal smoking during pregnancy on atopic dermatitis (AD) is not clear. The purpose of this study was to evaluate the effect of gestational smoke exposure on the development of AD in the offspring on the basis of the maternal and cord blood cotinine. We recruited 261 mother and newborn pairs in 2004. Cord blood and information on perinatal factors of children were gathered at birth. At 2 yr of age, information about development of AD and environmental exposures were collected. We compared AD with non-AD children for the concentration of cotinine in cord and maternal blood measured by high performance liquid chromatography–mass spectrometry. Multiple logistic regressions were performed to estimate the relationship of cotinine levels and AD. About 150 mother and child pairs completed the follow-up study and specimen collection with 38 (25.3%) children developing AD. Two (1.3%) out of 150 mothers smoked during pregnancy, while 38 (25.3%) mothers reported having ETS exposure. Cotinine levels in cord blood and maternal blood were highly correlated ($r = 0.71$, $p < 0.001$). The risk of AD was found to increase with maternal and cord blood cotinine levels in a dose–response manner (p for trend = 0.01). Children exposed to high levels (> 75th percentile) had a significantly increased risk of AD. Smoke exposure during pregnancy might increase the risk of AD in children. Avoidance of prenatal smoke exposure may be warranted for early prevention.

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Key words: atopic dermatitis; cotinine; environmental tobacco smoke

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Accepted 2 May 2008

Most studies evaluating the effect of smoking and environmental tobacco smoke (ETS) exposure on health outcomes rely on questionnaires (1, 2). Self-reported measures, such as number of cigarettes smoked per day in smokers or hours per day exposed to ETS by non-smokers, are highly imprecise because of individual differences in smoking habits and environmental characteristics (3, 4). ETS exposure might change with the number of cigarettes smoked, proximity of non-smokers to smokers, room ventilation and other environmental characteristics as well as individual differences concerning the adverse effects of

ETS. At present, cotinine measured in blood, saliva, or urine is considered to be the most specific and the most sensitive biomarker for smoke exposure (5). Cotinine, the major metabolite of nicotine, has a longer half life (18–20 h) than nicotine (1–2 h) (6). Cord serum cotinine concentration is related to daily smoking rate during pregnancy (7). It appears to be the most adequate biomarker of fetal exposure to smoking at the end of pregnancy, distinguishing not only active smoking from passive smoking but also exposure to ETS from non-exposure (8). To avoid reporting bias or misclassification of

exposure and non-exposure, we based the calculation of smoking associated health risks on the concentrations of the plasma cotinine as an objective biomarker.

In the Taiwan 2001 survey, 4.5% reproductive age women smoke while 57–59% of non-smoking women of childbearing ages were exposed to ETS (9). Unborn children are at risk from cigarette smoke, either from the mother smoking or from ETS. They can be underweight at birth and have decreased lung function (10). In contrast to low birth weight and respiratory tract diseases, there is no consistent association between smoke exposure and atopic dermatitis (AD) in children. Krämer et al. reported that school beginners were at a higher risk of developing AD when exposed to ETS (11). Maternal smoking during pregnancy or lactation increased the risk of the offspring developing AD (12, 13) while some studies revealed negative relationships (14–18).

As AD causes significant family stress and healthcare expenditures, identification of risk factors for early prevention is of critical importance. Therefore, in this study, we will investigate if ETS exposure and maternal active smoking during pregnancy will increase the risk of AD on the basis of the cord and maternal blood cotinine.

Methods

Study population

Considering potential environmental exposures and nationwide representativeness, we recruited our subjects from medical centers, regional hospitals, local hospitals, and clinics in Taiwan in 2004. Pregnant women during the third trimester of pregnancy, who had prenatal examination in selected hospitals, were invited to join in this study. Cord blood was collected at delivery. A total of 261 mother and child pairs were recruited with informed consents and the study was approved by the Joint Institution Review Board in Taiwan.

Cases of AD were defined by International Study of Asthma and Allergies in Childhood (ISAAC) questionnaire through all of the three questions 'Has your child ever had an itchy rash which was coming and going for at least 6 months at any time?', 'Has the itchy rash been coming and going over elbows, knees, face, wrists, or generalized (four or more localizations)?', and 'Has your child ever had AD diagnosed by a doctor?'. As described elsewhere, a dermatologist examined a subgroup of the participating young children, and the combination of answers that resulted in the highest sum

of sensitivity and specificity was determined (19, 20). Exclusion criteria include multiple gestation (twins, triplets, etc), inability to answer questions in Chinese, and plan to move out of the area before delivery.

At 2 yr of age, those who developed AD were gathered. We compared concentration of cord and maternal blood cotinine in AD cases and the left non-AD cases in the cohort.

Questionnaire survey

The parents were asked by home interview questionnaires at birth for birth year, parental education levels and occupation, family income, parental history of atopic diseases (such as AD, allergic rhinitis, or asthma), alcohol and drug use, diets and supplements (such as multivitamins) during pregnancy, history of smoke exposure including ETS exposure, and maternal active smoking during pregnancy. From the records of the cooperating hospitals, we gathered neonate's health data at birth, such as head circumference, birth body weight, height, weeks of gestation, and type of delivery. At 2 yr of age, ISAAC questionnaire was performed and some postnatal exposures such as duration of breast feeding and infant formula, early consumption of egg, wheat, soy bean, or shrimp before 1 yr of age, older siblings, furry pets or carpets at home, fungi at house walls, incensing at home, and postnatal ETS exposure were asked.

Laboratory method

The plasma samples of maternal venous blood and umbilical cord blood were stored at -80°C and were sent to National Health Research Institute in Taiwan for cotinine analysis. Average 0.5 ml of plasma was spiked with 15 μl of 10 ng/ml deuterium (D3) (ISOTEC, Miamisburg, OH, USA)-labeled cotinine (Sigma, St Louis, MO, USA) as an internal standard, 100 μl of 5 N KOH (Sigma), and 3 ml of dichloromethane (Mallinckrodt Baker, Phillipsburg, NJ, USA), followed by vigorous shaking for 1 min and centrifugation at 3000 rpm for 10 min. The clear organic phase was transferred to a tube and the aqueous phase was repeatedly extracted with dichloromethane. Then, the organic solutions were combined and dried under nitrogen. Each sample was raised in 150 μl of 5% methanol (Mallinckrodt Baker) in 20 mM ammonium acetate (Merck, Darmstadt, Germany). Finally, 20 μl was injected into high-performance liquid chromatography (PerkinElmer, Boston, MA,

USA) coupled to a triple-quadruple tandem mass spectrometer (API 3000TM; Applied Biosystems, Foster, CA, USA) (HPLC-MS/MS) to measure the cotinine levels. The column was Purospher STAR RP-18 column (4'55 mm, 3 μ m; MERCK, Darmstadt, Germany). The cotinine concentration was determined by comparison of the peak area to a calibration curve which was constructed by injecting nine standard cotinine concentrations ranging from 0.02 to 100 ng/ml and plotted using peak area versus the concentration of cotinine. All-tubes were deactivated. To evaluate the HPLC-MS/MS performance, a calibration curve standard was injected after every 24 samples were analyzed. The detection limit was set at 0.04 ng/ml.

Statistical analysis

Cotinine concentrations were analyzed as categorical variables after division into quartiles, with the lowest quartile used as the reference category. We compared sociodemographic data of mothers and newborns in terms of cotinine levels using chi-squared tests for categorical variables and ANOVA statistics for continuous variables. The trend test was performed by chi-square tests. The relationship of cotinine levels and AD were estimated by means of multiple logistic regression. Analyses were performed in both univariate and multivariate models. Potential confounders from the literature reviews, including infant gender, premature birth, maternal education and occupation, alcohol consumption during pregnancy, family income, parental history of atopy, duration of breast feeding, early consumption of solid food, postnatal ETS exposure, carpets at home, and fungi on house walls were taken into consideration. Only those who made 10% change in point estimate were left in the final model. All hypothesis testing was two-sided at the significance level of 0.05 and performed with the SAS software version 8.2 (SAS Institute Inc., Cary, NC, USA).

Results

At 2 yr of age, 150 mother and child pairs completed the follow-up study and specimen collection. The prevalence of most variables in those lost in the follow-up and in those completed the follow-up was not significantly different from each other. There were 38 (25.3%) children developing AD. Eight (5.4%) out of 150 mothers indicated that they smoked before this pregnancy. Two (1.3%) out of 150 mothers smoked during pregnancy while 38 (25.3%)

mothers reported having ETS exposure with 0.44 ± 1.01 cigarettes/day. Of two mothers with active smoking during pregnancy, one (50%) of their children developed AD. Of 38 mothers with gestational ETS exposure, eight (21.1%) of their children had AD.

As shown in Table 1, maternal education (p for trend = 0.001), family income per year (p for trend = 0.001), maternal occupation (p for trend = 0.015), premature birth (p for trend = 0.055), ETS exposure during pregnancy (p for trend = 0.006), and postnatal ETS exposure (p for trend = 0.002) were found to be significantly different in the four groups on cord blood cotinine levels. Higher cotinine levels were associated with lower maternal education and family income, full-time house wives, premature birth, and ETS exposure during pregnancy and postnatal ETS exposure. The level of maternal blood cotinine was higher than that of cord blood cotinine (mean \pm s.d., 3.61 ± 14.04 ng/ml vs. 2.73 ± 18.48 ng/ml, $p = 0.642$). The geometric mean of maternal blood cotinine was 0.32 ng/ml while cord blood cotinine was 0.11 ng/ml. The maternal and cord plasma concentrations of cotinine were highly correlated ($r = 0.71$, $p < 0.001$) (Fig. 1). Table 2 revealed the association of maternal and cord blood cotinine levels with self-reported smoke exposure during pregnancy.

With respect to Table 3, the risk of AD was found to increase with maternal and cord blood cotinine levels in a dose-response manner (p for trend = 0.01). High cotinine levels (50–75th percentile and >75th percentile) were significantly associated with AD after adjusting for potential confounders. There was no significantly positive association between self-reported ETS exposure amount during pregnancy and AD, [0–3 cigarettes/day vs. 3–5 cigarettes/day; aOR 1.84 (95% CI: 0.23–15.01)] and [0–3 cigarettes/day vs. >5 cigarettes/day; aOR 1.98 (95% CI: 0.18–19.82)] after adjusting for maternal education, maternal occupation, and postnatal ETS exposure (data not shown).

Discussion

This study is an interesting contribution to the literature on the potential association between prenatal smoke exposure and pediatric AD. To the best of our knowledge, the effect of smoke exposure during pregnancy on the development of AD in the offspring on the basis of objective biomarker has not been studied before.

Most people might be very curious about where was smoke exposure from for these

Table 1. Characteristic of the study population according to cotinine levels

	Cord blood cotinine levels (ng/ml)			
	<0.0486 (n = 36)	0.0486–0.1072 (n = 39)	0.1072–0.1936 (n = 39)	>0.1936 (n = 36)
Maternal age (yr), Mean \pm s.d.	29.8 \pm 5.1	30.9 \pm 5.6	28.7 \pm 2.9	27.6 \pm 6.0
Maternal education*(%)				
High school and below	27.8	38.5	46.2	86.1
College and above	72.2	61.5	53.8	13.9
Maternal occupation*(%)				
House wives	27.8	23.1	15.4	44.4
Non-house wives	72.2	76.9	84.6	55.6
Maternal alcohol consumption Yes (%)	5.6	7.7	2.6	5.6
Family income per year* (NT dollars) (%)				
<600,000	14.3	15.8	15.4	50.0
600,000–1,500,000	65.7	60.5	59.0	47.2
>1,500,000	20.0	23.7	25.6	2.8
Infant gender (%)				
Male	47.2	53.8	46.2	58.3
Birth weight (g) Mean \pm s.d.	3189.2 \pm 436.0	3123.3 \pm 549.8	3277.6 \pm 415.7	3131.3 \pm 374.9
Premature birth (< 37 weeks)* Yes (%)	3.1	5.4	2.6	9.7
Parental history of atopy Yes (%)	22.2	13.2	28.2	19.4
Duration of breast feeding (months)				
<6	60.0	69.2	69.2	74.3
\geq 6	40.0	30.8	30.8	25.7
Incensing at home Yes (%)	50.0	48.7	46.2	69.4
ETS exposure during pregnancy* Yes (%)	14.3	17.9	23.7	47.2
Postnatal ETS exposure* Yes (%)	25.7	28.2	23.1	61.8
Recurrent wheezing (>three episodes)* Yes (%)	19.4	23.1	30.8	47.2
Atopic dermatitis* Yes (%)	13.9	20.5	30.8	36.1

ETS, environmental tobacco smoke.

* $p < 0.05$.

pregnant women because most women quit smoking after they were pregnant. In our study, pregnant women had smoke exposure mainly from ETS and only trivial from active smoking. Furthermore, nicotine vapor may persist over the succeeding few days after the last cigarette was smoked in the room because of emission from contaminated room surfaces, people's clothing, or house dust (21). Surprisingly, we found ETS exposure in Taiwan was worse than that in the United States. The cotinine level in our exposed non-smokers was much higher than that reported in California (the geometric mean of maternal cotinine was 0.32 ng/ml vs. 0.08 ng/ml) (22). Though there are regulations to restrict smoking in public and work places, ETS exposure could be obtained at home in Taiwan, where residences are often quite crowded because of limited space (23). Table 1 provides further insight into this

issue. Full-time house wives had higher levels of cotinine than women with other jobs. ETS exposure was particularly serious in the room not well ventilated. These special situations in Taiwan may explain the higher cotinine levels in maternal blood in our population.

Some people might question about the objectivity of the cotinine because its measurements might be influenced by individual differences in rates or patterns of metabolism or excretion, the presence of other sources (such as nicotine-containing food), and the sensitivity and specificity of the analytical methods. In our study, cotinine was analyzed by HPLC–MS/MS, which had excellent validity (24). In addition, recent studies indicated that the pharmacokinetics of cotinine was similar in both smokers and non-smokers (25), suggesting that the degree of metabolism variability was likely to be similar

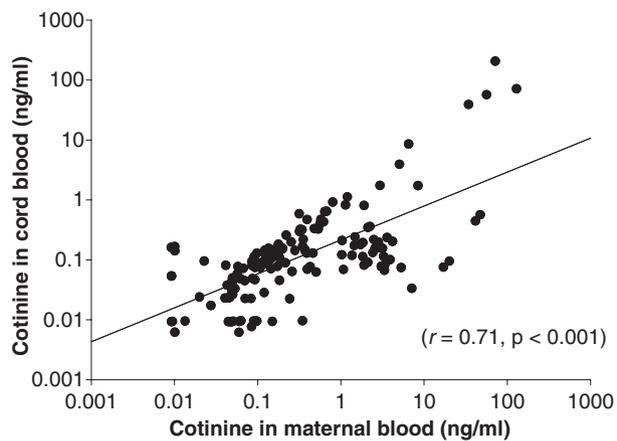


Fig. 1. The correlation of maternal and cord blood cotinine.

Table 2. The association of maternal and cord blood cotinine levels with self-reported smoke exposure during pregnancy

	No. (%)	Maternal blood cotinine levels (ng/ml), Mean \pm s.d.	Cord blood cotinine levels (ng/ml), Mean \pm s.d.
Total	150	3.61 \pm 14.04	2.73 \pm 18.48
Non-exposed non-smokers	110 (73.3)	2.05 \pm 6.18	0.58 \pm 3.96
Exposed non-smokers	38 (25.3)	5.69 \pm 21.40	5.91 \pm 30.49
Number of cigarettes/day			
<3	23 (15.3)	4.35 \pm 13.05	0.19 \pm 0.20
3–5	8 (5.3)	3.91 \pm 14.73	8.40 \pm 41.03
>5	7 (4.7)	17.58 \pm 45.15	10.19 \pm 24.76
Smokers	2 (1.3)	39.26 \pm 46.28	106.94 \pm 139.16

for non-smokers. While nicotine is contained in some foods, such as tomatoes, potatoes, cauliflower, and black tea (26), at usual food consumption levels dietary nicotine is negligible compared with moderate ETS exposure and may not significantly confound determination of cotinine level in our study. Besides, we discovered that maternal and cord blood cotinine showed stronger association with AD than self-reported ETS exposure from our results. Based

on the above reasons, we believe our exposure assessment is reliable. Data from this study provided evidence that exposure to smoking either from maternal smoking or ETS exposure during pregnancy may play a role in the development of AD. Even after adjusting for potential confounders, the risk of AD was significantly associated with cotinine levels and had dose-response relationship. Some studies supported our observation and also demonstrated that exposure to ETS in genetically predisposed children was associated with an increased risk of sensitization to house dust mites (11, 12). Another study indicated that during early childhood, prenatal and postnatal ETS-exposed children had a significantly higher risk of sensitization to food allergens compared with children never exposed to ETS (27). In contrast with our study, Mills et al. found that cigarette smoking was not a risk factor for AD (14). Miyake et al. discovered that maternal smoking and household smoking in the same room as the infant were not related to the risk of AD (15). These discrepancies might be because of recall bias or different age groups. Furthermore, the possible association might have been masked by the fact that families at risk have changed their smoking habits following recent health education and prevention programs. Magnusson et al. and Linneberg et al. reported that gestational smoke exposure had mild protective effect on AD (16, 17). However, lack of the objective biomarker for smoke exposure and absence of the control options for hereditary factors in their studies may have affected the results. In addition, Noakes et al. found that there were no effects of maternal smoking on the rates of allergen sensitization and AD at 12 months of age (18). Differences of the adjusting potential confounders and the population that failed to follow up might account for the variation of the results.

If smoke exposure during pregnancy is a significant risk factor for AD, people might be

Table 3. The association of atopic dermatitis and blood cotinine levels by univariate and multivariate logistic regression

Maternal blood cotinine levels (ng/ml)	<0.0843 (n = 36)	0.0843–0.1920 (n = 39)	0.1920–1.4563 (n = 38)	>1.4563 (n = 36)	Trend test
Crude OR (95% CI)	1.00	2.76 (0.78–9.76)	2.86 (0.81–10.13)	5.09 (1.48–17.53)*	
Adjusted OR (95% CI)†	1.00	3.51 (0.85–14.42)	4.59 (1.12–18.70)*	5.33 (1.33–21.41)*	0.01*
Cord blood cotinine levels (ng/ml)	<0.0486 (n = 36)	0.0486–0.1072 (n = 39)	0.1072–0.1936 (n = 39)	>0.1936 (n = 36)	Trend test
Crude OR (95% CI)	1.00	1.65 (0.49–5.63)	2.76 (0.86–8.82)	3.50 (1.09–11.21)*	
Adjusted OR (95% CI)†	1.00	2.60 (0.62–10.92)	4.44 (1.12–17.66)*	5.71 (1.40–23.32)*	0.01*

OR, odds ratio; CI, confidence interval; ETS, environmental tobacco smoke.

* $p < 0.05$.

†Model adjusted for premature birth, maternal education, and postnatal ETS exposure.

curious about how smoke enters the body of the fetus. Consistent with previous study (28), we found that there was a positive linear relationship between maternal and neonatal cotinine concentrations (Fig. 1) and the mean cotinine concentration was lower in cord blood than maternal blood in our study. This implied that cotinine, a metabolite of nicotine, used to quantify exposure to tobacco smoke, readily gained access from maternal to fetal circulation and accumulated in the fetal body. In addition, Seymour et al. reported that exposure of ovalbumin-sensitized mice to ETS by inhalation elicited a rapid and prolonged exaggerated response with respect to IgE, IgG1, eosinophils, and Th2 cytokines [particularly interleukin (IL)-4 and IL-10] (29). Magnusson et al. discovered that cord serum IgE was increased in newborns if the mother smoked (13). Further evidence of altered immune responses from maternal smoking was found in a study of Australian birth cohort. It demonstrated that maternal smoking in pregnancy was associated with significantly higher neonatal Th2 (IL-13 protein) responses to both house dust mite and ovalbumin (30). Sensitization to allergens along with the adjuvant effect of smoking on the Th2 response may be responsible for the prevalence of allergic symptoms seen in individuals with tobacco smoke exposure. Furthermore, tobacco smoke, like other air pollutants, may have irritant effects on skin and mucous membranes, thereby facilitating the penetration of potential allergens into the body and causing more AD symptoms and an elevated risk of sensitization (29). Then, there raised another question about which kind of ingredients in cigarettes mainly contribute to AD. The diversity of chemical ingredients found in tobacco (4000 chemicals, of which 300 were known carcinogens) was matched by the diversity of tobacco-related health problems (31). People might suppose that nicotine was the arch criminal because it was the main constituent of tobacco. Harkavy had shown that nicotine was not the responsible antigenic component of tobacco leaf, although its role as a hapten was a possibility (32). Relevant information about the leading element in cigarettes to AD, however, was not available. Further studies are warranted.

The strengths of our study included the population-based prospective cohort design and objective biomarkers of smoking exposure. Cotinine was a valid quantitative predictor of the level of smoke exposure for population studies. However, because the smoke exposure was intermittent and cotinine was a biomarker of recent exposure with a half-life of approximately

20 h (6), one of our limitations was that the cotinine measurement from cord blood was a single opportunistic measurement at delivery. In our study, cord blood cotinine level revealed a good correlation to self-reported smoke exposure amount (Table 3) and smoking habits investigated in our questionnaire were relatively stable over time. According to previous study, cotinine levels remained fairly constant and at near steady-state values throughout the day (6). Furthermore, if measurement error did occur, it tended to be toward the null and the effect of exposure was likely to be underestimated. Another limitation was that there is probably a considerable overlap between pregnancy and postnatal exposure and therefore also an association between cotinine levels and postnatal exposure, which makes it very difficult to disentangle the relevance of the different time windows. As the correlation and the collinearity of pregnancy and postnatal ETS exposure was low ($r = 0.3$, $p = 0.06$) in our study, we put postnatal ETS exposure into the model as a potential confounder. Levels of cord cotinine concentration were still positively related to development of AD even after adjusting for postnatal ETS exposure. The other limitation was that a potential selection bias could have occurred if non-participants were different on sociodemographic or smoking characteristics from participants. However, the prevalence of most variables in those lost in the follow-up and in those completed the follow-up was not significantly different from each other. If a selection bias would occur, it might be trivial.

In conclusion, we observe that ETS exposure in Taiwan is more serious than expected. Our study supports the hypothesis that ETS exposure and maternal smoking during pregnancy might be a risk factor for pediatric AD. Avoidance of prenatal smoke exposure should be paid more attention.

Acknowledgments

This study was supported in part by grants from the Bureau of Health Promotion, Department of Health (BHP-PHRC-92-4 and DOH93-HP-1702), and the National Science Council (NSC95-2314-B-002-269 and NSC96-2314-B-192-001) of Taiwan.

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