

**PREPARATION OF 2,3,6,3',4'-PENTA-O-ACETYL SUCROSE,  
THE PRECURSOR OF SUCRALOSE, BY ENZYMATIC METHODS**

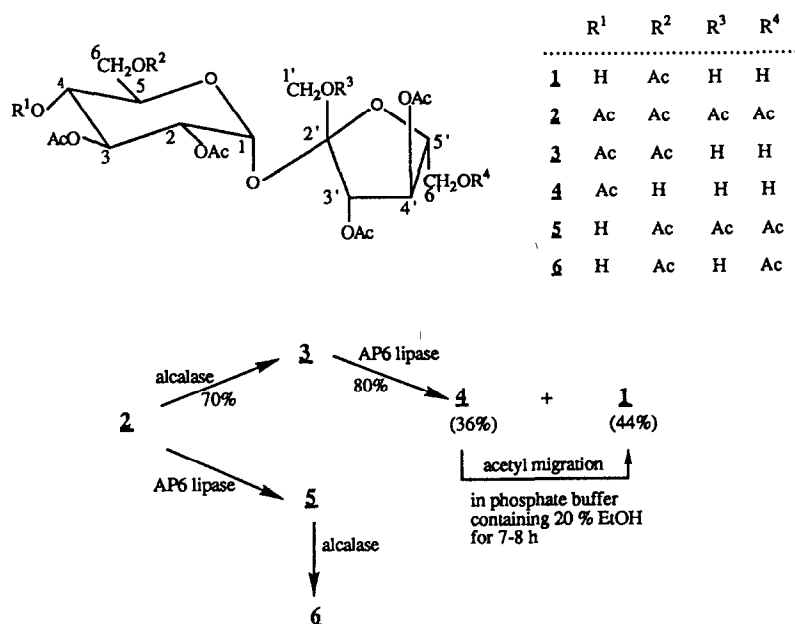
Geok-Toh Ong, Shih-Hsiung Wu\* and Kung-Tsung Wang  
Institute of Biological Chemistry, Academia Sinica and  
Graduate Institute of Biochemical Sciences,  
National Taiwan University  
P.O.Box. 23-106, Taipei, Taiwan

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**Summary :** 2,3,6,3',4'-penta-O-acetyl sucrose, the precursor of sucralose, can be prepared from the sequential hydrolysis of sucrose octaacetate by alcalase and AP-6 lipase.

In 1976, Hough and Phadnis reported the synthesis of 4-chloro-4-deoxy- $\alpha$ -D-galactopyranosyl 1,6-dichloro-1,6-dideoxy- $\beta$ -D-fructofuranoside, called sucralose, and found it to possess sweetness several hundred times that of sucrose<sup>1,2</sup>. Because of non-nutrition, non-carcinogen and resistance to be hydrolyzed by  $\alpha$ -galactosidase and  $\beta$ -fructofuranosidase, the compound is quite safe for human use. The compound had been synthesized from 2,3,6,3',4'-pentaacetyl sucrose (1) by replacing hydroxyl groups with chlorine, especially the inversion of chirality at C-4 (gluco ---> galacto)<sup>3</sup>. Compound 1 was obtained from sucrose which was tritylated, acetylated, then detritylated and C<sub>4</sub> ---> C<sub>6</sub> acetyl migration<sup>4,5</sup>, but the whole procedure was tedious and inefficient. A strategy that has more applicability for the production of 1 is to selectively remove the acetyl groups from sucrose octaacetate (2). Selective removal of acetyl groups from 2 for the preparation of partially acetylated sucrose has been studied in chemical<sup>6-13</sup> and enzymatic<sup>14-16</sup> ways. Enzymatic methods

are generally more selective than chemical methods. Recently, a British group has been developed a enzymatic method for preparing **1** directly by the subtilisin-catalyzed hydrolysis of **2**<sup>17</sup>. However, the yield was extremely low (about 1.6 %) and quite impractical. Here, we report that **2** prepared from the sequential hydrolysis of **1** by alcalase and lipase AP-6 with 55% of a total yield was shown in Scheme 1.



Scheme 1.

In order to prepare **1** efficiently and practically, compound **2** was first hydrolyzed by alcalase (subtilisin Carlsberg from Novo, Denmark) to produce 2,3,4,6,3',4'-hexa-Q-acetyl sucrose (**3**) and then **3** was further hydrolyzed by AP-6 lipase (*Aspergillus*

nigar from Amano, Japan) to form 2,3,4,3',4'-penta-Q-acetyl sucrose (4) and 1. Compound 4 could be converted to 1 by C<sub>4</sub> ---  
-> C<sub>6</sub> acetyl migration in the phosphate buffer (0.2 M, pH 7.0) containing 20 % ethanol. The structure of 1 and 4 were determined by <sup>1</sup>H-<sup>1</sup>H and <sup>1</sup>H-<sup>13</sup>C 2D-COSY NMR spectra<sup>18</sup>. However, if compound 2 was first hydrolyzed by AP-6 lipase to produce 2,3,6,1',3',4',6'-hepta-Q-acetyl sucrose (5) and then further hydrolyzed by alcalase, the major product was found to be 2,3,6,3',4',6'-hexa-Q-acetyl sucrose (6) with little sucrose penta-acetate. It evidently showed that sucrose hexaacetates were not good substrates for subtilisin. That is reason why 2 could not be obtained directly from the subtilisin-catalyzed hydrolysis of 1 with substantial yield<sup>17</sup>. The method we developed has much higher yield and more practical than that of the British group.

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  18. N.m.r. data of **1** (acetonitrile- $d_3$ ):  $^1\text{H}$ ,  $\delta$  1.97-2.10 (m, 15 H, 5 Ac), 3.20-3.30 (t, 1 H, OH-6'), 3.30-3.45 (m, 2 H, OH-1', 4), 3.50 (d, 1 H, H-1'), 3.52-3.72 (m, 3 H, H-4, 6', 6'), 3.90-4.12 (m, 2 H, H-5, 5'), 4.18-4.38 (m, 2 H, H-6, 6), 4.65-4.75 (dd, 1 H,  $\underline{J}_{1,2}$  3.7,  $\underline{J}_{2,3}$  10.3 Hz, H-2), 5.15-5.28 (dd, 1 H,  $\underline{J}_{3,4}$  9.7 Hz, H-3), 5.30-5.42 (dd, 1 H,  $\underline{J}_{3,4}$  =  $\underline{J}_{4,5}$  = 7.8 Hz, H-4'), 5.52 (d, 1 H, H-3'), 5.55 (d, 1 H, H-1');  $^{13}\text{C}$ ,  $\delta$  20.40-20.52 (5 C, -COCH<sub>3</sub>), 62.12 (C<sub>6</sub>), 63.10 (2C, C<sub>6,1'</sub>), 68.41 (C<sub>4</sub>), 70.70 (C<sub>2</sub>), 71.03 (C<sub>5</sub>), 72.10 (C<sub>3</sub>), 74.77 (C<sub>4</sub>), 75.98 (C<sub>3</sub>), 81.71 (C<sub>5</sub>), 90.26 (C<sub>1</sub>), 105.19 (C<sub>2</sub>), 170.28-171.77 (5 C, -COCH<sub>3</sub>).
  - N.m.r. data of **4** (CDCl<sub>3</sub>):  $^1\text{H}$ ,  $\delta$  1.97-2.15 (m, 15 H, 5 Ac), 3.18 (t, 1 H, OH-6'), 3.37-3.80 (m, 6 H, H-1', 1', 6, 6, 6', 6'), 3.95-4.00 (m, 1 H, H-5'), 4.07-4.14 (m, 1 H, H-5), 4.76-4.83 (dd, 1 H,  $\underline{J}_{1,2}$  3.7,  $\underline{J}_{2,3}$  10.3 Hz, H-2), 4.85-4.96 (dd, 1 H,  $\underline{J}_{3,4}$  =  $\underline{J}_{4,5}$  = 7.8 Hz, H-4'), 5.34-5.47 (m, 3 H, H-3, 3', 4'), 5.64 (d, 1 H, H-1);  $^{13}\text{C}$ ,  $\delta$  20.71-20.81 (5 C, -COCH<sub>3</sub>), 62.89 (C<sub>6</sub>), 63.19 (C<sub>1</sub>), 63.99 (C<sub>6</sub>), 68.89 (C<sub>4</sub>), 70.30 (C<sub>2</sub>), 70.84 (C<sub>5</sub>), 71.96 (C<sub>3</sub>), 74.90 (C<sub>4</sub>), 76.50 (C<sub>3</sub>), 78.80 (C<sub>5</sub>), 90.15 (C<sub>1</sub>), 105.34 (C<sub>2</sub>), 170.21-171.56 (5 C, -COCH<sub>3</sub>).