

Differential *in Vitro* Suppressive Effects of Steroids on Leukocyte Phagocytosis in Two Teleosts, Tilapia and Common Carp

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The objectives of this study were to investigate the potential roles of cortisol and gonadal steroids in the phagocytic activity of peripheral blood leukocytes in two teleosts, tilapia (*Oreochromis niloticus* × *O. aureus*) and common carp (*Cyprinus carpio*). An *in vitro* microtiter plate assay, measuring incorporation of FITC-latex beads into peripheral blood leukocytes, was developed for the first time in teleosts. Peripheral blood leukocytes were cultured in AL medium with tested compounds in a microfluor black plate at 25°C. FITC-latex beads were further incubated for phagocytosis and engulfed fluorescent intensity in phagocytes was detected fluorometrically. Cortisol suppressed leukocyte phagocytosis in a dose (10^{-14} to 10^{-4} M)- and time (0.5 to 8 h)-dependent manner in tilapia. The glucocorticoid agonist dexamethasone had a suppressive effect similar to that of cortisol, while cortisone and the mineralocorticoid aldosterone had only a weak effect in tilapia. High doses of estradiol and ethynylestradiol, but not of estrone, suppressed phagocytosis in tilapia. No suppressive effect on phagocytosis was observed with various concentrations of progesterone, testosterone, and 11-ketotestosterone. Triiodothyronine was also inactive on phagocytosis. A combination of estradiol and cortisol potentiatingly suppressed phagocytosis. Actinomycin D and cycloheximide blocked the sup-

pressive effects of cortisol and estradiol. Cortisol had weaker suppressive effects on the phagocytosis of leukocytes in common carp than tilapia. Other steroids had no suppressive action on phagocytosis in common carp. It is concluded that the suppressive effects of cortisol and estradiol on phagocytosis in tilapia are mediated by specific glucocorticoid receptors and estrogen receptors, respectively. Cortisol would play a main and important role on the down-regulation of phagocytic activity. Sexual steroids, such as estradiol, also could interact with cortisol to further suppress immunity in tilapia. Differential responsiveness of the immune system to suppressive effects of steroids, among teleosts species, has been demonstrated. © 2001 Academic Press

Key Words: fish; cortisol; estradiol; steroids; phagocytosis; leukocyte; tilapia; common carp.

INTRODUCTION

The interaction of the endocrine and immune systems is an important regulatory process in vertebrates. The development of the immune system and immunostimulation is likely to be dependent on growth hormone and prolactin in mammals (Blalock, 1994; Berczi, 1997; Clark, 1997) and fish (Calduch-Giner *et al.*, 1995; Balm, 1997; Yada *et al.*, 1999). Thyroid hor-

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mones stimulated phagocytosis activity in masu salmon, *Oncorhynchus masou* (Nagal *et al.*, 1994). Suppression of the immune system by glucocorticoids has been shown in mammals (Goulding and Flower, 1997) and fish (Balm, 1997). Thus, the effects of growth hormone/prolactin and of glucocorticoids on immune and inflammatory reactions are antagonistic in nature. Stress via cortisol could inhibit the leukocyte function in the specific and/or nonspecific immunity (T and B lymphocytes, natural killer cells, macrophages, and neutrophils) in mammals (Goulding and Flower, 1997). However, cortisol may also stimulate monocyte phagocytosis mediated by the β -glucan receptor in human (Kay and Czop, 1994) and increase phagocytosis of neutrophils in bovine milk (Fox and Heald, 1981). Cortisol effects on the immunity of fish include a decreased number of blood leukocytes and antibody *in vivo* in Atlantic salmon, *Salmo salar* (Ellis, 1981; Mazur and Iwama, 1993), a decreased number of lymphocytes and increased susceptibility to infection in brown trout, *Salmo trutta in vivo* (Pickering and Dutton, 1983; Pickering, 1984), decreased phagocytosis in the macrophage of spleen and anterior kidney in rainbow trout, *O. mykiss in vivo* (Narnaware *et al.*, 1994), inhibition of neutrophil apoptosis in common carp, *Cyprinus carpio in vitro* (Weyts *et al.*, 1998), decreased numbers and mitogenic activities of blood lymphocytes but increased neutrophils in channel catfish, *Ictalurus punctatus in vitro* (Ellsaesser and Clem, 1987), and a reduced mitogenic activity of blood lymphocytes in Atlantic salmon, *S. salar, in vitro* (Espelid *et al.*, 1996). Dexamethasone but not cortisol significantly depressed macrophage phagocytosis *in vitro* in the anterior kidney and spleen of rainbow trout, *O. mykiss* (Narnaware *et al.*, 1994). Cortisol even could have permissive, suppressive, or stimulatory actions in various animals (Sapolsky *et al.*, 2000).

There are few consistent data on the effects of sex steroids on the immune system (Grossman, 1985; Balm, 1997). The phagocytosis of leukocytes was increased but the numbers of blood lymphocyte and monocytes were reduced by estradiol treatment in the sow (Magnusson and Einarsson, 1990). Estradiol inhibited cell-mediated immunity and phagocytosis in porcine leukocytes (Magnusson, 1991; Josefsson *et al.*, 1992). Estradiol also suppressed phagocytosis in the macrophage cell line of goldfish (Wang and Belosevic, 1995). Testosterone inhibited macrophage phagocytosis

in chicken (Al-Afaleq and Homeida, 1998) and killed leukocytes in the anterior kidney of chinook salmon, *O. tshawytscha* (Slater and Schreck, 1997). Progesterone but not estradiol significantly reduced neutrophil function in steers (Roth *et al.*, 1982) and suppressed cell-mediated immunity in hamster (Kincl and Ciaccio, 1980).

Phagocytosis is an important process in the nonspecific immune system. Macrophages and neutrophils are the main cells responsible for phagocytosis. Leukocytes collected from anterior kidney or spleen are often used for the *in vitro* study of phagocytosis (Narnaware *et al.*, 1994; Narnaware and Baker, 1996; Calduch-Giner *et al.*, 1997; Narnaware *et al.*, 1997; Mondal and Rai, 1999). The circulatory system contains large numbers of phagocytic cells, such as hemocytes in invertebrates and macrophages/neutrophils in vertebrates, and they have been used to obtain reproducible, quantitative, and nonsubjective phagocytosis data via a microtiter plate assay. This system provides a valuable tool for further investigating the interaction between the endocrine and immune systems. The present objectives were to investigate the potential roles of cortisol and gonadal steroids on the phagocytosis activity of peripheral blood leukocytes in a microtiter plate assay in two teleost species, tilapia and common carp.

MATERIALS AND METHODS

Fish

Male tilapia (*Oreochromis niloticus* \times *O. aureus*, average body weight 920 ± 161 g, body length 54 ± 8 cm) and common carp (*C. carpio*, average body weight 575 ± 100 g, body length 33.0 ± 1.5 cm) were purchased from the fish market. The fish were kept in a freshwater system for more than 1 month before study.

Preparation of Peripheral Blood Leukocytes

Fish were anesthetized in 0.02% benzocaine solution (Sigma; St. Louis, MO) and blood was collected from the caudal vasculature of fish with heparin (Sigma). The preparation of blood leukocytes was modified

from the methods of Miller *et al.* (1994). Blood was mixed with AL medium (AIM-V medium and Leibovitz's L 15 medium w/1-glutamine, w/o NaHCO₃, GIBCO, 1/1, v/v) (1/3, v/v) and streptomycin and gentamicine. Lymphoprep (density, 1.077, Nycomed Pharma As, Norway) (4/3, v/v) was added to mixed blood solution and then it was centrifuged at 350g for 20 min (10°C). The leukocytes were obtained from the interface and washed with AL medium. After centrifugation (600g for 10 min, 10°C), the leukocytes were suspended in AL medium containing 5.5 mM glucose. The viability and number of leukocytes were counted by adding trypan blue with a hemacytometer.

The Analysis of Leukocyte Phagocytosis by a Microtiter Plate Assay

The methods were modified from the procedures of Anderson and Mora (1995). About 1.5×10^5 leukocytes in 50 μ l AL medium with glucose were added in a microfluo black plate (Nunc-Immuno Modules, Roskilde, Denmark) in the presence or the absence of test compounds, and it was then incubated for several hours (4 h in tilapia, 8 h in common carp) at 25°C in air. After incubation at 25°C, 50 μ l of FITC-latex beads (0.105 μ m, Sigma) in 0.02 M phosphate-buffered saline solution (pH 7.0, Sigma) and 50 μ l of trypan blue quenching buffer (trypan blue in citrate buffer, pH 4.5, Sigma) were added and incubated at 25°C. The fluorescence remaining in the cultured medium was blocked by the trypan blue quenching buffer. The fluorescent intensity of the engulfed cells was measured by a MFX Microtiter Plate Fluorometer (Dynex, U.S.A.). The phagocytosis index was 100% in the control and the relative phagocytosis index in the treated group was calculated. The tested compounds (Sigma) were as follows: cortisol, cortisone, aldosterone, dexamethasone, testosterone, 11-ketotestosterone, progesterone, estradiol, estrone, ethynylestradiol, and triiodothyronine. Actinomycin D and cycloheximide (Sigma) were also used to test the blocking of the suppressive effects of steroids.

To obtain optimal experimental conditions, experimental conditions such as different ratios of latex beads per leukocyte (2, 3, 4, 5, and 6×10^4 beads/cell), the incubation time (from 5 to 180 min) of FITC-latex beads and leukocytes, and the quenching time (from 2 to 30 min) of trypan blue quenching buffer in the

mixture of latex beads/leukocytes were assessed, respectively. The optimal condition for the experiment was as follows: 5×10^4 latex beads/leukocyte, 40 min incubation between latex beads and leukocytes, and 10 min quenching with a trypan blue quenching buffer.

Statistical Analysis

One-way analysis of variance followed by Duncan's multiple range test was conducted to test the significant difference ($P < 0.05$) (Steel and Torrie, 1980). Results are given as a mean \pm standard error of the mean (SEM).

RESULTS

Phagocytosis in Tilapia: Dose and Time Dependency of the Suppressive Effect of Cortisol

Tilapia macrophage-engulfed FITC-latex beads appeared fluorescent green under fluorescent field microscopy (Fig. 1). Leukocyte viability, before and after the treatment with a tested compound, was more than 95% and not different. Various final concentrations (0, 0.01 pM, 1 pM, 0.1 nM, 10 nM, 1 μ M, and 0.1 mM) of cortisol were incubated with tilapia leukocytes for 0.5, 1, 2, 4, 6, and 8 h, respectively. The dose-dependent response and time course of the phagocytosis effects of cortisol are shown in the Fig. 2. Incubations for as little as 2 h with a high dose of 1 μ M cortisol or for 8 h with a low dose of 1 pM cortisol could significantly suppress phagocytosis. The maximal suppression of phagocytic activity (40% phagocytosis index) was observed with cortisol at 0.1 mM for 8 h incubation.

Comparative Effects of Various Corticosteroids on Phagocytosis in Tilapia

Four hours of incubation between tested chemicals and leukocytes was selected to investigate the phagocytic index in the presence of the various compounds. Both cortisol and the glucocorticoid agonist dexamethasone had a significant effect on the suppression of phagocytosis (Fig. 3). Dexamethasone is slightly more active than cortisol. Similar values and maximal ef-

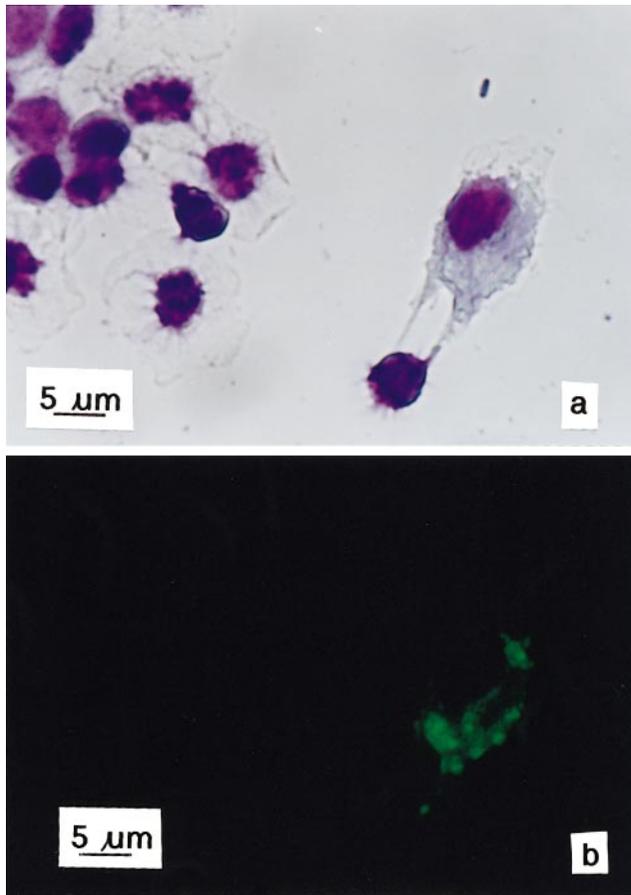


FIG. 1. Giesma stain of tilapia attached peripheral blood leukocytes (PBL). PBL were observed with bright-field (a) and fluorescent-field (b) microscopes. A macrophage engulfing FITC-latex beads was shown with green color for fluorescence.

fects were observed in 1 μM and 0.1 mM dexamethasone and 0.1 mM cortisol (Fig. 3). Cortisone and a mineralocorticoid, aldosterone, had very weak effects.

Comparative Effects of Various Sex Steroids and Thyroid Hormone on Phagocytosis in Tilapia

High doses (10 nM, 1 μM , and 0.1 mM) of estradiol and ethynylestradiol but not estrone significantly inhibited phagocytosis (Fig. 4a). All three compounds at a 10 nM dose, cortisol, estradiol and ethynylestradiol, started to show suppressive phagocytic activity (Figs. 3 and 4a). The phagocytic indices were 60, 82, and 80% in treatments with 0.1 mM cortisol, estradiol, and ethynylestradiol, respectively (Fig. 4a). The maximal suppressive action on phagocytosis of estrogenic com-

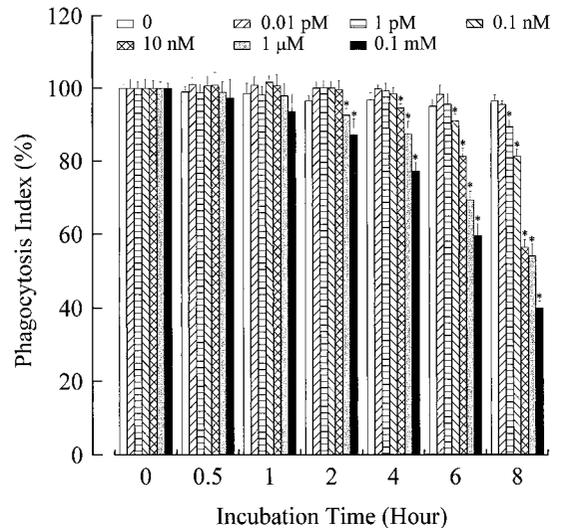


FIG. 2. The effects of cortisol (0–0.1 mM) and incubation time (0–8 h) on the phagocytosis index (%) in tilapia leukocytes. Values represent means \pm SEM. An asterisk indicates a significant difference ($P < 0.05$) from the respective control (without cortisol treatment).

pounds was detected at 1 μM and 0.1 mM ethynylestradiol (Fig. 4a). In contrast, there was no significant effect on phagocytosis with various concentrations of other gonadal steroids, progesterone, testosterone, and 11-ketotestosterone, nor of thyroid hormone, triiodothyronine (Fig. 4b).

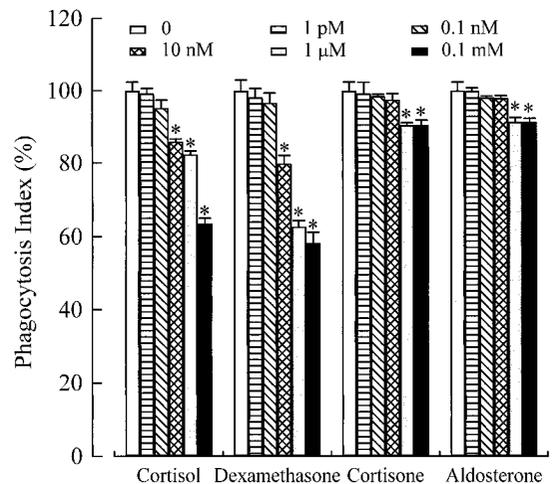


FIG. 3. The effects of cortisol, dexamethasone, cortisone, and aldosterone (0–0.1 mM for each chemical) on the phagocytosis index (%) in tilapia leukocytes. Values represent means \pm SEM. An asterisk indicates a significant difference ($P < 0.05$) from the respective control (without steroid or analog treatment).

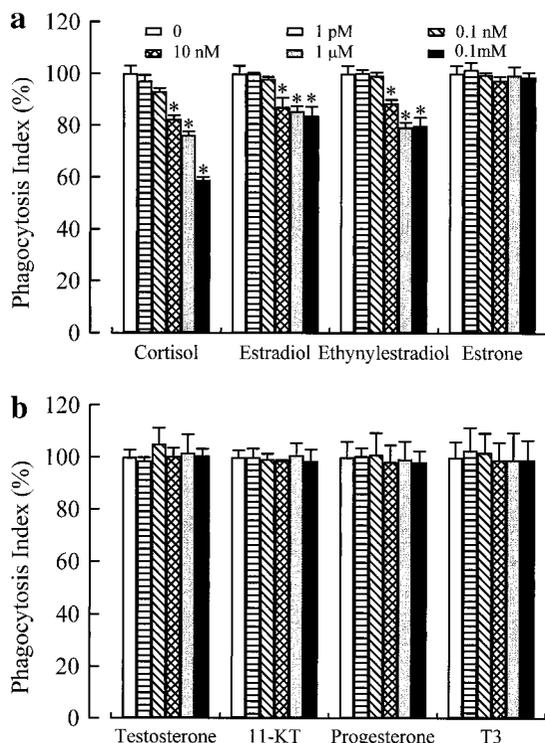


FIG. 4. The effects of sexual steroids and thyroid hormone (0–0.1 mM for each hormone) on the phagocytosis index (%) in tilapia leukocytes; (a) cortisol, estradiol, ethynylestradiol, and estrone; (b) testosterone, 11-ketotestosterone (11-KT), progesterone, and triiodothyronine (T₃). Values represent means \pm SEM. An asterisk indicates a significant difference ($P < 0.05$) from the respective control (without hormone or analog treatment).

Effects of Combined Treatments with Cortisol and Estradiol on Phagocytosis in Tilapia

The effects of combinations of different concentrations of estradiol and cortisol are shown in Fig. 5a. In the presence of an inactive dose of estradiol (0.1 nM), which is inactive alone, the suppressive action of cortisol on phagocytosis was enhanced. The maximal inhibitory effect corresponding to 60% is at a cortisol concentration of 1 μ M instead of 0.1 mM (Fig. 5a). Furthermore, in the presence of a low dose of estradiol (10 nM), the effective dose of cortisol was significantly reduced (10 nM vs 1 pM), and its maximal inhibitory effect was enhanced (up to 40% versus 60% of the phagocytosis index) (Fig. 5a). In comparison, no further suppression of phagocytosis was observed when testosterone was added to different concentrations of cortisol (Fig. 5b).

Effects of Actinomycin D and Cycloheximide on Cortisol or Estradiol Suppression of Phagocytosis in Tilapia

Whether the suppressive effects of cortisol and estradiol could be reduced by inhibitors of transcription and translation was investigated. As shown in Fig. 6, actinomycin D and cycloheximide blocked the suppressive effects of cortisol and estradiol in a dose-dependent manner (Fig. 6).

Leukocyte Phagocytosis in Common Carp: Dose and Time Dependence of the Suppressive Effect of Cortisol

Only high doses of cortisol (1 μ M or 0.1 mM) over a long incubation period (6 or 8 h) significantly inhibited leukocyte phagocytosis in common carp (Fig. 7). Comparison with tilapia showed that leukocyte

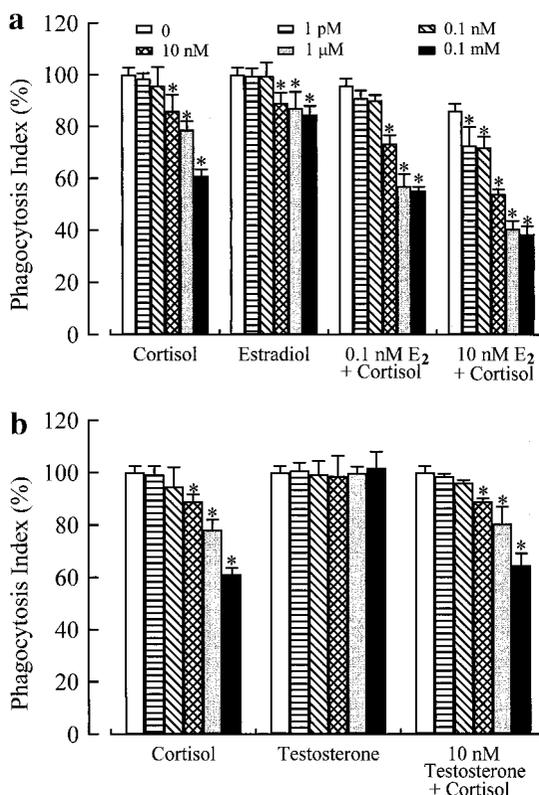


FIG. 5. Interaction of cortisol (0–0.1 mM) with (a) estradiol (E₂) (0.1 and 10 nM) and (b) testosterone (10 nM) on the phagocytosis index (%) in tilapia leukocytes. Values represent means \pm SEM. An asterisk indicates a significant difference ($P < 0.05$) from the respective control (without steroid treatment).

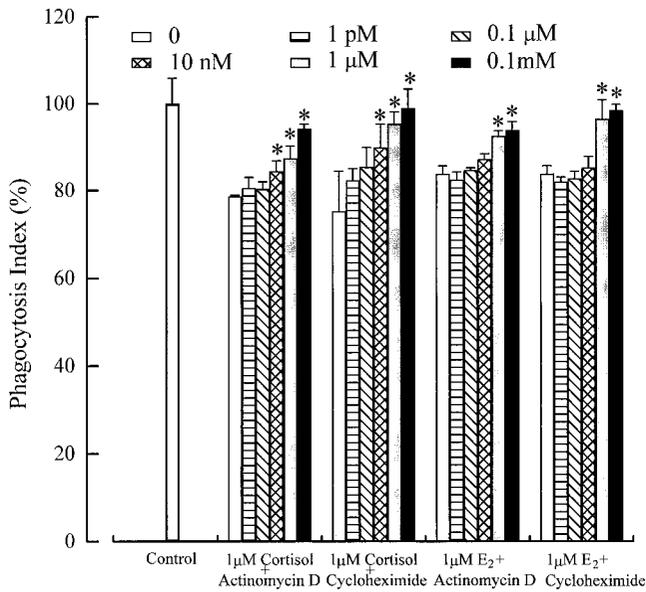


FIG. 6. Actinomycin D (0–0.1 mM) and cycloheximide (0–0.1 mM) inhibited cortisol (1 μ M) and estradiol (1 μ M) effects on phagocytosis in tilapia leukocytes. Values represent means \pm SEM. An asterisk indicates a significant difference ($P < 0.05$) from the respective control (without inhibitor treatment).

phagocytosis in common carp was much less sensitive to cortisol inhibition (Fig. 8). Therefore, further study of leukocyte phagocytosis in common carp was conducted for 8 h incubation.

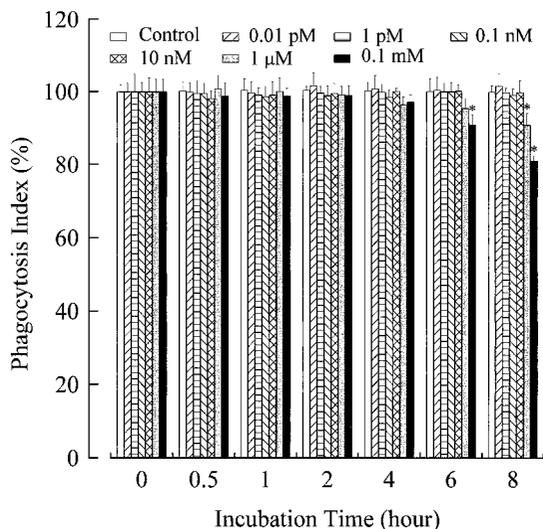


FIG. 7. The effects of cortisol (0–0.1 mM) and incubation time (0–8 h) on the phagocytosis index (%) in common carp leukocytes. Values represent means \pm SEM. An asterisk indicates a significant difference ($P < 0.05$) from the respective control (without cortisol treatment).

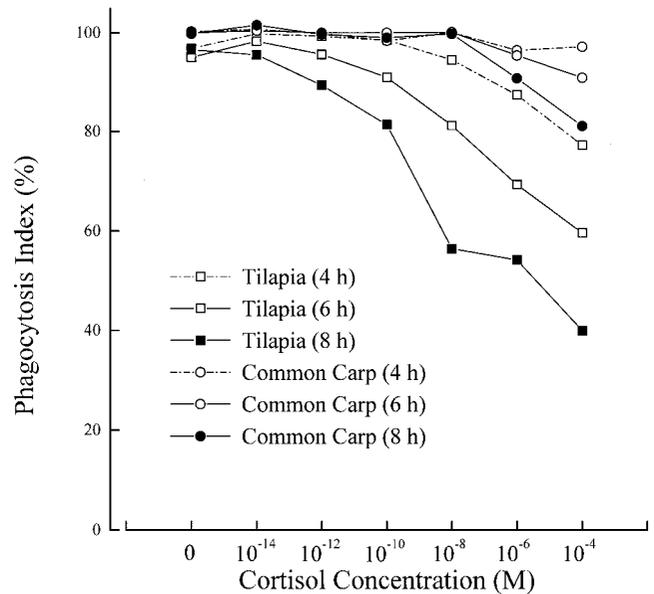


FIG. 8. Comparison of cortisol (0–0.1 mM) effects on the phagocytosis index (%) of leukocytes in tilapia and common carp. Incubation times of leukocytes and cortisol were 4, 6, and 8 h, respectively.

Lack of Gonadal Steroid Effect on Phagocytosis in Common Carp

There were no inhibitory actions of various doses (1 pm, 0.01 nM, 10 nM, 1 μ M, and 0.1 mM) of estradiol, testosterone, and 11-ketotestosterone on leukocyte phagocytosis (Fig. 9a). Estradiol also had no additive effects on the cortisol-inducing inhibition of phagocytosis (Fig. 9b).

DISCUSSION

Phagocytes have a major role in the cellular immune responses of fish, especially in fighting against bacteria. The factors that are involved in fish for the regulation of phagocytosis remain to be established. Peripheral blood provides a more consistent and easily handled source for the collection of leukocytes compared to the anterior kidney or spleen. The detected sensitivity, repeatability, and easy handling of engulfed fluorescent intensity in the whole well by the fluorescent photometer in the present experiment was much better than observations from microscopy (in-

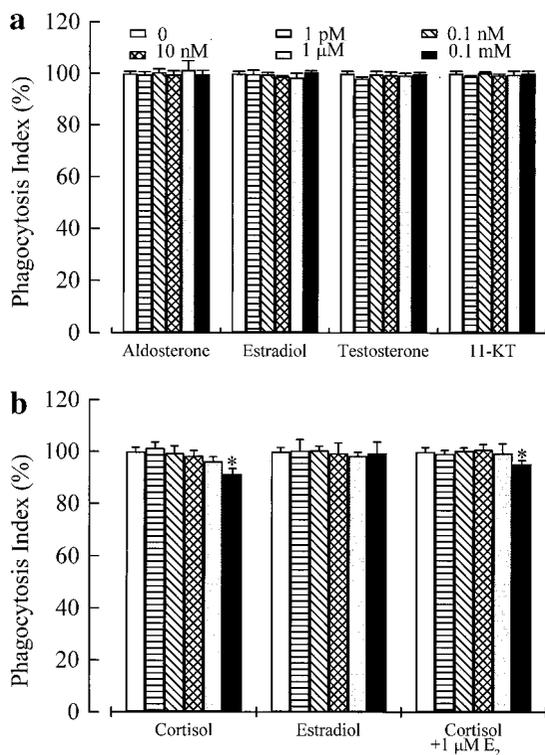


FIG. 9. The effects of (a) estradiol, testosterone, and 11-ketotestosterone (11-KT) (0–0.1 mM each steroid) and (b) interaction of cortisol (0–0.1 mM) and estradiol (E₂, 1 μM) on the phagocytosis index (%) in common carp leukocytes. Values represent means ± SEM. An asterisk indicates a significant difference ($P < 0.05$) from the respective control (without steroid treatment).

specting 200 leukocytes for calculation of the phagocytosis index) (Narnaware *et al.*, 1994, 1997). The FITC-latex beads are also better than yeast (Narnaware *et al.*, 1994; Narnaware and Baker, 1996) or sheep red blood cells (Carlson *et al.*, 1993; Calduch-Giner *et al.*, 1997) because of ease of handling, preserving, and selecting of smaller particle sizes. After evaluating the ratio of latex beads per leukocyte, incubation time of latex beads and leukocytes, and quenching time, it is considered that peripheral blood leukocytes could provide a valuable tool for the first time in teleosts to study phagocytosis in a microtiter plate assay with FITC-latex beads.

The present data demonstrated that cortisol significantly inhibited leukocyte phagocytosis in tilapia, as in some other fish and other vertebrates. The inhibition depends on dosage and incubation time. Plasma cortisol levels are significantly elevated and could be 10^{-7} M when animals are exposed to stressful environ-

ments and the elevated levels could last for several hours to days (Strange *et al.*, 1977; Leach and Taylor, 1980; Sun *et al.*, 1994). Therefore, cortisol levels in leukocyte phagocytosis experiment in the present study were from physiological (10^{-8} to 10^{-7} M) to pharmacological (10^{-6} and 10^{-4} M) doses. Inhibitory effects on phagocytosis were consistently observed at various doses of cortisol in this experiment. Cortisol also reduced phagocyte ability to generate a chemiluminescence response in striped bass when exposed to bacteria or phorbol esters (Stave and Robertson, 1985). A suppressive effect of cortisol on lipopolysaccharide mitogenesis was demonstrated in peripheral blood cells from Atlantic salmon *in vitro* (Espelid *et al.*, 1996). In contrast, cortisol at a low dose enhances phagocytosis in human (Kay and Czop, 1994) and bovine (Fox and Heald, 1981). No effect of cortisol on phagocytosis was found in anterior kidney and spleen of rainbow trout (Narnaware *et al.*, 1994). In the present study, the responsiveness of leukocytes from two different teleost species, tilapia and common carp, were compared by using the same experimental conditions. Tilapia was much more susceptible to cortisol effects than the common carp. There are clearly species differences in the regulation of leukocyte phagocytosis in fish.

There are a few studies of gonadal steroid effects of nonspecific immunity in animals. In the present studies, estradiol and the estradiol pharmacological agonist ethynylestradiol significantly suppressed leukocyte phagocytosis in tilapia but only at higher doses (10^{-8} , 10^{-6} , and 10^{-4} M). The amplitude of the suppressive effects was lower with estradiol than with cortisol. Other gonadal steroids, such as androgens (testosterone, 11-ketotestosterone) and progesterone, had no significant effects, indicating the estrogen specificity of the suppressive effects in tilapia.

In contrast to the suppressive effect of estradiol on phagocytosis in tilapia, gonadal steroids had no effects in common carp under similar experimental conditions. Inhibition by estradiol of nonspecific immunity occurs in some mammals, such as sow and porcine, but not in steers (Magnusson, 1991; Josefsson *et al.*, 1992). Testosterone can also suppress nonspecific immunity in chicken (Al-Afaeq and Homeida, 1998). Dihydrotestosterone strongly, but estradiol only weakly, inhibited splenic macrophage phagocytosis in wall lizards (Mondal and Rai, 1999). In teleosts, estra-

diol inhibits nonspecific immunity in goldfish (Wang and Belosevic, 1995). Five or more days of incubation with testosterone was required to elicit significant *in vitro* immunosuppression in anterior kidney leukocytes in chinook salmon (Slater and Schreck, 1997). Testosterone and cortisol administered *in vitro* together had a significantly greater inhibitory effect on antibody-producing cells than did either administered alone in chinook salmon (Slater and Schreck, 1993). It is apparent that gonadal steroid effects on nonspecific immunity vary according to the type of steroid and the species of teleost.

In the present study, cortisol and estradiol had a potentiating effect on the suppression of phagocytosis in tilapia. High estradiol levels are found in female fish, especially during oocyte development (vitellogenesis) (Fostier *et al.*, 1983). The data might suggest that reproduction (increased sex steroids) and stress (increased cortisol) might have a profound interaction on immunity in fish during the spawning season, as also indicated in brown trout, *S. trutta* (Pickering, 1984). This might explain the different resistance to disease infection between female and male fish and also seasonal effects. (Slater and Schreck, 1993). The present results, which showed no effect on leukocyte phagocytosis of the main steroid produced in male fish (aromatizable androgen, such as testosterone, nor nonaromatizable androgen, such as 11-ketotestosterone) may account for the lower susceptibility of males than females to disease infection in some species. The strong inhibitory effect on phagocytosis of the combination of cortisol and estradiol, as demonstrated in this study, may provide an endocrine bases for the high susceptibility to disease infection and the change of immune function at certain reproductive stages and/or in gender in human and animals (Roth *et al.*, 1983; Schuurs and Verheul, 1990; Grossman *et al.*, 1991; Mondal and Rai, 1999). It is male fish that appear to be more susceptible to disease during sexual maturation in rainbow trout (Sumpter *et al.*, 1987). Reduction in the magnitude of the cortisol level in plasma to stress was observed in the mature male compared with immature rainbow trout (Pottinger *et al.*, 1995). Sexual dimorphism in the phagocytic activity of the wall lizard's splenic macrophages was detected because of sex steroid effects of dihydrotestosterone in males (Mondal and Rai, 1999). The interaction between sex steroids and cortisol in the induction of

immunosuppression should be further studied *in vivo* and *in vitro* in more species.

The present data also suggest that specific glucocorticoid receptors and estrogen receptors are responsible for the involvement of cortisol and estradiol actions in tilapia. Glucocorticoid receptors have been identified in the anterior kidney leukocytes in coho salmon (Maule and Schreck, 1990). Further identification of glucocorticoid receptors and estrogen receptors in the peripheral blood leukocytes of tilapia is needed. Stimulation of gene expression and protein synthesis may mediate the action of cortisol and estradiol on leukocyte phagocytosis because actinomycin D and cycloheximide could block inhibitory actions. It has been shown that cortisol stimulated the production of a new protein ($I\kappa B\alpha$), and $I\kappa B\alpha$ hold NF- κB in an inactive form to prevent the synthesis of cytokines (mediators for immune system) (Marx, 1995; McKay and Cidlowski, 1999). It is possible that the pathway of estradiol action is similar to that of cortisol (McKay and Cidlowski, 1999). However, not all the ligands that bound to the nuclear receptors would automatically suppress phagocytosis because testosterone, 11-ketotestosterone, progesterone, and triiodothyronine did not have any effect in tilapia. Another consideration is whether specific receptors for these steroids are present in tilapia leukocytes.

It is also interesting that the common carp is much more resistant to the inhibitory effects of steroids on phagocytosis than tilapia. There is about a 10^5 - to 10^6 -fold difference between tilapia and common carp in terms of cortisol concentrations to suppress a similar degree of phagocytosis. Estradiol also did not have any effects on phagocytosis in common carp. The data may suggest that the common carp is more resistant than tilapia when exposed to environmental stressors (low temperatures) or disease infections (Sun *et al.*, 1994; Chen *et al.*, 1995). It is still unclear whether cyprinids in general are less sensitive to cortisol than other groups of fish (such as tilapia).

The affinity of glucocorticoid receptors is in the nanomolar range in the liver (K_d of ~ 0.5 nM) of rainbow trout, *O. mykiss* (Pottinger, 1990), leukocytes (K_d of ~ 1.0 nM) of coho salmon, *O. kisutch* (Maule and Schreck, 1990), and peripheral blood leukocytes (K_d of 3.8 nM) of common carp (Weyts *et al.*, 1998). Tilapia peripheral leukocytes were very sensitive to cortisol concentrations as low as 1 pM (after 8 h incubation) in

the present experiment. The detected binding affinity of receptors may not truly reflect the cortisol levels in physiological action. The binding affinity of the glucocorticoid receptor is apparently not different in fish. However, the present data demonstrated that carp leukocytes are less sensitive to steroids than are those of tilapia. The rationale for these differences between carp and tilapia is not clear. The binding sites of glucocorticoid receptors could be down-regulated by high levels of cortisol or stressors in rainbow trout (Pottinger, 1990). Any difference in the basal and stressed levels of plasma cortisol in tilapia and common carp may contribute to the different sensitivity to steroids of these two species. Further investigation also could address the question of whether the differential effects of steroids on phagocytosis between tilapia and common carp are associated with differential expression of specific steroid receptors in leukocytes.

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REFERENCES

- Al-Afaleq, A. I., and Homeida, A. M. (1998). Effects of low doses of estradiol, testosterone and dihydrotestosterone on the immune response of broiler chicks. *Immunopharmacol. Immunotoxicol.* **20**, 315–327.
- Anderson, R. S., and Mora, L. M. (1995). Phagocytosis: A microtiter plate assay. In "Techniques in Fish Immunology, Vol. 4: Immunology and Pathology of Aquatic Invertebrates" (J. S. Stolen, T. C. Fletcher, S. A. Smith, J. T. Zelikoff, S. L. Kaattari, R. S. Anderser, K. Soderhall, and B. A. Weeks-Perkins, Eds.), pp. 109–112. SOS, Fair Haven, NJ.
- Balm, P. H. M. (1997). Immune-endocrine interactions. In "Fish Stress and Health in Aquaculture" (G. K. Iwama, A. D. Pickering, and C. B. Schreck, Eds.), pp. 195–221. Cambridge Univ. Press, Cambridge.
- Berczi, I. (1997). Prolactin, growth hormone and host defence. In "Stress, Stress Hormones and the Immune System" (J. C. Buckingham, G. E. Gillies, and A. M. Cowell, Eds.), pp. 337–356. Wiley, New York.
- Blalock, J. E. (1994). The syntax of immune-neuroendocrine communication. *Immunol. Today* **15**, 504–511.
- Calduch-Giner, J. A., Sitja-Bobadilla, A., Alvarez-Pellitero, P., and Perez-Sanchez, J. (1995). Evidence for a direct action of GH on haemopoietic cells of a marine fish, the gilthead sea bream (*Sparus aurata*). *J. Endocrinol.* **146**, 459–467.
- Calduch-Giner, J. A., Sitja-Bobadilla, A., Alvarez-Pellitero, P., and Pérez-Sánchez, J. (1997). Growth hormone as an *in vitro* phagocyte-activating factor in the gilthead sea bream (*Sparus aurata*). *Cell Tissue Res.* **287**, 535–540.
- Carlson, R. E., Anderson, D. P., and Bodammer, J. E. (1993). *In vivo* cortisol administration suppresses the *in vitro* primary immune response of winter flounder lymphocytes. *Fish Shellfish Immunol.* **3**, 299–312.
- Chen, G. R., Sun, L. T., Lee, Y. H., and Chang, C. F. (1995). Characteristics of blood in common carp, *Cyprinus carpio*, exposed to low temperature. *J. Appl. Aquacult.* **5**, 21–31.
- Clark, R. (1997). The somatogenic hormone and insulin-like growth factor-I: Stimulators of lymphopoiesis and immune function. *Endocr. Rev.* **18**, 157–179.
- Ellis, A. E. (1981). Stress and the modulation of defense mechanisms in fish. In "Stress and Fish" (A. D. Pickering, Ed.), pp. 147–169. Academic Press, London.
- Ellsaesser, C. F., and Clem, L. W. (1987). Cortisol-induced hematologic and immunologic changes in channel catfish (*Ictalurus punctatus*). *Comp. Biochem. Physiol.* **87A**, 405–408.
- Espelid, S., Løkken, G. B., Steiro, K., and Bøgwald, J. (1996). Effects of cortisol and stress on the immune system in Atlantic salmon (*Salmo salar* L.). *Fish Shellfish Immunol.* **6**, 95–110.
- Fostier, A., Jalabert, B., Billard, R., Breton, B., and Zohar, Y. (1983). The gonadal steroids. In "Fish Physiology" (W. S. Hoar, D. J. Randall, and E. M. Donaldson, Eds.), Vol. IX, pp. 277–372. Academic Press, New York.
- Fox, L. K., and Heald, C. W. (1981). Effect of cortisol on the bactericidal function of the bovine milk neutrophil *in vitro*. *Am. J. Vet. Res.* **42**, 1933–1936.
- Goulding, N. J., and Flower, R. J. (1997). Glucocorticoids and the immune system. In "Stress, Stress Hormones and the Immune System" (J. C. Buckingham, G. E. Gillies, and A. M. Cowell, Eds.), pp. 337–356. Wiley, New York.
- Grossman, C. J. (1985). Interactions between the gonadal steroids and the immune system. *Science* **227**, 257.
- Grossman, C. J., Roselle, G. A., and Mendenhall, C. L. (1991). Sex steroid regulation of autoimmunity. *J. Steroid Biochem. Mol. Biol.* **40**, 646–659.
- Josefsson, E., Tarkowski, A., and Carlsten, H. (1992). Antiinflammatory properties of estrogen I. *In vitro* suppression of leukocyte production in bone marrow and redistribution of peripheral blood neutrophils. *Cell. Immunol.* **142**, 67–78.
- Kay, J., and Czop, J. K. (1994). Enhancement of human monocyte beta-glucan receptors by glucocorticoids. *Immunology* **811**, 96–102.
- Kincl, F. A., and Ciaccio, L. A. (1980). Suppression of immune responses by progesterone. *Endocrinol. Exp.* **14**, 27–33.
- Le Morvan, C., Clerton, P., Deschaux, P., and Troutaud, D. (1997). Effects of environmental temperature on macrophage activities in carp. *Fish Shellfish Immunol.* **7**, 209–212.

- Leach, G. J., and Taylor, M. H. (1980). The role of cortisol in stress-induced metabolic changes in *Fundulus heteroclitus*. *Gen. Comp. Endocrinol.* **42**, 219–227.
- Magnusson, U. (1991). *In vitro* effects of prepartum concentrations of oestradiol-17 β on cell-mediated immunity and phagocytosis by porcine leukocytes. *Vet. Immunol. Immunopathol.* **28**, 117–126.
- Magnusson, U., and Einarsson, S. (1990). Effects of exogenous oestradiol on the number and functional capacity of circulating mononuclear and polymorphonuclear leukocytes in the sow. *Vet. Immunol. Immunopathol.* **25**, 235–247.
- Marx, J. (1995). How the glucocorticoid suppress immunity. *Science* **270**, 232–233.
- Maule, A. G., and Schreck, C. B. (1990). Glucocorticoid receptors in leukocytes and gill of juvenile coho salmon (*Oncorhynchus kisutch*). *Gen. Comp. Endocrinol.* **77**, 448–455.
- Mazur, C. F., and Iwama, G. K. (1993). Handling and crowding stress reduces number of plaque-forming cells in Atlantic salmon. *J. Aquatic Anim. Health* **5**, 98–101.
- McKay, L. I., and Cidlowski, J. A. (1999). Molecular control of immune/inflammatory responses: Interactions between nuclear factor- κ B and steroid receptor-signaling pathways. *Endocr. Rev.* **20**, 435–459.
- Miller, N. W., Chinchar, V. G., and Clem, L. W. (1994). Development of leukocyte cell lines from the channel catfish (*Ictalurus punctatus*). *J. Tissue Cult. Methods* **16**, 117–123.
- Mondal, S., and Rai, U. (1999). Sexual dimorphism in phagocytic activity of wall lizard's splenic macrophages and its control by sex steroids. *Gen. Comp. Endocrinol.* **116**, 291–298.
- Nagal, M., Fuda, H., Jara, A., Sazneyoshi, M., and Yamauchi, K. (1994). Changes in serum concentrations of immunoglobulin M (IgM), cortisol and thyroxine (T₄) during smoltification in the masu salmon, *Oncorhynchus masou*. *Fish. Sci.* **60**, 241–242.
- Narnaware, Y. K., Baker, B. I., and Tomlinson, M. G. (1994). The effects of various stresses, corticosteroids and adrenergic agents on phagocytosis in the rainbow trout *Oncorhynchus mykiss*. *Fish Physiol. Biochem.* **13**, 31–40.
- Narnaware, Y. K., and Baker, B. I. (1996). Evidence that cortisol may protect against the immediate effects of stress on circulating leukocytes in the trout. *Gen. Comp. Endocrinol.* **103**, 359–366.
- Narnaware, Y. K., Kelley, S. P., and Woo, N. Y. S. (1997). Effect of injected growth hormone on phagocytosis in silver sea bream (*Sparus sarba*) adapted to hyper- and hypo-osmotic salinities. *Fish Shellfish Immunol.* **7**, 515–517.
- Pickering, A. D. (1984). Cortisol-induced lymphocytopenia in brown trout, *Salmo trutta* L. *Gen. Comp. Endocrinol.* **53**, 252–259.
- Pickering, A. D., and Duston, J. (1983). Administration of cortisol to brown trout, *Salmo trutta* L., and susceptibility to saprolegnia infection and furunculosis. *J. Fish Biol.* **23**, 163–175.
- Pottinger, T. G. (1990). The effect of stress and exogenous cortisol on receptor-like binding of cortisol in the liver of rainbow trout, *Oncorhynchus mykiss*. *Gen. Comp. Endocrinol.* **78**, 194–203.
- Pottinger, T. G., Balm, P. H. M., and Pickering, A. D. (1995). Sexual maturity modifies the responsiveness of the pituitary-interrenal axis to stress in male rainbow trout. *Gen. Comp. Endocrinol.* **98**, 311–320.
- Roth, J. A., Kaeberle, M. L., Appell, L. H., and Nachreiner, R. F. (1983). Association of increased estradiol and progesterone blood values with altered bovine polymorphonuclear leukocyte function. *Am. J. Vet. Res.* **44**, 247–253.
- Roth, J. A., Kaeberle, M. L., and Hsu, W. H. (1982). Effect of estradiol and progesterone on lymphocyte and neutrophil functions in steers. *Infect. Immunol.* **35**, 997–1002.
- Sapolsky, R. M., Romero, L. M., and Munck, A. U. (2000). How do glucocorticoids influence stress responses? Integrating permissive, suppressive, stimulatory, and preparative actions. *Endocr. Rev.* **21**, 55–89.
- Schuurs, A. H., and Verheul, H. A. (1990). Effects of gender and sex steroids on the immune response. *J. Steroid Biochem.* **35**, 157–172.
- Slater, C. H., and Schreck, C. B. (1993). Testosterone alters the immune response of chinook salmon, *Oncorhynchus tshawytscha*. *Gen. Comp. Endocrinol.* **89**, 291–298.
- Slater, C. H., and Schreck, C. B. (1997). Physiological levels of testosterone kill salmonid leukocytes *in vitro*. *Gen. Comp. Endocrinol.* **106**, 113–119.
- Stave, J. W., and Robertson, B. S. (1985). Cortisol suppresses the chemiluminescent response of striped bass phagocytes. *Dev. Comp. Immunol.* **9**, 77–84.
- Steel, R. G. D., and Torrie, J. Y. H. (1980). "Principles and Procedures of Statistics." McGraw-Hill, New York.
- Strange, R. J., Schreck, C. B., and Golden, J. T. (1977). Cortisol stress response to handling and temperature in salmonids. *Trans. Am. Fish. Soc.* **106**, 213–218.
- Sumpter, J. P., Carragher, J., Pottinger, T. G., and Pickering, A. D. (1987). The interaction of stress and reproduction in trout. In "Reproductive Physiology of Fish" (D. R. Idler, L. W. Crim, and J. M. Walsh, Eds.), pp. 299–302. Memorial Univ. Press, St John's, Newfoundland, Canada.
- Sun, L. T., Chen, G. R., and Chang, C. F. (1994). Characteristics of blood parameters and gill Na⁺-K⁺-ATPase in chilled comatose tilapia cultured in various salinities. *Comp. Biochem. Physiol.* **107A**, 641–646.
- Wang, R., and Belosevic, M. (1995). The *in vitro* effects of estradiol and cortisol on the function of a long-term goldfish macrophage cell line. *Dev. Comp. Immunol.* **19**, 327–336.
- Weyts, F. A. A., Flik, G., and Verburg-van Kemenade, B. M. L. (1998). Cortisol inhibits apoptosis in carp neutrophilic granulocytes. *Dev. Comp. Immunol.* **22**, 563–572.
- Weyts, F. A. A., Verburg-van Kemenade, B. M. L., and Flik, G. (1998). Characterization of glucocorticoid receptors in peripheral blood leukocytes of carp, *Cyprinus carpio* L. *Gen. Comp. Endocrinol.* **111**, 1–8.
- Yada, T. Y., Nagae, M., Moriyama, S., and Azuma, T. (1999). Effects of prolactin and growth hormone on plasma immunoglobulin M levels of hypophysectomized rainbow trout, *Oncorhynchus mykiss*. *Gen. Comp. Endocrinol.* **15**, 46–52.