

行政院國家科學委員會專題研究計畫 成果報告

狼瘡病人血清中抗中性白血球抗體的免疫活性, 對應抗原的
鑑定, 及臨床應用(3/3)

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中文摘要：

關鍵詞：白血球減少，抗中性白血球抗體，全身性紅斑狼瘡，SSB/La 蛋白，細胞凋亡，活化誘導細胞死亡，死亡訊息

白血球減少在全身性紅斑狼瘡病人(SLE)是一種常見的血液學異常。文獻報告指出約有 50-60%的 SLE 病人的血清中有抗中性白血球抗體 (anti-PMN autoantibodies) 的存在，而且與狼瘡的活動性有關。我們利用 PMN 的 cellular ELISA 測定法發現 SLE-sera 的抗白血球活性遠比正常血清為高。而且這個抗 PMN 活性會誘發白血球的細胞凋亡。另外，本抗體也會促進正常 PMN 的吞食能力及產生更多量的 IL-8。這些結果使我們相信 SLE sera 中的抗 PMN 抗體會經由 activation-induced cell death (AICD)來傷害 PMN 並引發細胞凋亡。為了更進一步鑑定 anti-PMN 抗體在 PMN 細胞膜上對應抗原的存在及其特性，我們將 biotinylated PMN 製成 cell lysate 後，先以 normal serum 或 SLE serum 作免疫沉澱之後，再經 10% SDS-PAGE 電泳分離，最後以 HRP-avidin 作探針及呈色反應。我們發現 47-50kDa 的 PMN 膜上分子會成為 SLE 血清中 anti-PMN 抗體的對應抗原。經蛋白質體學(Proteomics)的分析，鑑定出這個 50 kDa 左右的膜蛋白的氨基酸序列有 99.55%與 SSB/La (一種 RNA polymerase III binding protein) 相符。為了大量製造純化的 SSB/La 來作進一步的研究，我們首先利用人類由來的白血球 cDNA library，以含有 anti-SSB/La 抗體的 SLE sera 來篩選會轉錄 SSB/La protein 的 DNA。並使此 DNA 增殖之後，轉殖進入 E-coli 使之大量製造混成 SSB/La，再以 His-tag column 大量純化蛋白質。最後再以 SSB/La-Sepharose affinity column 由富含 anti-SSB/La 抗體的 SLE 病人血清加以純化。相當有趣的是這些純化的 anti-SSB/La 抗體及 SSB/La 分子兩者對 PMN 均具有相同的活性。即是有刺激活化 PMN 的功能導至 PMN apoptosis。但當兩種分子共同添加於懸浮液時並無加成作用。這些結果顯示 SSB/La 分子可能扮演對 innate immune system 有 danger signal 的角色。當細胞因任何原因受到傷害時，細胞核內的 SSB/La 會流出細胞外，這些扮演 danger signal 的分子會活化吞食細胞以發揮清除死細胞及消除引起細胞死亡的病因。這個假設仍需更多的實驗來證實。

ABSTRACT

Keywords: leucopenia, anti-neutrophil antibodies, systemic lupus erythematosus, SSB/La protein, apoptosis, activation-induced cell death, death signal

Leukopenia is a quite common hematological abnormality found in patients with systemic lupus erythematosus (SLE). It had been reported that the prevalence rate of anti-polymorphonuclear neutrophil antibodies (anti-PMN) is found in 50-60% of SLE sera in parallel with lupus activity. By using cellular ELISA of normal PMN as substrate, we demonstrated that the anti-PMN activity in SLE sera is significantly higher than normal sera. In addition, the anti-PMN activity of SLE-sera may elicit PMN apoptosis as detected by flow cytometry. Furthermore, the binding of SLE anti-PMN autoantibodies enhanced phagocytic activity and IL-8 production by normal PMN. These results render us to conclude that neutropenia in SLE is caused by the presence of anti-PMN in the sera that cytotoxic to PMN via the mechanism of activation induced cell death (AICD). To identify the cognate antigen(s) of anti-PMN autoantibodies on the PMN surface, Western blot analysis of biotinylated normal PMN lysates previously immunoprecipitated by either normal or SLE sera and were then probed by HRP-avidin and. We found that the 47-50kDa membrane molecule was the cognate antigen of SLE anti-PMN. Proteomics study revealed the 50kDa protein was 99.5% homologous with human nuclear SSB/La molecule. Molecular cloning of the gene-encoding SSB/La was obtained from human derived leukocyte cDNA library. After insertion, proliferation, induction and expression of the SSB/La encoding gene, the expressed protein was further purified by His-tag column. Using affinity column of SSB/La-Sepharose column, human polyclonal anti-SSB/La autoantibodies were partially purified from SLE sera containing high anti-SSB/La activity. The anti-SSB/La autoantibodies were proved to be

cytotoxic to normal human PMN. Interestingly, both anti-SSB/La auto-antibodies and SSB/La protein exhibited PMN activation effects including phagocytosis and IL-8 production as mentioned in the above paragraph. However, the addition of SSB/La and anti-SSB/La antibodies did not increase these activation. These results suggest that SSB/La protein may probably act as a “danger signal” to the innate immune system in that release of SSB/La molecules from the injured cell alert the phagocytes to activation. However, further investigations are needed to clarify this observation.

Anti-SSB/La is one of the antineutrophil autoantibodies responsible for neutropenia and functional impairment of polymorphonuclear neutrophils in patients with systemic lupus erythematosus

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Summary

Decreased number and impaired functions of polymorphonuclear neutrophils (PMN) due to the presence of anti-PMN autoantibodies in the serum render patients with systemic lupus erythematosus (SLE) susceptible to bacterial infections. However, the cognate antigens and pathological mechanisms of anti-PMN autoantibodies in SLE are rarely reported in the literature. In this study, we found approximately 20% of SLE sera contained anti-PMN autoantibodies detected by human PMN-coated cellular ELISA. A membrane protein with molecular weight of 50 kDa was identified as the cognate antigen of anti-PMN in Western blot after membrane-biotinylation and streptavidin column elution. The 50 kDa molecule was proved to be SSB/La after immunoscreening, molecular cloning and nucleotide sequencing of the gene from the human leucocyte cDNA library. Human anti-SSB/La autoantibodies purified from active SLE sera passing through the recombinant SSB/La conjugated Sepharose 4B affinity column could bind and penetrate into normal human PMN. Functional analysis revealed that the anti-SSB/La autoantibodies exerted a number of potent effects on human PMN, including suppressed phagocytosis, accelerated apoptosis and enhanced IL-8 production. These *in vitro* results suggest that anti-SSB/La is one of the anti-PMN autoantibodies capable of penetrating into PMN and responsible for neutropenia and functional impairment of PMN in patients with SLE.

Introduction

Systemic lupus erythematosus (SLE), the archetype of systemic autoimmune disorder, is characterized by the presence of a diverse spectrum of autoantibodies in the serum. Some of the autoantibodies bind to circulating antigens forming immune complexes and deposit in different tissues to elicit tissue inflammation [1,2]. Some of the autoantibodies target the surface expressed antigens on the tissues directly, causing cytotoxicity [3,4]. Neutropenia is found in about 50–60% of patients with active SLE in the long-term course of the disease [5]. Polymorphonuclear neutrophils (PMN) are regarded traditionally as the first-line cell component of the body defence mechanisms against bacterial pathogens. However, recent investigations demonstrated that PMN play a role in the afferent and efferent limbs of immune responses that react to and produce different cytokines, including IL-1, IL-6, IL-10, IL-12, TNF- α , TGF- β ₁, G-CSF, M-CSF, GM-CSF and chemokines such as IL-8, MIP-1 α , MIP-1 β and MCP-1 [6,7]. Clinically, increased susceptibility to infections is a major cause of morbidity and mortality in patients with SLE [8,9]. Treatment with adrenal corticosteroids and/or immunosuppressants is obviously responsible for the increased infections. However, decreased numbers [10] and/or impaired functions [11–14] of PMN *per se* are equally crucial for susceptible to bacterial infection in SLE. The documented functional defects of SLE-PMN included reduced phagocytosis [11], presence of a serum inhibitor for phagocytosis [12,13], decreased nitroblue tetrazolium dye reduction [14] and impaired cytokine/chemokine production

[15]. However, the real causes of the neutropenia and functional defects in SLE-PMN have not yet been entirely elucidated.

Drew and Terasaki [16] reported that autoimmune cytotoxic antigranulocyte antibodies were found in approximately 50% of SLE patients detected by a panel of 70 different granulocytes from random normal individuals. Lalezari *et al.* [17] and Jiang *et al.* [18] demonstrated that granulocytes possess a set of alloantigenic specificities encoded by separate genes, loci NA, NB and NC, other than HLA antigens when detected by leuco-agglutination assays. Starkebaum *et al.* [19] found that increased IgG binding on the surface of PMN was the major cause of neutropenia in SLE. These bound monomeric IgG anti-PMN autoantibodies facilitated the opsonization of PMN engulfed by mononuclear phagocytes. The F(ab')₂ fragment of the IgG molecules was confirmed as the true antibody in nature [20]. Furthermore, Hardley *et al.* [21] noted that the PMN-bound antibodies from SLE were capable of penetrating into cytosol and nuclei. Reports also demonstrated that SLE sera were able to opsonize PMN and fix C3 that correlated inversely with neutrophil counts in SLE [22,23]. The binding specificity of anti-PMN antibodies in SLE was investigated further by Sipos *et al.* [24], who demonstrated that the antibodies reacted with two membrane antigens on PMN with molecular weights of 50–60 kDa and 30 kDa. Functional analysis revealed that the anti-PMN autoantibodies inhibited the binding of mouse monoclonal antibodies with CD15 (granulocyte antigen, 46 kDa) and CD16 (Fc γ RIIIb, 50–80 kDa) on normal human neutrophils. However, whether the cognate antigens of anti-PMN in SLE serum are identical to CD15 or CD16 were not confirmed in that study.

To explore the nature of the cognate antigen(s) and immunopathological roles of anti-PMN in SLE, immunoscreening, molecular cloning and *Escherichia coli* expression of the gene(s) encoding the anti-PMN cognate antigens from human leucocyte λ -phage expression library was conducted. We found that 50 kDa SSB/La is one of the cognate autoantigens expressed on human PMN surface. Anti-SSB/La autoantibodies purified from SLE sera could penetrate into PMN after binding with the PMN surface responsible for increased apoptosis and functional impairments of normal PMN *in vitro*. These results suggest that SSB/La autoantibody is one of the antineutrophil autoantibodies responsible for the neutropenia and functional defects of PMN *in vivo* in patients with SLE.

Materials and methods

Patients and controls

The sera were collected from normal individuals and the stocked SLE sera containing anti-dsDNA antibodies titre \geq 100 IU/ml after routine serological measurements in a clinical immunological laboratory. These SLE sera were expected to be discarded after 3–6 months storage. The normal sera were confirmed negative for antinuclear antibody test and normal haemogram.

Preparation of polymorphonuclear neutrophils from normal human peripheral blood

Heparinized venous blood obtained from healthy volunteers was mixed with one-fourth volume of 2% dextran solution (mol. wt. 464 000) (Sigma Chemical Company, St Louis, MO, USA) and sedimented at room temperature for 30 min. Leucocyte-enriched supernatant was collected and layered over Ficoll-Hypaque (specific gravity 1.077) (Pharmacia Biotech, Uppsala, Sweden) density gradient solution. After centrifugation at 300 *g* for 25 min, PMN were obtained from the bottom. In some experiments, mononuclear cells (MNC) were aspirated concomitantly from the interface after centrifugation. The contaminating red blood cells in PMN suspension were lysed by suspending in chilled 0.83% ammonium chloride solution at 4°C for 10 min. Cells concentration was adjusted to 5×10^6 /ml in 10% fetal bovine serum (FBS) in RPMI-1640 (10% FBS-RPMI). Both purity and viability of PMN and MNC were greater than 95%, as confirmed by Wright's stain and trypan blue dye exclusion, respectively.

Preparation of surrogate immune complexes

The method reported by Gamberale *et al.* [25] was adapted. Briefly, human γ -globulin (Sigma) was heat-aggregated at 63°C for 1 h at a concentration of 5 mg/ml. The aggregated human IgG (aggIgG) was centrifuged at 10 000 *g* for 10 min and the precipitates were discarded. The collected supernatant was stored at 4°C as surrogate immune complex. In all the following experiments (PMN-binding, phagocytosis, PMN penetration and IL-8 production), freshly prepared PMN were usually incubated with aggIgG (50 μ g/ml) at 37°C for 1 h in advance for ligating the Fc γ receptors on the surface of PMN. After two washes with PBS, pH 7.2, the aggIgG-treated PMN were used for the following experiments. Fc γ R ligation by aggIgG (50 μ g/ml) did not affect the viability (MTT test) and function (phagocytosis) of PMN (data not shown). In addition, ligation by aggIgG and then blockade by specific anti-Fc γ RIII antibody (anti-CD16) of Fc γ R on PMN did not derange the phagocytosis (data not shown).

Detection of PMN-binding capacity of different sera by cellular ELISA

The method of Doughty *et al.* [26] was used for detecting the amount of neutrophil-binding immunoglobulins in the sera. Briefly, 50 μ l of aggIgG-pretreated neutrophil suspension (5×10^6 cells/ml) were placed in 96-well U-shaped flat-bottomed microtitre plates (Linbro/Titertek[®], Flow Laboratories, Inc., McLean, VA, USA). The microtitre plate was then centrifuged at 150 *g* for 10 min to precipitate the cells at the bottom. After removing the supernatant carefully, the neutrophils were fixed by 0.24% glutaraldehyde solution at 4°C for 30 min followed by incubation with 1% bovine serum albumin (Sigma) for 2 h to reduce the non-specific binding with protein molecules. One hundred microlitres of SLE or control sera by 1 : 50 dilution was added to the wells and incubated for 1 h at room temperature. After several washes with PBS, pH 7.2, 100 μ l of 1 : 5000 diluted HRP-conjugated goat antihuman IgG (Chemicon International Inc., Temecula, CA, USA) was added and incubated for 1 h at room temperature followed by reacting with 150 μ l of substrate containing ortho-phenilen diamine (Sigma) for 10 min in the dark. The reaction was stopped by the addition of 50 μ l 4 N H₂SO₄ solution. The amount of antibodies bound to PMN was measured by an ELISA reader (Dynex Technologist, Chantilly, VA, USA) at O.D._{490nm} absorbance. We defined anti-PMN(+) SLE sera as those exhibited PMN-binding capacity (O.D._{490nm}) \geq mean + 2 s.d. of normal sera. Only these anti-PMN(+) SLE sera were used in the following experiments. Because at least two confounding factors, hypergammaglobulinaemia and immune complexes, existed in SLE sera, the binding of SLE sera with PMN could be enhanced by false positivity. However, after 50 \times dilution of sera and previous ligation of IgGf_cR on PMN by heat-aggregated human IgG, these two confounding factors in SLE sera could be minimized. The anti-PMN(+) SLE serum was adequate for the detection of cognate antigen(s) and immunoscreening for the gene(s) encoding the cognate antigen(s) in the human leucocyte cDNA library.

Detection of cognate antigen(s) for antineutrophil antibodies on PMN surface by membrane biotinylation, streptavidin-column elution and Western immunoblot

PMN lysates were prepared according to the method reported by Garrels and Gibson [27]. One hundred microlitres of sample buffer (1 \times = 125 mM Tris-HCl pH 6.8, 2% SDS, 5% glycerol, 0.03% bromophenol blue and 1% β -mercaptoethanol) were added to lyse 1×10^7 freshly prepared neutrophils in pellets followed by boiling the lysates for 7 min with frequent vigorous vortexing. The precipitates were removed after centrifugation at 11 800 *g* at 4°C for 15 min. The clear PMN lysates (15 μ l/well) were electrophoresed in 10% SDS-PAGE. The dispersed proteins in gel were transferred to a nitrocellulose paper in a semidry transfer system. The membrane was immersed in the blocking buffer (5% non-fat milk in wash buffer containing 10 mM Tris pH 7.5, 100 mM NaCl, and 0.1% Tween-20) for 1 h at room temperature and then probed by either anti-PMN(+) SLE or normal serum at 1 : 4 diluted in 5% non-fat milk containing wash buffer. HRP-conjugated goat antihuman IgG antibodies (Chemicon) diluted at 1 : 5000 in wash buffer was used as the secondary antibodies. The antigen-antibody reaction was detected by the enhanced chemiluminescence (ECL) protein detection system (Amersham International, Amersham, UK). To detect the cognate antigen(s) of anti-PMN expressed on the cell surface of PMN, freshly isolated PMN (5×10^6 cells/ml) suspended in borate buffer (50 mM

sodium borate in 150 mM NaCl, pH 8.0) were membrane biotinylated by incubation with 1 ml of biotin-labelling buffer (0.1 mg biotin-7-NHS/ml in borate buffer) (Boehringer-Mannheim Biochemicals, GmbH, Mannheim, Germany) at 4°C for 1 h with gentle shaking. After several washes with PBS, 1 ml of the extraction buffer containing 50 mM sodium borate, 150 mM NaCl, 1% NP-40, 0.5% sodium deoxycholate and 0.1 mg/ml PMSF, pH 8.0 were added to the pellet to lyse the membrane biotinylated PMN. The cell lysates were sonicated further in an ice-bath. After centrifugation at 5200 *g* for 10 min, the biotinylated membrane proteins were absorbed into the streptavidin beads packing column (ImmunoPure[®], Pierce, Rockford, IL, USA) and eluted by glycine buffer, pH 2.5. The detailed procedures were described in the manufacturer's instruction sheets. The eluted biotinylated membrane proteins of PMN were then treated with 2 × sample buffer and the cognate antigens of anti-PMN in PMN membrane lysates were identified by Western immunoblot probed by anti-PMN(+) SLE serum.

Peptide mapping of surface membrane expressed cognate antigen(s) for anti-PMN(+) SLE sera

The surface membrane-expressed molecules on PMN reactive with anti-PMN(+) SLE sera in Western immunoblot gel were excised and eluted. After proteases (Lys C) cleavage and alkylation to prevent the linkage of multiple peptides through disulphide bonds, the digested peptides were separated further by reverse-phase HPLC on octadecyl silica gel. The peptide mapping was analysed using the ProFound computer program at the Beckman Center, Stanford University Medical Center.

Immunoscreening and molecular cloning of the genes encoding the cognate antigens of anti-PMN from human leucocyte cDNA library by anti-PMN(+) SLE sera

The screening and cloning of the genes encoding the cognate antigens of anti-PMN autoantibodies was carried out according to the method reported by Young and Davis, with some modifications [28,29]. A human leucocyte cDNA expression library (λTriplExTM and λTriplEx2TM) purchased from Clontech Laboratories Inc. (Palo Alto, CA, USA) was immunoscreened by SLE sera containing anti-PMN autoantibodies. Briefly, recombinant phages were plated on *E. coli* strain XL1-Blue at 1×10^5 plaques per 15-cm diameter dishes. β-galactosidase fusion protein expression was induced and transferred to nitrocellulose filters presoaked with 10 mM isopropyl thiogalactopyranoside (IPTG). The filters were screened with SLE sera containing high titre anti-PMN autoantibodies by Western immunoblot. The filters were then reacted with HRP-conjugated goat antihuman IgG (Chemicon) and peroxidase colour development reagent 4-chloro-1-naphthol (Sigma). Positive regions (discoloration) were excised, eluted, replated and rescreened until the plaques were uniformly positive. The insert sizes of positive clones were determined by PCR. The nucleotide sequence of the inserts was determined by a dsDNA cycle sequencing system (Gibco BRL Inc., Gaithersburg, MD, USA) and identified by the nucleotide blast program from the National Center for Biotechnology Information (NCBI). The detailed procedures were described in the manufacturer's instruction booklet.

Cloning and expression of human full-length SSB/La protein

Total cellular RNA was extracted from Jurkat cells (human T-lymphoma cell line) (1×10^7 /ml) after 6 h incubation with PHA (100 ng/ml) (Sigma) at 37°C, 5% CO₂ –95% air using the RNeasy[®] Mini kit (Qiagen GmbH, Hilden, Germany). cDNA was synthesized with 1 μg of denatured total RNA at 42° for 1 h in a final volume of 25 μl containing 1 μg of SSB/La antisense primer, 200 nmole of each dNTP (Pharmacia Fine Chemicals, Piscataway, NJ, USA) and 200 U M-MLV reverse transcriptase (Promega Corporation, Madison, WI, USA). The cDNA was amplified by PCR using paired oligonucleotide primers specific for human SSB/La (5'Sall-SSB, ACGCGAGTCGACATGGCTGAAAATGGTG AT AATGAAAAG; 3'XhoI-SSB(H), CATTATCTCGAGCTG GTCTCCAGCACCATT TCTG). A HYBAID OmniGene DNA thermal cycler (Teddington, Middlesex, UK) was used to run 35 cycles of denaturation at 95°C for 1 min, annealing at 56°C for 1 min and extension at 60°C for 2 min. The PCR product of 1227 bp SSB/La cDNA was purified by GFXTM PCR

DNA and Gel Band purification kit (Amersham Pharmacia Biotech, Inc., Piscataway, NJ, USA), was used later as the insert.

Two microlitres of *E. coli* XL1-Blue were incubated in 3 ml LB broth (1 g NaCl, 1 g tryptone peptone and 0.5 g yeast extract in 100 ml LB broth) with shaking at 37°C overnight. After refreshing in 40 ml LB for 1 h and on ice for 10 min, the competent cell XL1-Blue was prepared by adding 100 mM CaCl₂ and incubating on ice for 20 min. Plasmid vector pET23b (Novagen Inc., Madison, WI, USA) transformed into competent XL1-Blue by heat shock at 42°C for 2 min was incubated in LB broth with shaking at 37°C overnight. The pET23b was extracted and purified by GFX™ Micro Plasmid Prep kit (Amersham Pharmacia).

Five hundred nanograms of purified SSB/La PCR product and vector pET23b, respectively, were digested at 37°C by restriction enzyme Sall (Promega) and XhoI (Promega) overnight, respectively. The same amount of 50 ng digested SSB/La and pET23b was incubated with 1 μl T4 DNA ligase (Epicentre Technologies, Madison, WI, USA) and ATP (1 mM) at 4°C overnight for ligation. After cleaning with the GFX™ PCR DNA purification kit, 10 ng of ligated pET23b vector was transformed into competent XL1-Blue *E. coli* by heat shock. The transformed XL1-Blue was refreshed in LB broth at 37°C for 1 h and cultured on LB agar plates containing ampicillin (75 μg/ml) at 37°C overnight. The success of plasmid transformation enables the growth of XL1-Blue in LB agar plates with ampicillin at 37°C overnight. The single colony of XL1-Blue was transferred to 1 ml LB broth with ampicillin and incubated at 37°C overnight. After centrifugation at 1300 **g** for 10 min, the plasmid pET23b was purified by use of the GFX™ Micro Plasmid Prep kit. The pET23b vector containing SSB/La insert was screened by PCR using oligonucleotide primers of SSB/La and confirmed by DNA sequencing.

Competent cells of BL21(DB3)pLysS (Promega) were prepared and the pET23b containing SSB/La was transformed into these competent cells, as described above. The transformed BL21(DB3)pLysS was cultured in LB agar plates with ampicillin (75 μg/ml) at 37°C overnight. After refreshing the cultured BL21(DB3)pLysS colony in LB broth by shaking at 37°C for 3 h (bacterial concentration O.D._{600nm} = 0.6–1.0), the induction of recombinant SSB/La was conducted by adding IPTG (0.4 mM) and incubation at 37°C for 3 h. After sonication of IPTG-induced BL21(DB3)pLysS in 150 μl PBS and centrifugation at 11800 **g** for 30 min, the pellet (insoluble form) and supernatant (soluble form) were prepared for Western blotting, as mentioned above.

The SSB/La protein from IPTG-induced transformed BL21(DB3)pLysS was purified by use of a HisTrap kit (Amersham Pharmacia). In brief, the induced BL21(DB3)pLysS was sonicated completely after incubation with imidazole (10 mM) containing binding buffer. The supernatant was collected and eluted through HisTrap Chelating HP columns with immobilized divalent cations (Ni²⁺). The target SSB/La expressed protein was recovered by elution with gradient imidazole concentrations from 10 mM, 20 mM and 40 mM to 500 mM. The eluted purified protein was dialysed in PBS buffer, pH 7.2 and then concentrated with disposable microconcentrator. The detailed procedures were described in the manufacturer's instruction booklet. The purity and identification of the recombinant SSB/La protein were detected by 10% SDS-PAGE and Western blot probed by anti-SSB/La autoantibodies purified from anti-PMN(+) SLE sera.

Purification of anti-SSB/La antibodies from anti-PMN(+) SLE sera by recombinant SSB/La affinity column

The recombinant SSB/La protein conjugated CNBr-activated Sepharose 4B gel packing column (Pharmacia Biotech, Wisnoms, Sweden) was prepared by the method reported elsewhere in the literatures. The anti-PMN(+) SLE sera were incubated with the SSB/La affinity column at 4°C overnight. After washing with Tris buffer, pH 8.0, the anti-SSB/La antibodies were eluted by glycine buffer, pH 2.5. The detailed procedures were described in the manufacturer's instruction sheets. The antigen-binding activity of the purified human anti-SSB/La antibodies was detected by Western immunoblot using the same recombinant SSB/La as substrate. In addition, we used a commercial immunoassay kit (line immunoassay) (anti-ENA) (SLR Research Corporation, Carlsbad, CA, USA) to detect the presence of anti-extractable

nuclear autoantibodies, including anti-RNP, anti-Scl-70, anti-SSA/Ro, anti-SSB/La, anti-Jo-1 and anti-Sm specificities in these affinity column-purified anti-SSB/La autoantibodies. The detailed procedures were described in the manufacturer's instruction sheets. We found that different eluted samples contained different potencies of anti-SSB/La but seldom other anti-ENA activity (data not shown).

Measurement of PMN apoptosis by flow cytometry

PMN apoptosis was measured by flow cytometry according to the method of Nicoletti *et al.* [30]. Briefly, 400 μ l of agglgG-pretreated PMN (2×10^6 cells/ml) were incubated with 100 μ l non-specific human IgG (0.5 mg/ml) or anti-SSB/La (0.5 mg/ml) purified from anti-PMN(+) SLE in a conical tube for 2 h at 37°C in 5% CO₂ –95% air. After three washes, the treated PMN were incubated in 1 ml of 10% FBS-RPMI for 24 h and 48 h at 37°C in 5% CO₂ –95% air. At the end of each culture period, the PMN was stained in 0.5 ml of hypotonic fluorochrome solution (50 μ g/ml propidium iodide in 0.1% sodium citrate and 0.2% Triton X-100) at 4°C in the dark for 1 h to stain the nuclei. The fluorescence intensity of the cell nuclei was measured by FACSsort flow cytometer (Becton-Dickinson, Mountain View, CA, USA) with 488 nm excitation. The percentage of hypodiploid DNA appeared in the left of the diploid DNA peak was regarded as percentage apoptotic PMN.

Measurement of PMN phagocytosis by flow cytometry

The method described by Shalaby *et al.* [31] was used with some modification for measuring PMN phagocytosis. Briefly, the fluoresbrite carboxylate microspheres (0.75 μ m in diameter, Polyscience Inc., Washington, PA, USA) were opsonized with fresh normal human serum at 37°C for 1 h. After two washes, the opsonized microspheres were adjusted to 1×10^9 particles/ml in PBS, pH 7.2. Freshly prepared PMN (2×10^6 /ml) were preincubated with culture medium, LPS (100 ng/ml), non-specific human IgG (0.5 mg/ml) or purified human anti-SSB/La (0.5 mg/ml) for 45 min, followed by incubation with 100 μ l opsonized fluorescence microspheres for another 45 min. The treated PMN was then fixed with 2% paraformaldehyde to stop phagocytosis. After several washes, the percentage and mean fluorescence of PMN phagocytosis was determined by FACSsort flow cytometer (Becton-Dickinson) at 488 nm excitation.

Detection of anti-SSB/La antibody penetration into PMN by flow cytometry

AgglgG-pretreated PMN (1×10^6 /ml) were incubated with culture medium, purified human anti-SSB/La (0.5 mg/ml) or non-specific human IgG (0.5 mg/ml) in ice-bath for 30 min. After washing three times with PBS, pH 7.2, the PMN was fixed with 4% paraformaldehyde for 15 min, treated with 0.1% Triton X-100 for 5 min and stained with 1 : 100 diluted FITC-conjugated goat antihuman IgG antibodies (Jackson ImmuniResearch Laboratories Inc., West Grove, PA, USA) in an ice-bath for 30 min. To prevent the non-specific endocytosis of PMN, the cells were pretreated with 10 ng/ml of cytochalasin B (CyB, Sigma) for 45 min. The percentage of active penetration of anti-SSB/La or non-specific IgG was calculated by the following formula after flow cytometric determination:

$$\% \text{ penetration of protein} = (\text{PMN} + \text{CyB} + \text{protein})\% - (\text{PMN} + \text{CyB} + \text{medium})\%.$$

ELISA for IL-8 concentration of PMN cultured supernatants

AgglgG-pretreated PMN (1×10^6 /ml) were preincubated with 0.5 mg/ml of human IgG or anti-SSB/La antibodies in conical tubes for 2 h as mentioned above. After washing, the treated PMN were cultured continuously in 10% FBS-RPMI for 24 h at 37°C in 5% CO₂ –95% air. The culture supernatants were collected for IL-8 quantification using a commercially available human IL-8 ELISA kit (Endogen, Woburn, MA, USA). The detailed procedures were described in the manufacturer's instruction booklet. The minimal detectable concentration of IL-8 was 18.1 Pg/ml.

Statistical analysis

Results represent the mean \pm s.d. in this study. Statistical significance was assessed by Student's *t*-test or

non-parametric Wilcoxon rank sum test.

Results

PMN-binding activity of SLE sera

Seventy-one sera from patients with active SLE and an equal number of normal individuals were compared for PMN-binding capacity using the cellular ELISA method. A significant difference of PMN-binding activity was found between SLE sera ($O.D_{.490nm} = 1.683 \pm 0.312$) and normal sera ($O.D_{.490nm} = 1.337 \pm 0.347$) (data not shown). For further investigation, the anti-PMN(+) serum was defined as the serum with a PMN-binding capacity \geq mean + 2 s.d. of normal sera. We found that 16/71 (22.5%) of SLE sera were positive for anti-PMN, as shown in Fig. 1a. These anti-PMN(+) SLE sera were used for the detection and immunoscreening of the genes encoding the cognate antigen(s) of anti-PMN from human leucocyte cDNA library, as described below.

Detection of cognate antigen(s) of anti-PMN autoantibodies on PMN by Western immunoblotting after membrane biotinylation and streptavidin-column elution

We used total cell lysates and the lysates after membrane biotinylation and streptavidin-column purification as substrates in Western blot that were probed by anti-PMN(+) SLE or normal serum. As demonstrated in Fig. 1b(1), several cognate antigens in total cell lysates of PMN (lane 3) and MNC (lane 1) were detected by anti-PMN(+) SLE serum, but not by normal serum [Fig. 1b(2)]. The molecular weights of the cognate antigens in total PMN lysates detected by anti-PMN autoantibodies were 15, 30, 50 and 100 kDa proteins (lane 3). However, when PMN membrane lysates were probed by anti-PMN(+) SLE in Western blot after membrane-biotinylation and streptavidin column purification, we found 30% of anti-PMN(+) SLE sera, but not normal sera, reactive with 50 kDa surface expressed molecules on PMN [Fig. 1b(1), lane 4]. This 50 kDa band was cut for peptide digestion and amino acid sequence identification. The preliminary result demonstrated that the protein is 46% homologous with SSB/La molecule (data not shown).

Immunoscreening, molecular cloning and nucleotide sequencing of the genes encoding cognate antigens of anti-PMN antibodies

Immunoscreening of human leucocyte cDNA expression library by anti-PMN(+) SLE sera exhibited three positive clones (one 0.8 Kb and two 0.9 Kb) encoding the cognate antigens of anti-PMN autoantibodies [Fig. 2a(1), (2), (3)]. The nucleotide sequencing of the PCR amplified inserts revealed that the 0.8Kb clone A [Fig. 2a(1)] was 97% homologous with the gene encoding SSB/La protein (47.7 kDa) [Fig. 2b]. The 0.9Kb clone B was 99% homologous with human histone 3 (15.3 kDa), but the other 0.9Kb clone C was a hypothetical gene after computer analysis of nucleotide blast program provided by NCBI.

Cloning and expression of full-length SSB/La DNA from Jurkat T cells

Full-length cDNA of SSB/La was prepared from PHA-stimulated Jurkat T cells by RT-PCR. After digestion by restriction enzyme, plasmid pET23b and SSB/La were ligated and transformed into *E. coli* XL1-Blue. The insertion of SSB/La in pET23b was amplified by PCR and confirmed further by direct DNA sequencing (data not shown). The SSB/La carried vector pET23b and was then transformed into the BL21(DB3)pLysS expression system. After induction with ITPG, the SSB/La protein was collected and conjugated with the HisTrap chelating HP column. The human recombinant SSB/La protein was eluted effectively by a high concentration (500 mM) of imidazole and the purity and molecular weight of the recombinant SSB/La were determined by 10% SDS-PAGE [Fig. 3a, lane 4].

Purification of anti-SSB/La antibodies from anti-PMN(+) SLE sera by affinity column

Anti-PMN(+) SLE serum was absorbed in a recombinant SSB/La conjugated Sepharose 4B gel column after incubation at room temperature for 2 h. After several washes, the anti-SSB/La antibodies were eluted by glycine buffer, pH 2.5 and

the anti-SSB/La containing elute was neutralized immediately with PBS, pH 7.2. The antigen-binding activity of the purified anti-SSB/La antibodies was confirmed by Western blot using recombinant SSB/La as antigen (Fig. 3b, lane 3). Normal serum (lane 1) and mouse monoclonal anti-dsDNA (lane 2) did not react with 50 kDa SSB/La protein. Alternatively, monoclonal anti-dsDNA antibody did not cross-react with SSB/La but anti-PMN(+) SLE serum (lane 4), the purified anti-SSB/La (lane 3) and anti-His C-terminal antibodies (lane 5) reacted potently with SSB/La. In addition, a line immunoassay that is considered as the 'gold standard' in detecting anti-extractable nuclear protein autoantibodies used in the clinical immunological laboratory also confirmed that the purified anti-SSB/La autoantibodies reacted mainly with SSB/La (data not shown). These purified anti-SSB/La antibodies were tested for immunopathological effectiveness on normal PMN functions.

Impaired PMN phagocytosis by human anti-SSB/La autoantibodies

Normal human neutrophils (1×10^6 /ml) were incubated with human anti-SSB/La (0.5 mg/ml), non-specific human IgG (0.5 mg/ml), LPS (100 ng/ml) or culture medium for 1 h. One hundred microlitres of opsonized fluorescence microspheres (1×10^9 beads/ml) was added to PMN suspension and incubated for another 45 min for phagocytosis assay. A representative case demonstrating impaired PMN phagocytosis in both percentage (%) and mean fluorescence intensity (#MFI, denoted by mean fluorescence channel) by anti-SSB/La compared to non-specific human IgG is showed in Fig. 4. The same experiment was repeated three times with a similar tendency.

Increased neutrophil apoptosis by human purified anti-SSB/La autoantibodies

We preincubated normal human PMN with purified anti-SSB/La autoantibodies (0.5 mg/ml) or non-specific human IgG (0.5 mg/ml) for 24 h and 48 h. PMN apoptosis was determined by flow cytometry. The percentage of hypodiploid DNA in the pre-G0/G1 phase of cell cycle was regarded as the percentage PMN apoptosis. A representative case showed that the percentage apoptosis of PMN incubation with anti-SSB/La was higher than with non-specific human IgG after 24 h (Fig. 5a, left panel) and 48 h (Fig. 5a, right panel) incubation. The same experiment was repeated four times with a similar tendency.

Binding and penetration ability of anti-SSB/La autoantibodies toward PMN

For evaluating the penetrating activity of anti-SSB/La autoantibodies into PMN cells, we preincubated normal human PMN \pm cytochalasin B (10 ng/ml) for 45 min and then reacted with anti-SSB/La or non-specific human IgG in an ice-bath for 30 min. After fixation and permeabilization, the cells were stained with FITC-conjugated goat antihuman IgG. A representative case is presented in Fig. 5b in that the percentage of active penetration of anti-SSB/La (51.24%) was much higher than IgG (21.09%) by calculation. The same experiment was repeated six times with a similar tendency.

Increased neutrophil interleukin 8 production by anti-SSB/La

As demonstrated in Fig. 6, anti-SSB/La antibodies stimulated IL-8 production of human PMN more than non-specific human IgG. Combining the results of Figs 5 and 6, it is possible that anti-PMN autoantibodies activated simultaneously the genes mediating IL-8 synthesis and apoptosis that seems to be compatible with the phenomenon of activation-induced cell death (AICD).

Discussion

Polymorphonuclear neutrophils are regarded traditionally as terminally differentiated cells incapable of protein synthesis and are committed to death via apoptosis within 72 h [32–34]. However, recent studies indicate that neutrophils are not only capable of receiving signals from different proinflammatory cytokines, bacterial lipopolysaccharide and glucocorticosteroids to suppress apoptosis [35–38], but also synthesize many important proinflammatory cytokines,

chemokines and haematopoietic growth factors after stimulation to modulate immune responses [6,7]. Neutropenia, one of the common haematological abnormalities in systemic lupus erythematosus, correlates with disease activity and becomes a major cause of morbidity/mortality in patients [19,23,39]. It is conceivable that both accelerated apoptosis and impaired functions of the neutrophils render SLE patients susceptible to bacterial infection [11–15,39]. Many authors have demonstrated that increased neutrophil apoptosis in SLE is owing to the cytotoxic effect of anti-PMN autoantibodies in the circulation [19–24,40]. However, the real immunopathological roles and the cognate antigen(s) of the anti-PMN autoantibodies were not elucidated in the literature.

Four original findings were derived from the present study: (a) approximate 20% active SLE sera contained PMN binding autoantibodies as demonstrated in Fig. 1a. (b) The cognate antigens of anti-PMN autoantibodies including 15, 30, 40, 50 and 80 kDa molecules (Fig. 1b, lane 3) were present in total PMN lysates, but only 50 kDa molecule (Fig. 1b(1), lane 4) is expressed on the surface of PMN. (c) Increased PMN apoptosis (Fig. 5a) and decreased PMN phagocytosis (Fig. 4) were exerted by anti-SSB/La autoantibodies. (d) Anti-SSB/La autoantibodies activated PMN to increase IL-8 production (Fig. 6). Biologically, apoptosis is linked intimately to cell stimulation and proliferation [41,42]. Accordingly, cell activation and cell death are inseparable once the cells are activated. The activation-induced cell death (AICD) phenomenon is found commonly in antigen- or mitogen-activated immune-related cells [43,44]. The immunopathological effects of anti-SSB/La on PMN were seen through the AICD mechanism, because both IL-8 and apoptotic genes in PMN are elicited simultaneously by these antibodies. Interestingly, our results are similar to those in a number of reports. Muhl *et al.* [45] reported the expression and release of chemokines associated with apoptotic cell death in human promonocytic U937 cells and peripheral blood mononuclear cells. Hagimoto *et al.* [46] demonstrated that Fas receptor ligation induced IL-8 secretion and apoptosis in bronchiolar epithelial cells. Lai *et al.* [47] demonstrated that polyclonal anti-dsDNA autoantibodies enhanced the gene expression of IL-8, TGF- β and nitric oxide synthase in cultured umbilical vein endothelial cells in parallel with the degree of cell apoptosis. However, whether these immunopathological effects of anti-SSB/La autoantibodies on PMN are compatible with AICD needs further investigation.

Because many IgG Fc receptors are expressed on the PMN surface, it is possible that the immunopathological effects of anti-SSB/La autoantibodies on PMN were through the interactions of Fc portion of antibody and Fc γ receptors on PMN to transduce certain signals. To avoid these non-specific effects of Fc γ receptor, we used heat-aggregated human γ -globulins as surrogate immune complexes to ligate Fc γ R on PMN in advance in some of our experiments. Our results indicate that the immunopathological effects of anti-SSB/La autoantibodies on PMN including cytotoxicity and decreased neutrophil phagocytosis were mediated by specific anti-PMN activity *per se*, but not by the non-specific interactions of IgG Fc portion and Fc γ receptors on PMN (data not shown). Our preliminary results also confirmed that anti-SSB/La F(ab')₂ fragments as well as intact IgG molecules could impose the above-mentioned immunopathological effects on human PMN (data not shown).

Interleukin 8 (IL-8) can stimulate many neutrophil functions, such as chemotaxis, release of lysosomal enzymes and generation of reactive oxygen metabolites [48,49]. In addition, IL-8 acts as an autocrine stimulator to enhance IL-8 production of neutrophils. A growing body of evidence suggests that IL-8 prolongs the life span of neutrophils by suppressing the apoptotic process [35]. Thus, PMN *per se* could amplify the recruitment of PMN, rescue the short life span and enhance inflammatory strength in the inflammatory sites by autocrine IL-8 synthesis. In our previous report, we demonstrated that the spontaneous and bacterial lipopolysaccharide-stimulated production of IL-8 *in vitro* were impaired in neutrophils of patients with SLE [13]. Furthermore, our preliminary data suggested that the expression of CXCR2, a specific ligand for IL-8, on SLE-PMN was significantly decreased compared to normal PMN. This result suggests that intrinsic defects of PMN *per se* or the presence of certain undiscovered suppressor factors in the blood cause hyporesponsiveness of PMN to external stimulation in SLE patients. Accordingly, it is possible that anti-PMN

autoantibody is one of the suppressor factors in SLE serum to derange the PMN functions via their cytotoxic effects. In the present study, we have successfully immunoscreened and cloned the gene-encoding SSB/La protein by anti-PMN(+) SLE sera and proved that anti-SSB/La antibody is one of the anti-PMN autoantibodies mediating immunopathological effects on PMN.

Several unique immunopathological activities of anti-SSB/La autoantibodies on PMN in SLE serum were noted: (a) anti-SSB/La is one of the anti-PMN autoantibodies in SLE serum. (b) Anti-SSB/La bind not only to the membrane expressed autoantigen SSB/La on PMN, but also penetrate into PMN. (c) The binding/penetration of anti-SSB/La enhances IL-8 production, but impairs the phagocytosis and accelerates the apoptosis of normal human PMN. SSB/La, a 43–52-kDa ribonucleoprotein, participates in the transcriptional regulation of RNA polymerase III as well as some messenger RNAs. Anti-SSB/La appears in as many as 87% of Sjögren's syndrome in association with extraglandular manifestations such as neurological involvement, vasculitis, purpura, leucopenia and lymphopenia [50]. Approximately 10–15% of lupus patients produce anti-SSB/La autoantibodies, especially in association with late-onset SLE, secondary Sjögren's syndrome and the neonatal lupus syndrome [50]. However, reports on the immunopathological roles and clinical relevance of anti-SSB/La in lupus pathogenesis are limited. It has been recognized that many cytoplasmic and nuclear proteins such as SSA/Ro, Sm, proteinase 3 and ribosomal phosphoproteins can translocate to the cell surface after stimulation or in apoptosis. These surface-translocated molecules then become the major autoantigens to induce autoantibody production in certain autoimmune diseases [51–53]. On the other hand, these induced autoantibodies against nuclear/cytoplasmic autoantigens exert their adverse effects on normal tissues via direct cytotoxicity/apoptosis, formation of immune complexes or penetration into cell interior [21,54,55]. In the present study, we have demonstrated that SSB/La autoantigen translocates to the cell surface of PMN (data not shown) and anti-SSB/La autoantibodies impose a pathological effect on PMN via cellular surface binding and then penetration into the cells to induce apoptosis. This may lead to neutropenia and functional impairment of PMN in patients with SLE. However, other autoantigen/autoantibody systems such as antibodies against dsDNA (56), ribosomal P proteins or HLA molecules should be considered as other potential contributory factors for neutropenia in patients with SLE.

In conclusion, we are the first authors to find that anti-SSB/La is one of the antineutrophil autoantibodies binding to the surface-expressed SSB/La autoantigens on PMN *in vitro* and is probably responsible for neutropenia and impaired neutrophil functions *in vivo* in patients with SLE.

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Fig. 1. Detection of cognate antigen(s) of anti-PMN autoantibodies on normal human neutrophil (PMN) surface (b) after screening the high titre of anti-PMN sera from patients with SLE (a). (a) Comparison of PMN-binding activity of SLE and normal sera by PMN cellular ELISA (please see [Materials and Methods](#)). (b) Identification of cognate antigen(s) of anti-PMN autoantibodies on PMN surface after membrane biotinylation, streptavidin-column purification and Western blot analysis of PMN lysates probed by (1) SLE serum, (2) normal serum. Lane 1: total human MNC lysate; lane 2: MNC lysate after membrane biotinylation and streptavidin column purification; lane 3: total PMN lysate; and lane 4: PMN lysate after membrane biotinylation and streptavidin column purification. A 50–60 kDa surface-expressed molecule on PMN was found reactive with anti-PMN autoantibodies in SLE serum [b-(1)], but not reactive with normal serum [b-(2)]. The same experiment has been conducted 10 times with a similar tendency.

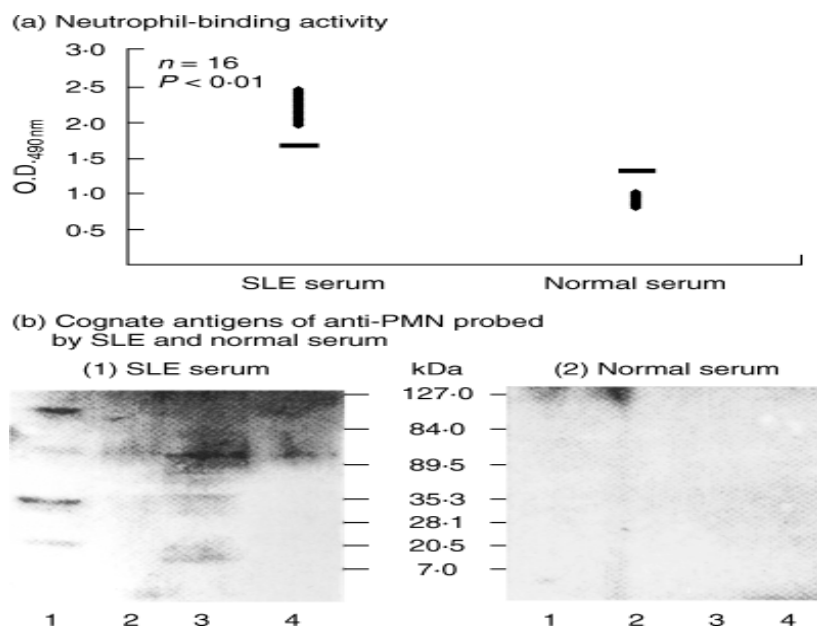


Fig. 2. Immunoscreening, molecular cloning, and nucleotide sequencing of the gene(s) encoding cognate autoantigen(s) of anti-PMN antibodies from human leucocyte cDNA expression library probed by anti-PMN(+) SLE sera. (a) Immunoscreen of a human leucocyte cDNA library, λ TripIExTM and λ TripIEx2TM, probed by high titres of anti-PMN(+) SLE serum. Three clones, clones A, B and C, were obtained. The DNA inserts in the positive clones were amplified by PCR and then electrophoresed in agarose gel. (1) Eight representative colonies were selected from clone A for PCR amplification and all exhibited 0.8Kb. (2) Six representative colonies are selected from clone B for PCR amplification and all exhibited 0.9Kb. (3) Three representative colonies were selected from clone C for PCR amplification and all of them exhibited 0.9Kb. M: DNA marker (λ /HindIII + Φ X174/HaeIII). (b) Nucleotide sequence of

clone A (525 bp) showed 97% (510/525) homology with La/SSB (47.7 kDa) analysed by nucleotide BLAST program of NCBI.

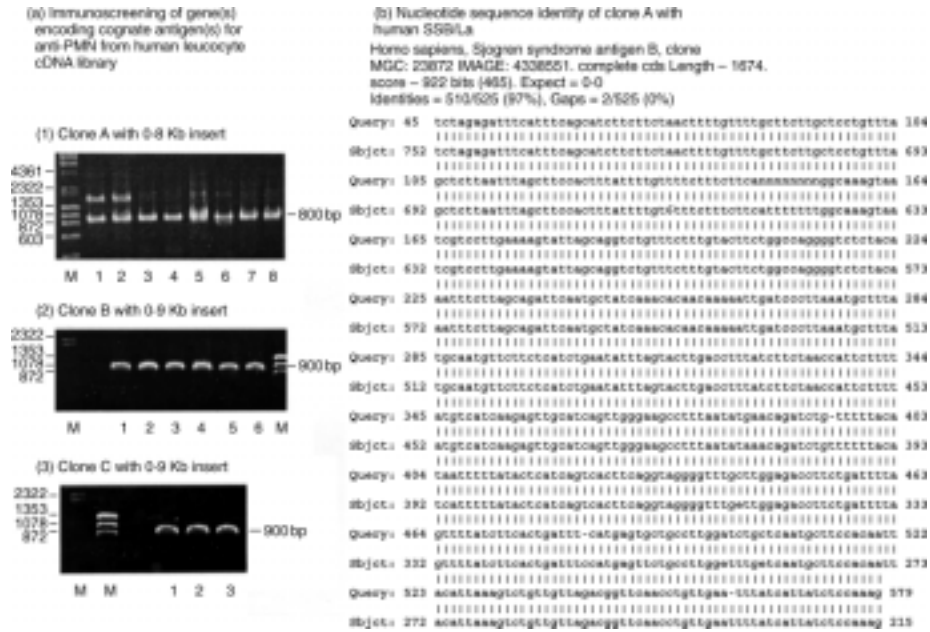


Fig. 3. Purification and identification of recombinant human SSB/La protein expressed in *E. coli*. (a) Detection of recombinant SSB/La protein by 10% SDS-PAGE after elution with different concentrations of imidazole through HisTrap chelating HP column. Lane 1: crude *E. coli* extract; lane 2: elute with PB buffer pH 7.2; lane 3: elute with 10 mM imidazole in PB buffer; and lane 4: elute with 500 mM imidazole in PB buffer. (b) Western immunoblotting of the eluted SSB/La protein probed by different sera and antibodies; lane 1: 1 : 50 diluted normal human serum; lane 2: 1 : 20 diluted monoclonal mouse anti-dsDNA antibody; lane 3: 1 : 50 diluted human anti-SSB/La antibodies purified from anti-PMN(+) SLE sera; lane 4: 1 : 50 diluted SLE sera; and lane 5: 1 : 5000 diluted monoclonal anti6xHis C-terminal antibody.

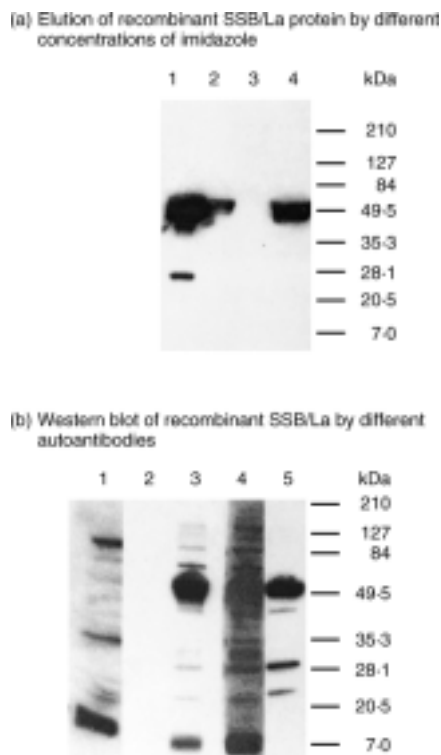


Fig. 4. A representative case demonstrates the effect of purified human anti-SSB/La on PMN phagocytosis. PMN was preincubated with anti-SSB/La (0.5 mg/ml) for 45 min, followed by reacting with opsonized fluorescence microspheres. The percentage (%) and mean fluorescence (MFI#) of PMN phagocytosis were detected by flow cytometry. The same experiment was repeated for three times with a similar result.

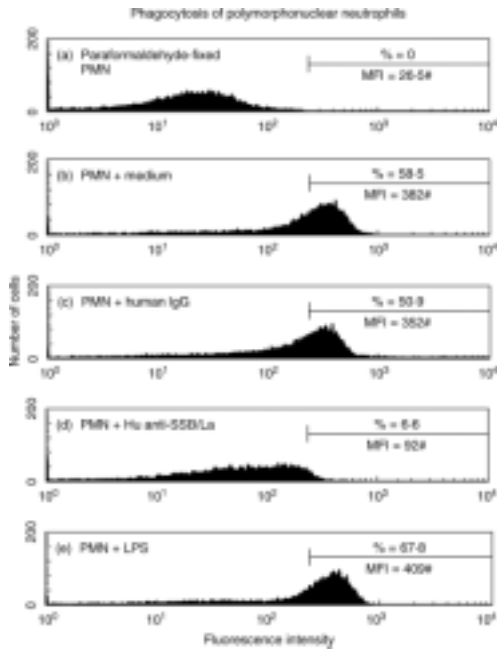


Fig. 5. The effect of purified human anti-SSB/La autoantibodies on agglgG-treated PMN apoptosis and the penetrating activity of the autoantibodies into human neutrophils. (a) The effect of purified human anti-SSB/La (0.5 mg/ml), and human IgG (0.5 mg/ml) on PMN apoptosis was determined by flow cytometry. PMN was incubated with medium, human IgG and anti-SSB/La for 24 h and 48 h. The percentage of hypodiploid DNA appeared before the G₀/G₁ phase of cell cycle was regarded as percentage apoptosis. The same experiment was repeated for four times with a similar tendency on each occasion. (b) A representative case demonstrates the binding and penetrating activities of the purified anti-SSB/La autoantibodies into PMN. PMN was preincubated with/without cytochalasin B (10 ng/ml) and then reacted with anti-SSB/La (0.5 mg/ml) or non-specific human IgG (0.5 mg/ml) for 30 min in an ice-bath. The percentage of intracellular IgG was determined by flow cytometry after fixed with 4% paraformaldehyde and permeabilization with 0.1% Triton X-100. The same experiment was repeated three times with a similar tendency.

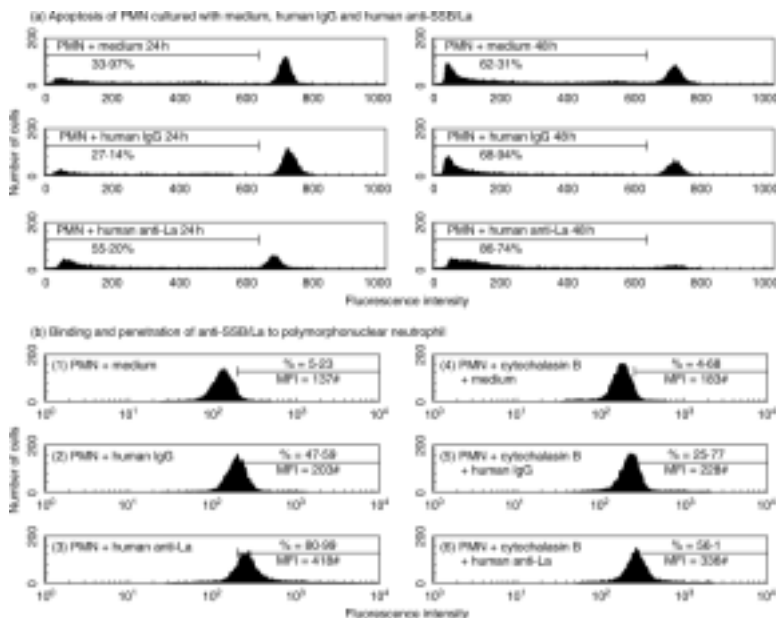
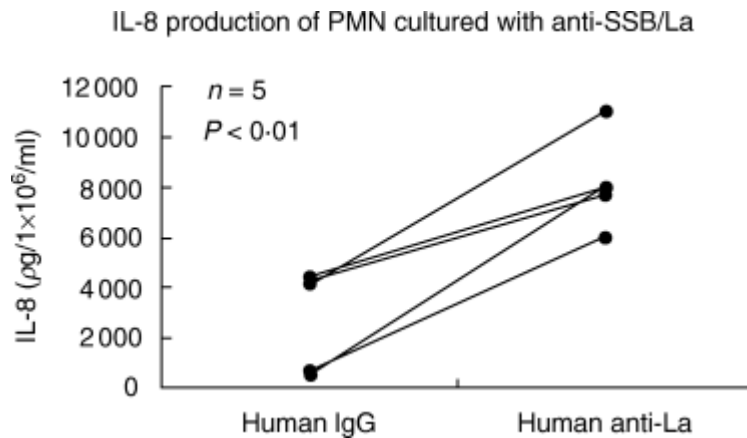


Fig. 6. Comparison of IL-8 production by normal human PMN after incubation with purified anti-SSB/La (0.5 mg/ml) and human IgG (0.5 mg/ml) for 24 h detected by commercial ELISA kit.



討論

本三年期研究計劃旨在探討全身性紅斑狼瘡患者血液中白血球減少的免疫病理學機轉，即是患者血清中是否有白血球傷害抗體的存在。我們的結果顯示 SLE 血清中有抗白血球抗體(anti-PMN)的存在。本抗體除會引發白血球的細胞凋亡之外，在引發細胞凋亡之前，會先活化 PMN 包括增強 PMN phagocytosis 及 IL-8 的產生。這些結果顯示 SLE 血清中確有傷害性的 anti-PMN 抗體的存在。其次，為了進一步證明及鑑定抗 PMN 抗體在 PMN 細胞膜上對應抗原的存在，我們以 Western blot 分析法將 biotinylated PMN cell lysates 先行經過 normal 或 SLE sera 的 immunoprecipitation，再經 10% SDS-PAGE 電泳之後，以 HRP-avidin 為探針及呈色。我們發現 PMN 膜上 47-50kDa 分子量的蛋白質是 anti-PMN autoantibodies 的對應抗原。經 proteomics 分析得知其為 SSB/La protein。我們再以分子生物學的方法，由市販的人由來白血球 cDNA library，以富含 anti-PMN 抗體活性的 SLE 血清為探針，選殖出會轉錄 SSB/La protein 的基因。經核酸序列鑑定出此基因與是存在於細胞核內的 SSB/La 有 99.8%的吻合度，即是 RNA polymerase III binding protein 的基因。而其分子量為 47.5kDa。為了大量獲取純化的 SSB/La，我們經 DNA 選殖，繁衍，殖入，表現及純化之後，獲得了純粹的 SSB/La。我們將含 anti-SSB/La activity 的 active SLE sera 經 SSB/La 的 affinity column 純化出 anti-SSB/La 抗體。這個抗體經證實與 SLE sera 的活性相仿。這些結果發表於 *Clinical and Experimental Immunology*, 2003.

為了使基因混成的 SSB/La 有其臨床應用，我們希冀本蛋白質分子可以中和 SLE 血清中抗 PMN 抗體。但是當我們將 anti-SSB/La 與 SSB/La 先行混合之後，與正常 PMN 作用時，發現 SSB/La 會有不甚明顯的中和 anti-PMN 的細胞傷害作用。這些結果令我們相當困惑。但經多次實驗之後，我們證實 SSB/La 本身也具有與 anti-SSB/La 相同的免疫活性，即是會先活化 PMN，使 PMN 的 phagocytosis 及 IL-8 的產生增加之後，接續引發 AICD 使細胞凋亡。雖然其真正的免疫病理機轉仍不清楚，但可能扮演著先天性免疫系統 danger signal 的角色。即是當身體的細胞因各種原因受到傷害致使細胞核內/細胞質內的 SSB/La 流出細胞外時，先天性的免疫細胞會受這些 SSB/La 分子的活化，而增強消除危害身體的傷害源。我們今後將致力於解明 SSB/La 蛋白成為 danger signal 的分子機轉，以建立新的學術理論。以上的結果待整理之後，將提出學術期刊發表。

自評

本計劃的成果已有部分發表於國際 SCI 學術期刊。還有部分成果有待補強之後再投稿於國際學術期刊。為們認為本計劃的最終目標在於臨床實務的應用，例如如何利用 recombinant SSB/La 來 protein 減緩 SLE 患者因白血球減少所產生的免疫力降低及免疫反應的改變。我們在另外的研究中發現 PMN 會影響 MNC 的免疫反應 (Immunology 95:480-7, 1998)。因此，leukopenia 的影響所及不如想像中的單純，不可等閒視之。值得免疫病理學者更加重視。總之，本計劃的成果應有其學術及臨床實務的價值。