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TERPENOIDS AND FLAVONOIDS FROM *PSEUDOTSUGA WILSONIANA*

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Abstract—Four new and 27 known compounds were isolated from the heartwood of *Pseudotsuga wilsoniana*. The structures of the new compounds, 9-oxobisabola-1,3,5.7*E*-tetraen-15-oic acid methyl ester, 11-hyd-roxybisabola-1,3,5.9-tetraen-15-oic acid methyl ester, (S)-12-hydroxyagathic acid and 4',5-dihydroxy-7-methoxy-6-methylflavanone, were determined by chemical and spectroscopic methods. © 1998 Published by Elsevier Science Ltd. All rights reserved

INTRODUCTION

Pseudotsuga wilsoniana (Taiwan Douglas fir) is an endemic species in Taiwan [1]. The wood is often used in building. The chemical constituents of plants of the genus *Pseudotsuga* are rarely investigated. Previous reports [2–9] indicated that terpenes and flavones are the main constituents in the leaves, wood and root of *P. menziesii* (Douglas fir). We present herein the first report of the chemical constituents of the heartwood of *P. wilsoniana*.

RESULTS AND DISCUSSION

We isolated 31 compounds from the EtOAc-soluble part of the heartwood of P. wilsoniana. By analyses of the physical and spectroscopic properties (mp, $[\alpha]$, IR, UV, mass spectrometry, ¹H and ¹³C NMR), 27 known compounds were readily identified as borneol (1) [10], epijuvabiol (2) [11], epijuvabione (3) [12], methyl atlantonate (4) [13], atlantolone (5) [14], nerolidol (8) [15], 6,8-cycloeudesm-4(15)-en-1-ol (9) [16, 17], α-cadinol (10) [18], cis-communic acid (11) [19], 15-nor-14-oxolabda-8(17),12E-dien-19-oic acid (12) [20], dehydroabietic acid (14) [21], pomiferin A (15) [22], methyl 15-hydroxydehydroabietate (16) [23], isopimaric acid (17) [24], sandaracopimaric acid (18) [24], chrysin (19) [25], pinocembrin (20) [26], naringenin (21) [27], strobopinin (22) [28, 29], 4', 5, 7-trihydroxy-6-methylflavanone (poriol, 23) [30], 5-hydroxy-7methoxy-6-methylflavanone (24) [28, 29], meridinol (26) [31], 4-hydroxybenzaldehyde (27), vanillin (28),

trans-4,4'-dihydroxy-3,3'-dimethoxystilbene (**29**) [32], β -sitosterol (**30**) and β -sitostenone (**31**) [33]. The structures of the four new compounds **6a**, **7**, **13** and **25** were determined as follows.

The molecular formula $C_{16}H_{20}O_3$ of compound **6a** was deduced from its exact mass $[M]^+$ at m/z 260.1418. Based on the analyses of ¹H and ¹³C NMR spectra, the structure of **6a** was assigned as 9-oxobisabola-1,3,5,7*E*-tetraen-15-oic acid. The resonances at δ 7.51 (d, J = 8.4 Hz) and 8.02 (d, J = 8.4 Hz) corresponded to the aromatic protons of a phenyl moiety having two electron-withdrawing groups on para positions. The signals at $\delta_{\rm H}$ 3.91 (s) and $\delta_{\rm C}$ 166.6 (s) corresponded to the methyl ester group. An olefinic proton and a vinyl methyl group appeared at $\delta_{\rm H}$ 6.49 (s) and 2.51 (s). The resonances at δ_C 201.3 (s), 152.1 (s) and 125.8 (d) were ascribed to the conjugated enone moiety. The base peak at m/z 203, $[M-C_4H_9]^+$, was attributed to a Norrish-type I cleavage. An NOE study, i.e. irradiation of H-8 (δ 6.49) causing a 10% enhancement of the signal at δ 7.51 (H-3 or H-5), supported that 6a had the (E)-configuration. Compound 6aunderwent isomerisation in part, giving 6b, on standing in $CDCl_3$ solution. The (Z)-configuration in 6b was also verified by an NOE study, i.e. irradiation of H-8 (δ 6.17) causing a 10% enhancement of the methyl group (δ 2.14) at C-7. Due to the steric effect, the β phenyl group and the enone moiety in 6b might not be coplanar; thus 6b showed a UV absorption at shorter wavelength (λ_{max} 256 nm) than that of **6a** (λ_{max} 284 nm).

Compound 7 showed a characteristic IR absorption at 3431 cm⁻¹ for a hydroxyl group. The NMR signals at $\delta_{\rm H}$ 7.21 (*d*, J = 8.4 Hz), 7.94 (*d*, J = 8.4 Hz) and 3.88 (*s*) corresponded to a *para*-substituted benzoic acid methyl ester. The signals at δ 5.45 (*m*) and 5.54 (*d*,

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J = 15.6 Hz) were attributable to two olefinic protons with a *trans*-relationship. Compound 7 (C₁₆H₂₂O₃), showing the exact mass [M]⁺ at m/z 262.1569, was assigned as 11-hydroxy*bis*abola-1,3,5,9-tetraen-15-oic acid methyl ester. The structure was confirmed by the ¹³C and HMBC NMR spectra. Compound 7 exhibited an optical rotation, $[\alpha]_D$ + 27.8 but the C-7 chirality is unknown.

The structure of 13 was determined to be 12-hydroxyagathic acid from spectral analyses. The IR spectrum showed a strong broad absorption at 3600–2500 cm⁻¹. Two carboxyl groups appeared at $\delta_{\rm C}$ 173.2 (*s*) and 183.3 (*s*) in the ¹³C NMR spectrum. The resonances at $\delta_{\rm H}$ 4.80 (*brt*) and $\delta_{\rm C}$ 84.3 (*d*) corresponded to a secondary alcohol. The resonances at $\delta_{\rm H}$ 4.65 (*br s*), $\delta_{\rm H}$ 4.91 (*brs*), $\delta_{\rm C}$ 147.2 (*s*) and $\delta_{\rm C}$ 107.8 (*t*) were attributable to a 1,1-disubstituted double bond, whereas the resonances at $\delta_{\rm H}$ 5.74 (*br s*), $\delta_{\rm C}$ 116.7 (*d*) and $\delta_{\rm C}$ 169.3 (s) were ascribed to a trisubstituted double bond. Irradiation of Me-13 (δ 2.06) caused an enhancement of H-14 (δ 5.74), indicating the (Z)configuration. The resonances at $\delta_{\rm H}$ 0.59 (s) and 1.23 (s) corresponded to two methyl groups on tertiary carbinyl centres (C-10 and C-4). The C-10 methyl group, orienting on the β -face, appeared up-field (δ 0.59) due to the shielding effect of the C-4 carboxyl group. The structure of 13 was confirmed by COSY and HMBC spectra. According to the literature [34], the C-12 chirality can be determined from the chemical shifts of the vinyl protons at C-17. Due to the deshielding effect of the hydroxyl group at C-12, the H-17 protons in the (12S)-isomer, presumably existing as the conformer A, occurred at lower fields (near δ 4.7) than the corresponding protons (near δ 4.4) in the (12R)-isomer, presumably existing as the conformer B. Compound 13, exhibiting two H-17 protons







at δ 4.65 and 4.91, was considered to have the (12S)-chirality.

Compound 25 was readily recognised as a Cmethylflavanone from its characteristic NMR resonances. Its structure was assigned as 4',5-dihydroxy-7-methoxy-6-methylflavanone, a poriol methyl ether. The axially-oriented H-2 showed resonance at δ 5.33 (dd, J = 13, 3Hz). Two C-3 protons appeared at δ 2.75 (dd, J = 17, 3 Hz) and 3.08 (dd, J = 17, 13 Hz). The resonances at δ 6.87 (2 H, d, J = 8.5 Hz) and 7.33 (2 H, d, J = 8.5 Hz) were attributable to four protons on a para-disubstituted phenyl group (B-ring). Another aromatic proton, occurring at δ 6.05 as a singlet, was ascribed to H-8. Two methyl groups appeared at $\delta_{\rm H}$ 1.99 (s, ArCH₃) and 3.81 (s, ArOCH₃). The C-5 hydroxyl group, hydrogen-bonded with the carbonyl group, resonated at a low-field δ 12.05. The regiochemistry of the A-ring was supported by NOE experiments. Irradiation of the C-6 methyl group (δ 1.99) caused enhancements of the C-7 methoxyl and C-5 hydroxyl groups. An enhancement of H-8 (δ 6.05) was observed on irradiation of the C-7 methoxyl group (δ 3.81). The mass spectrum showed the [M]⁺ at m/z 300, a peak at m/z 207 [M-C₆H₄OH]⁺ due to fragmentation of the B-ring and an intense peak at m/z 180 due to fragmentation of the C-ring (a retro-Diels-Alder reaction of pyranone). On treatment with diazomethane, compound 23 vielded a monoether 23a and a bisether 23b. The monoether 23a and compound 25 were determined to be enantiomers by spectral analyses and comparison of optical rotations. As compound 23 is known to have the (2R)-configuration, compound 25 should have the (2S)-configuration.

In summary, terpenes (1-18) and flavones (19-25) are rich in the EtOAc-soluble part of the heartwood of *P. wilsoniana*, similar to that found in Douglas fir. In addition, one lignan (26), two sterols (30 and 31) and three phenols (27-29) were isolated from this species.

EXPERIMENTAL

General

HPLC: Hibar Lichrosorb Si 60 column (10 μ m, 25 cm \times 1 cm i.d.). TLC: Merck silica gel 60F sheets.

Plant material

A specimen of *P. wilsoniana* Hayata is deposited in our laboratory. The wood (12–16 cm diameter), collected from mountains of central Taiwan, was airdried (880 g) and sliced after removal of the bark. This material was exhaustively extracted with Me₂CO (7 1×3). The combined extracts were concd to give an oil (12.8 g). The oil was partitioned between EtOAc and H₂O (1:1). The EtOAc-sol. part was concd (10.5 g) and subjected to silica-gel CC by elution with gradients of hexane–EtOAc–Me₂CO. Appropriate frs were combined and components purified by HPLC or recrystallisation. Compounds 1 (6 mg), 2 (14 mg), 3 (16 mg), 4 (13 mg), 5 (13 mg), 6a (7 mg), 7 (5 mg), 8 (24 mg), 9 (6 mg), 10 (7 mg), 11 (69 mg), 12 (15 mg), 13 (5 mg), 14 (46 mg), 15 (4 mg), 16 (11 mg), 17 (20 mg), 18 (18 mg), 19 (37 mg), 20 (55 mg), 21 (14 mg), 22 (14 mg), 23 (28 mg), 24 (12 mg), 25 (4 mg), 26 (38 mg), 27 (6 mg), 28 (25 mg), 29 (28 mg), 30 (20 mg) and 31 (7 mg) were obtained.

Borneol (1)

Solid. Mp 209–210°. $[\alpha]_{\rm D}^{23}$ – 35.7 (MeOH; *c* 0.8).

Epijuvabiol (2)

Oil. $[\alpha]_{D}^{23}$ + 48.3 (MeOH; *c* 0.95).

Epijuvabione (3)

Oil. $[\alpha]_{D}^{23}$ + 56.0 (MeOH; c 1.01).

Atlantonic acid methyl ester (4)

Oil. $[\alpha]_D^{23} + 36.0$ (MeOH; *c* 1.21).

Atlantolone (5)

Oil. $[\alpha]_D^{23}$ + 16.5 (CHCl₃; *c* 0.41).

9-Oxobisabola-1,3,5,7*E*-tetraen-15-oic acid methyl ester (**6a**)

Oil. IR ν_{max}^{meat} cm⁻¹: 1717, 1678, 1276. UV $\lambda_{max}^{\text{MeOH}}$ nm (ε): 284 (26 600), 219 (10 800). ¹H NMR (CDCl₃, 300 MHz): δ 0.95 (d, J = 6.6 Hz, H-12, 13), 2.18 (m, H-11), 2.41 (d, J = 7.0 Hz, H-10), 2.51 (s, H-14), 3.91 (s, OMe), 6.49 (s, H-8), 7.51 (d, J = 8.4 Hz, H-3, 5), 8.02 (d, J = 8.4 Hz, H-2, 6). ¹³C NMR (CDCl₃, 75 MHz): δ 18.2 (C-14), 22.7 (C-12, 13), 25.2 (C-11), 52.2 (OMe), 54.0 (C-10), 125.8 (C-8), 126.5 (C-3, 5), 129.8 (C-2, 6), 130.4 (C-1), 147.1 (C-4), 152.1 (C-7), 166.6 (C-15), 201.3 (C-9). EIMS (70 eV) m/z (rel. int.): 260 [M]⁺ (16), 245 (21), 203 (100), 159 (40), 115 (40), 91 (12), 59 (21). HR-MS for C₁₆H₂₀O₃ requires 260.1413; found 260.1418.

9-Oxobisabola-1,3,5,7Z-tetraen-15-oic acid methyl ester (**6b**)

Oil. IR v_{max}^{meat} cm⁻¹: 1716, 1683, 1273. UV $\lambda_{max}^{\text{MeOH}}$ nm (ε): 256 (11 100), 230 (11 700). ¹H NMR (CDCl₃, 400 MHz): δ 0.79 (d, J = 6.4 Hz, H-12, 13), 2.02 (m, H-11), 2.11 (d, J = 7.2 Hz, H-10), 2.14 (s, H-14), 3.90 (s, OMe), 6.17 (s, H-8), 7.20 (d, J = 8.0 Hz, H-3, 5), 8.00 (d, J = 8.0 Hz, H-2, 6). ¹³C NMR (CDCl₃, 75 MHz): δ 22.5 (C-12, 13), 24.8 (C-11), 26.9 (C-14), 52.1 (OMe), 52.3 (C-10), 127.1 (C-3, 5), 127.4 (C-8), 129.5 (C-1), 129.6 (C-2, 6), 146.0 (C-4), 150.8 (C-7), 165.7 (C-15), 200.9 (C-9). EIMS (70 eV) m/z (rel. int.): 260 [M]⁺ (21), 245 (44), 203 (100), 159 (45), 115 (33), 91 (16), 59 (28). HR-MS for $C_{16}H_{20}O_3$ requires 260.1413; found 260.1420.

11-Hydroxybisabola-1,3,5,9-tetraen-15-oic acid methyl ester (7)

Oil. $[\alpha]_{23}^{D3}$ +27.8 (MeOH; *c* 0.36). IR ν_{max}^{neat} cm⁻¹: 3431, 1714. UV λ_{max}^{MeOH} nm (ϵ): 240 (12 500). ¹H NMR (CDCl₃, 200 MHz): δ 1.20 (*s*, Me), 1.21 (*s*, Me), 1.24 (*d*, *J* = 6.9 Hz, H-14), 2.28 (*m*, H-8), 2.81 (*m*, H-7), 3.88 (*s*, OMe), 5.45 (*m*, H-9), 5.54 (*d*, *J* = 15.6 Hz, H-10), 7.21 (*d*, *J* = 8.4 Hz, H-3, 5), 7.94 (*d*, *J* = 8.4 Hz, H-2, 6). ¹³C NMR (CDCl₃, 50 MHz): δ 21.1 (C-14), 29.8 (C-12, 13), 40.1 (C-7), 40.7 (C-8), 52.0 (OMe), 70.6 (C-11), 124.6 (C-9), 127.1 (C-3, 5), 128.0 (C-1), 129.7 (C-2, 6), 140.1 (C-10), 152.4 (C-4), 167.1 (C-15). EIMS (70 eV) *m*/*z* (rel. int.): 262 [M]⁺ (1), 247 (6), 244 (5), 215 (26), 204 (9), 163 (100), 149 (20), 82 (58). HR-MS for C₁₆H₂₂O₃ requires 262.1570; found 262.1569.

Nerolidol (8)

Oil. $[\alpha]_{D}^{23} + 16.0$ (CHCl₃; *c* 0.4).

6,8-Cycloeudesm-4(15)-en-1-ol (9)

Solid, mp 72–73°. $[\alpha]_D^{23}$ +13.7 (CHCl₃; *c* 0.3).

 α -Cadinol (10)

Solid. mp 74–75°. $[\alpha]_D^{23}$ –45.5 (CHCl₃; *c* 0.49).

Cis-Communic acid (11)

Oil. $[\alpha]_{D}^{23} + 3.3$ (MeOH; *c* 5.5).

15-Nor-14-oxolabda-8(17),12E-dien-19-oic acid (12)

Oil. $[\alpha]_{D}^{23} + 39.5$ (CHCl₃; *c* 0.15).

(S)-12-Hydroxyagathic acid (13)

Solid. mp 181–182°. $[\alpha]_D^{23}$ + 57.6 (MeOH; *c* 0.21). IR ν_{max}^{KBr} cm⁻¹: 3700–2500, 3077, 1751, 1684, 1633, 1442. ¹H NMR (CDCl₃, 300 MHz): δ 0.59 (*s*, H-20), 1.23 (*s*, H-18), 2.06 (*s*, H-16), 4.65 (*br s*, H-17), 4.80 (*br t*, *J* = 5.6 Hz, H-12), 4.91 (*br s*, H-17), 5.74 (*s*, H-14). ¹³C NMR (CDCl₃, 75 MHz): δ 12.4 (C-20), 14.7 (C-16), 19.7 (C-2), 25.9 (C-6), 27.5 (C-11), 28.9 (C-18), 37.7 (C-3), 38.5 (C-7), 39.3 (C-1), 40.7 (C-10), 44.1 (C-4), 51.8 (C-9), 56.2 (C-5), 84.3 (C-12), 107.8 (C-17), 116.7 (C-14), 147.2 (C-8), 169.3 (C-13), 173.2 (C-15), 183.3 (C-19). EIMS (70 eV) *m*/*z* (rel. int.): 332 [M-H₂O]⁺ (3), 286 (2), 235 (100), 217 (12), 189 (63), 121 (40), 107 (31). HR-MS for C₂₀H₂₈O₄ requires 332.1988; found 332.1995.

Dehvdroabietic acid (14)

The acid was subjected to methylation with CH_2N_2 to give the corresponding Me ester. Solid, mp 61–63°. [α]_D²³ + 56.8 (CHCl₃; *c* 0.2).

Pomiferin A (15)

Oil. $[\alpha]_D^{23} + 39.6$ (CHCl₃; *c* 0.19).

Methyl 15-hvdroxydehydrobietate (16)

Solid, mp 76–78°. $[\alpha]_D^{23}$ +46.8 (CHCl₃; *c* 0.2).

Isopimaric acid (17)

Solid, mp 160–161°. $[\alpha]_{D}^{23}$ +1.5 (CHCl₃; c 0.7).

Sandaracopimaric acid (18)

Solid, mp 167–169°. $[\alpha]_{D}^{23}$ –18.5 (CHCl₃; *c* 1.2).

Chrysin (19)

Solid, mp 274–275°.

Pinocembrin (20)

Solid, mp 193–194°. $[\alpha]_D^{23}$ – 47.3 (MeOH; *c* 5.48).

Naringenin (21)

Solid, mp 248–250°. $[\alpha]_{D}^{23}$ – 24 (MeOH; *c* 0.15).

Strobopinin (22)

Solid, mp 225–228°. $[\alpha]_{D}^{23}$ – 55.2 (MeOH: *c* 0.57).

4'5,7-Trihydroxy-6-methylflavanone (23)

Solid, mp 270–272°. $[\alpha]_D^{23} - 53.8$ (MeOH; *c* 0.4). Treatment of **23** (8 mg) with CH₂N₂ in Et₂O gave a monoether **23a** (2.7 mg) and a *bis*ether **23b** (3 mg), along with a recovery of **23** (1.8 mg). **23a**. Mp 91.5– 92°. $[\alpha]_D^{23} - 57.8$ (MeOH; *c* 0.27). **23b**. Mp 178–178.5°. $[\alpha]_D^{23} - 39.2$ (CHCl₃; *c* 0.3).

5-Hydroxy-7-methoxy-6-methylflavanone (24)

Solid, mp 77–78°. $[\alpha]_{D}^{23}$ +45 (MeOH; *c* 0.02).

4'5-Dihydroxy-7-methoxy-6-methylflavanone (25)

Solid, mp 92–93°. $[\alpha]_D^{23} + 74.4$ (MeOH; *c* 0.03). IR $\nu_{\text{max}}^{\text{KBr}}$ cm⁻¹: 3326, 1625, 1445. UV $\nu_{\text{max}}^{\text{MeOH}}$ nm (ε): 291 (9937), 224 (shoulder, 14,444), 214 (15,348). ¹H NMR (CDCl₃, 300 MHz): δ 1.99 (*s*, Me), 2.75 (*dd*, *J* = 17.3 Hz, H-3), 3.08 (*dd*, *J* = 17, 13 Hz, H-3), 3.81 (*s*, OMe), 5.33 (*dd*, *J* = 13, 3 Hz, H-2), 6.05 (*s*, H-8), 6.87 (*d*, *J* = 8.5 Hz, H-3', 5'), 7.33 (*d*, *J* = 8.5 Hz, H-2', 6'). ¹³C NMR (CDCl₃, 75 MHz): δ 6.9 (Me), 43.3 (C-3), 55.8 (OMe), 79.1 (C-2), 90.7 (C-8), 102.8 (C-10), 106.0 (C-6), 115.6 (C-3', 5'), 128.0 (C-2', 6'), 130.8 (C-1'), 156.0 (C-4'), 160.4 (C-5'), 161.2 (C-9), 165.7 (C-7), 196.0 (C-4). EIMS (70 eV) m/z (rel. int.): 300 [M]⁺ (100), 299 (42), 207 (18), 194 (20), 180 (85), 152 (77), 135 (20), 120 (32). HR-MS for C₁₇H₁₆O₅ requires 300.0998; found 300.1000.

Meridinol (26)

Solid, mp 121–122°. $[\alpha]_D^{23} = 27.3$ (CHCl₃; *c* 0.53).

4-Hydroxybenzaldehyde (27)

Solid, mp 114–116°.

Vanillin (28)

Solid, mp 83–84[°].

Trans-4,4'-*Dihydroxy*-3,3'-*dimethoxystilbene* (**29**) Solid, mp 211–212°.

β -Sitosterol (30)

Solid, mp 136–138°. $[\alpha]_D^{23}$ – 38 (CHCl₃; *c* 1.5).

 β -Sitostenone (31)

Solid, mp 94–96°. $[\alpha]_D^{23} + 79$ (CHCl; *c* 1.6).

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