

Analysis of Polymer Additives by Matrix-Assisted Laser Desorption Ionization/Time of Flight Mass Spectrometer Using Delayed Extraction and Collision Induced Dissociation

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Matrix-assisted laser desorption/ionization (MALDI) in combination with mass spectrometry proved to be a viable method for identifying additives in polyethylene extracts. The MALDI mass spectra of additives standards were found to be very simple consisting of $[M+H]^+$ and/or $[M+Na]^+$ pseudo molecular ions with little fragment ions. For real samples, which often contain more than one additive, the production of only one or two ions for each additive makes the tentative assignment much easier. Collision induced dissociation (CID) of the pseudomolecular ion is used to confirm the tentative assignment. The analysis of high molecular weight additives, such as Chimassorb 944 and Tinuvin 622 indicated that MALDI was superior to other ionization techniques such as electrospray ionization (ESI), desorption chemical ionization (DCI) and fast atom bombardment (FAB) in the analysis of high molecular weight polymer additives. Sample preparation was found to be more critical than DCI in the analysis of real sample. The signals were interfered by the low molecular weight polyethylene molecules, which were co-extracted with the additives. This problem was partially overcome by using acetone instead of methanol to precipitate the low molecular weight polyethylene molecules.

INTRODUCTION

The range of applications of polymeric materials is extending at an amazing rate and the number of additives available to modify polymer properties is increasing proportionately. These additives include antioxidants, UV stabilizers, Lubricants etc. The failures of products can often be attributed to the leaching of additives from polymer, the chemical transformation of certain additives and the omission of essential additives during the manufacturing process. Moreover, some additives are known as potential health hazards.¹ Therefore, reliable and rapid analytical methods are often needed for the identification of unknown additives in polymers and/or monitoring the stability of additives during processing or services.

Direct spectroscopic methods such as ultraviolet absorption, infrared, fluorescence, and X-ray fluorescence have been reported.²⁻⁵ The main disadvantage of the spectroscopic techniques is a lack of specificity. A more desirable approach requires extraction of the additives following by off- or on-line chromatographic separation and spectroscopic identification.⁶⁻¹⁰ These methods are generally time consuming.

Analysis of polymer additives by mass spectrometry

and its related techniques enhanced the capability of compound identification. Moving toward higher molecular weight additives with less volatility, conventional MS techniques such as electron ionization (EI), chemical ionization (CI) and GC-MS are generally not suitable for their analysis. Thus, soft ionization techniques such as field desorption, fast atom bombardment (FAB),^{11,12} desorption chemical ionization (DCI),^{13,14} laser desorption,¹⁵⁻²⁰ electrospray ionization (ESI)²¹ and atmospheric pressure chemical ionization (APCI)^{22,23} all have been investigated for their utility in the analysis of polymer additives either in polymer extracts or directly from polymer matrices.

Earlier studies from our laboratory suggested that DCI in combination with low and high energy CID was capable of detecting polymer additives in polyethylene without prior chromatographic separation.^{13,14} While DCI is quite useful to the analysis of a variety of additives, it does have limited capability in the analysis of high molecular weight additives.^{13,14} Since its development, matrix-assisted laser desorption/ionization (MALDI) has been widely used as a soft ionization technique for very large molecules. Furthermore, MALDI is known to have less matrix effect in comparison with FAB or ESI. However, despite its capability in the analysis of high

Dedicated to Professor Sheng-lieh Liu on the occasion of his ninetyeth birthday.

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mo lecu lar weight com pounds, there have been very few reports on the anal ysis of poly mer ad di tives by MALDI.^{24,25} In ad di tion, these re ports were also lim ited to a few ad di tives. Scrivens et al. re ported the anal ysis of poly mer ad di tives using dithranol as ma trix and add ing siliver trifluoroacetate to form siliver ad ducts.²⁴ The sig nals were re ported to be not very re pro duc ible. The siliver adduct ap proach has also been used by Kornfeld et al. in a poster pre sented at 1997ASMS conference.²⁵ A more ex ten sive study in clud ing the use of de layed ex trac tion, post source de cay (PSD) and CID in the anal ysis of poly mer ad di tives by MALDI was per formed in this lab o ra to ry. The mer its and lim i ta tions of the ap proach are re ported.

EXPERIMENTAL

Chemicals

Ad di tives stan dards were kindly pro vided by Ciba-Geigy (Tai wan). Ox i dized Naugard 524 was pre pared by ox i da tion of Naugard 524 with hy dro gen per ox ide. The struc tures and triv ial names of the ad di tives pre sented in this work are shown in Ta ble 1. The poly eth yl ene sam ples were ob tained from lo cal sup pli ers. 1,8-Dihydroxy-9-anthracenone (Dithranol) and 4-Hydroxy-3-methoxycinnamic acid (Ferulic acid) were purchased from Sigma (St. Louis, MO). 2,5-dihydroxybenzoic acid (2,5-DHB) was from Aldrich (St. Louis, MO). Tetrahydrofuran (THF) was from Riedel-de

Haen (Germany). An aly ti cal grade ace tone, meth a nol and to lu ene were sup plied by Lab-scan (Dub lin, Ire land).

Sample Preparation

Ad di tives and ma trix were dis solved in tetrahydrofuran (THF) to make *ca.* 10 mg/mL so lu tions. The sam ple so lu tion and the ma trix so lu tion were mixed 1:10 (v/v), and 1 μ L of the mix ing so lu tion was spot ted on the sam ple plate and dried in the air.

Polymer Extract

A weighted Poly eth yl ene sam ple (*ca.* 1 g) was refluxed with 100 mL of to lu ene in a 250-mL round-bottomed flask for 3 h. After cool ing, 20 mL of ace tone was added to pre cip i tate low mo lecu lar weight poly mer. The ex tract so lu tions were fil tered and dried by purg ing with ni tro gen gas.

Mass Spectrometry

MALDI was per formed on a PerSeptive Voy ager DE-RP time of flight (TOF) mass spec trom e ter (Framingham, MA, USA). This sys tem is equipped with a N₂ la ser (337 nm). For CID and PSD anal ysis, the pre cur sor ions were se lected us ing the timed ion se lec tor at a res o lu tion of about 50. For CID ex per i ments, the col li sion cell was filled with air un til the pres ure in the source cham ber reached ap prox i mately $3.5 \times 10^{-6} \sim 5 \times 10^{-6}$ torr. A Finnigan LCQ ion trap mass spec trom e ter (Finnigan MAT; San Joes, CA, USA) equipped with an electro spray ion iza tion source was used for anal ysis of

Table 1. Chemical Name and Trade Name

No.	Trade name	MW	Chemical name
1	Iragnox 245	586	Tri(ehtyl glycol)bis-3-(3- <i>t</i> -butyl-4-hydroxy-5-methylphenyl)propionate
2	Iragnox 259	638	1,6-Hexamethylene bis-(3,5-di- <i>t</i> -butyl-4-hydroxyhydrocinnamate)
3	Iragnox 1010	1176	Pentaerythritol tetrakis[3-(3,5-di- <i>t</i> -butyl-4-hydroxyphenyl)propionate]
4	Iragnox 1024	552	N,N-bis[1-oxo-3-(3,5-di- <i>t</i> -butyl-4-hydroxyphenyl)propane]hydrazine
5	Iragnox 1076	530	Octyldecyl 3-(3,5-di- <i>t</i> -butyl-4-hydroxyphenyl)propionate
6	Iragnox 1098	636	N,N-hexamethylene bis(3,5-di- <i>t</i> -butyl-4-hydroxyhydrocinnamide)
7	Iragnox 3114	783	Tris(3,5-di- <i>t</i> -butyl-4-hydroxybenzyl)isocyanurate
8	Naugard 524	646	Tris(2,4-dk- <i>t</i> -butylphenyl)phosphite
9	Tinuvin P	225	2-(2-hydroxy-5-methylphenyl)-2H-benzotriazole
10	Tinuvin 144	684	2- <i>t</i> -Butyl-2-(4-hydroxy-3,5-di- <i>t</i> -butylbenzyl)[bis(methy-2,2,6,6-tetramethyl-4-piperidinyl)]dipropionate
11	Tinuvin 320	323	2-(2-Hydroxy-3,5-di- <i>t</i> -butylphenyl)-2H-benzotriazole
12	Tinuvin 326	315	2-(3- <i>t</i> -Butyl-2-hydroxy-5-methylphenyl)-2H-5-chlorobenzotriazole
13	Tinuvin 328	351	2-(2-Hydroxy-3,5-di- <i>t</i> -amylphenyl)-2H-benzotriazole
14	Tinuvin 440	435	8-Acetyl-3-dodecyl-7,7,9,9-tetramethyl-1,3,8-triazaspiro(4,5)decane-2,4-dione
15	Tinuvin 770	480	Bis(2,2,6,6-tetramethyl-4-piperidinyl)sebacate
16	Tinuvin 622	≥ 3000	Poly-(N-b-hydroxyethyl-2,2,6,6-tetramethyl-4-hydroxy-piperdiyl succinate)
17	Chimassorb 994	≤ 2500	Poly-{6-[1,1,3,3-tetramethylbutyl]-imino}-1,3,5-triaine-2,4-diyl}{2-(2,2,6,6-tetramethylpiperinyl)-imino}



high molecular weight additives and the low energy CID experiments.

RESULTS AND DISCUSSION

MALDI mass spectra of polymer additives

In MALDI, sample is embedded in a low molecular

weight matrix that enhances sample ionization. Strong absorbance at the wavelength and good co-crystallization with sample are critical for ionization efficiency. Therefore, the success of this technique largely depends on the choice of the matrix. Different matrices, such as dithranol, ferric acid and 2,5-dihydroxybenzoic acids (2,5-DHB), have been used in the analysis of various synthetic polymers.²⁶⁻²⁸ These matrices were studied and the results showed that 2,5-DHB pro-

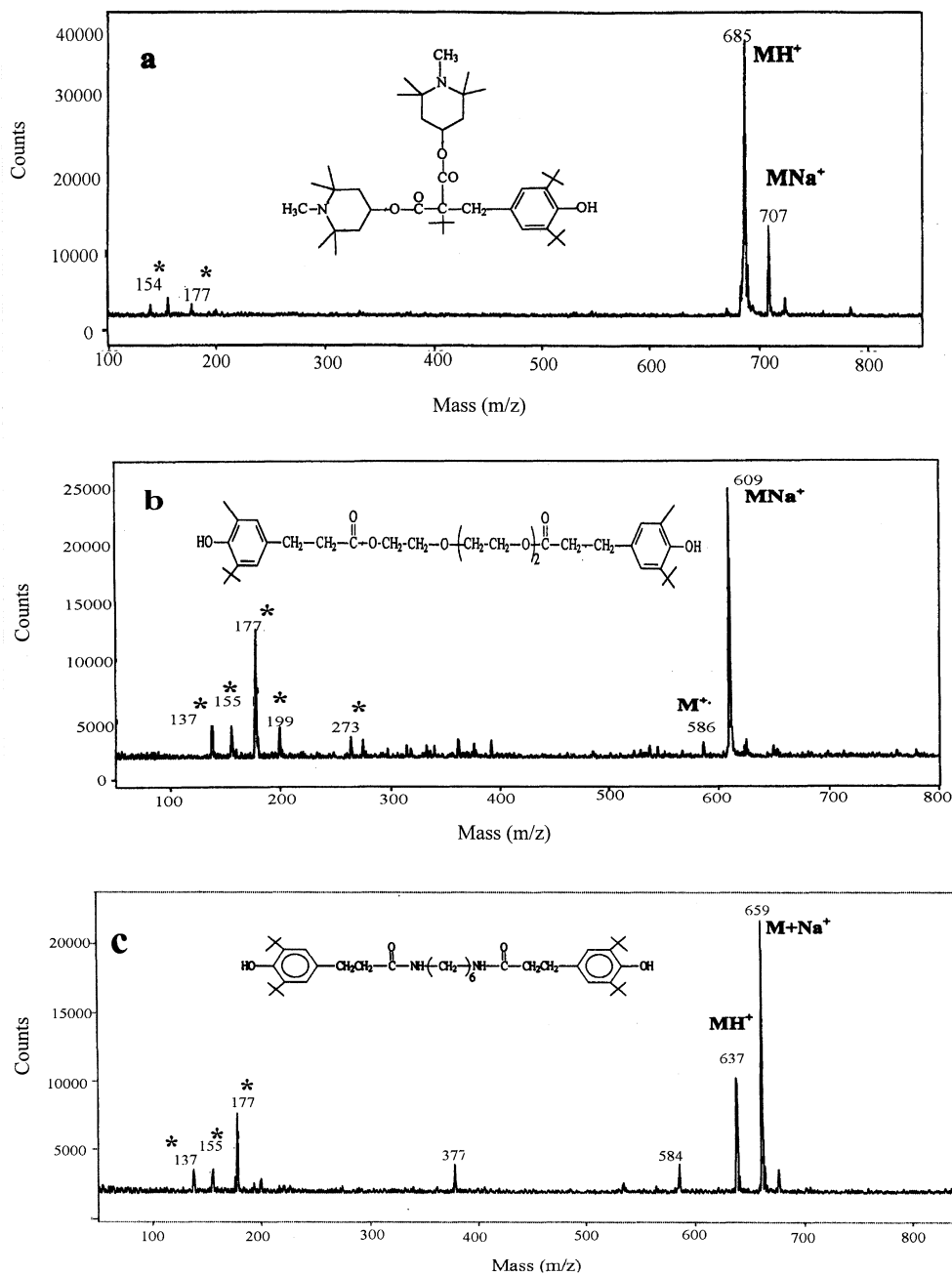


Fig. 1. (a) The MALDI mass spectrum of Tinuvin 144. (b) The MALDI mass spectrum of Irganox 245. (c) The MALDI mass spectrum of Irganox 1098 (*: matrix ion).

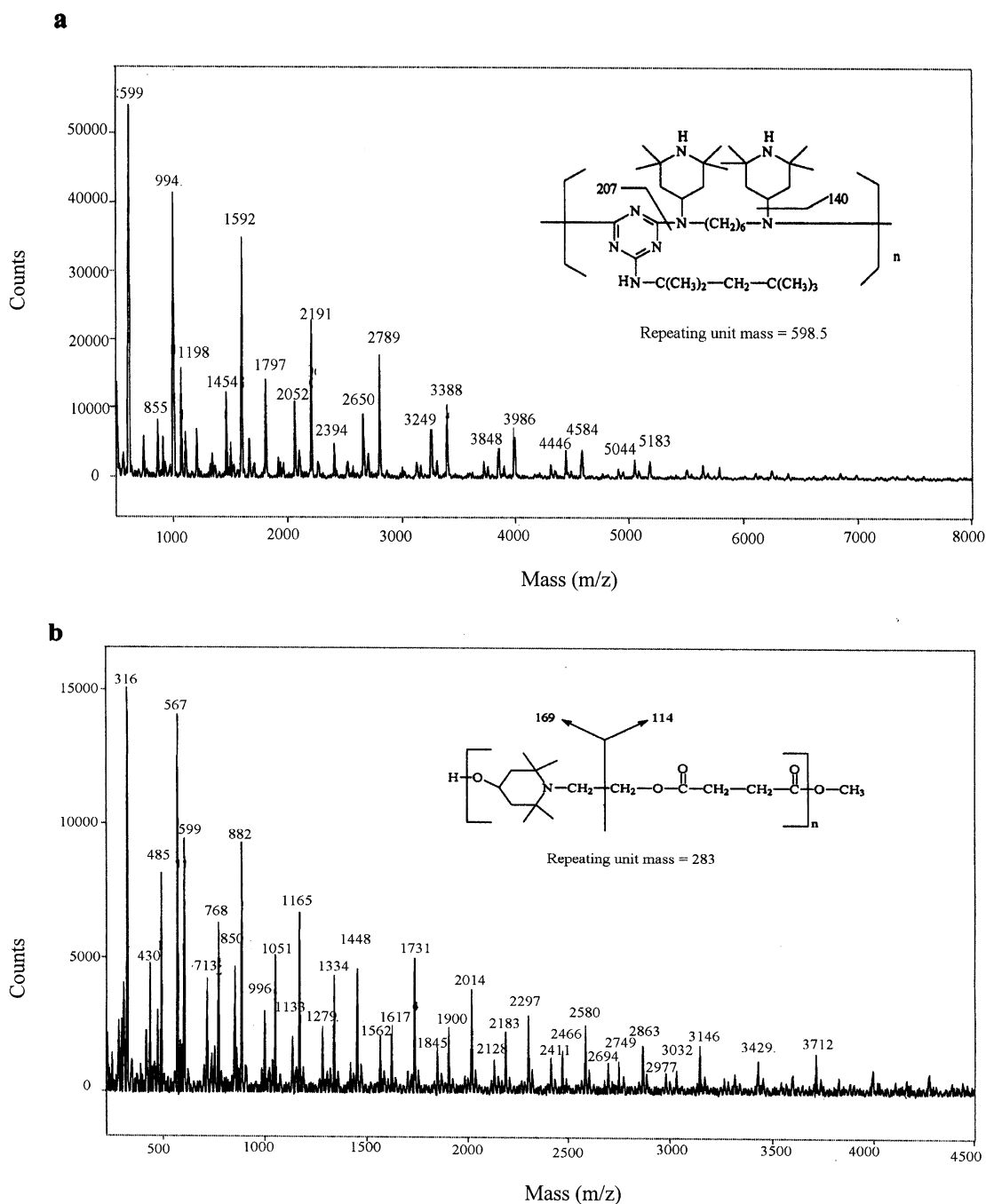


Fig. 2. The MALDI mass spec trum of (a) Chimassorb 944 (b) Tinuvin 622.

vided the most in tense analyte sig nals and the least abun dant ma trix ions. The pro duc tion of few ma trix ions would make the iden ti fi ca tion of ad di tives eas ier. Ac cord ingly, 2,5-DHB was used as the ma trix through out the study.

Con tin uous as well as de layed ex trac tions were stud ied for the anal y sis of poly mer ad di tives. The re sults showed that

there was less frag men ta tion in de layed ex trac tion. This is most likely due to the fact that the col li sion be tween the analyte ion and ma trix mol e cules is re duced greatly be cause of the de lay time. In the anal y sis of mix tures, such as the anal y sis of ad di tives in poly mer ex tract, it is eas ier to iden ti fy the ad di tives if there is less frag ment ions in the mass spec trum.

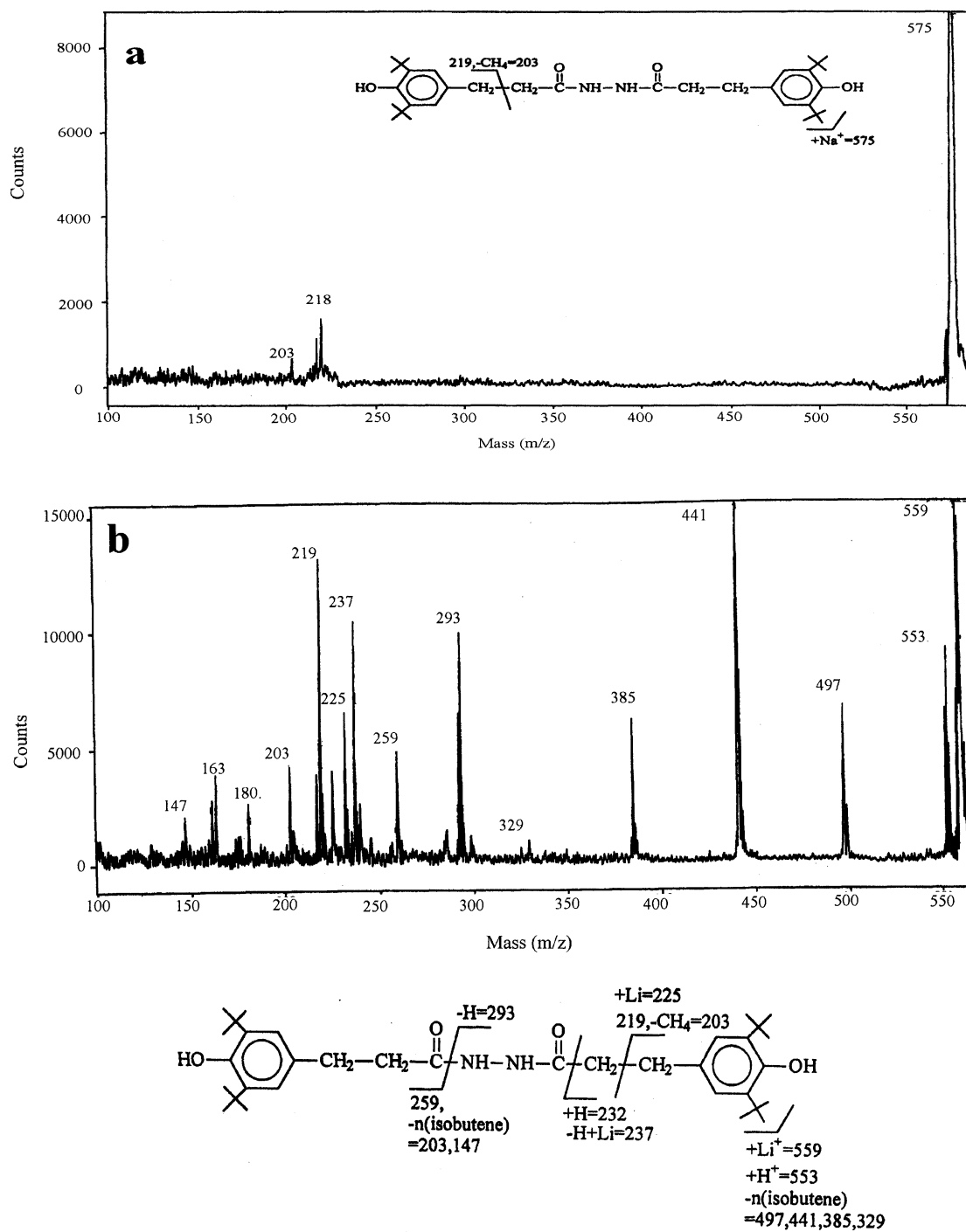


Fig. 3. The CID mass spectrum of (a) $[M+Na]^+$ ion (b) $[M+Li]^+$ ion of Irganox 1024.

Therefore, delayed extraction would make a better choice than continuous extraction. An other advantage of using delayed extraction is that the resolution is also higher than that of continuous extraction.

Additives listed in Table 1 were analyzed by MALDI. Typical MALDI mass spectra of polymer additives are shown in Fig. 1. The mass spectra of Tinuvin UV stabilizers were characterized with $[M+H]^+$ or $[M+H]^+/[M+Na]^+$ ion with es-

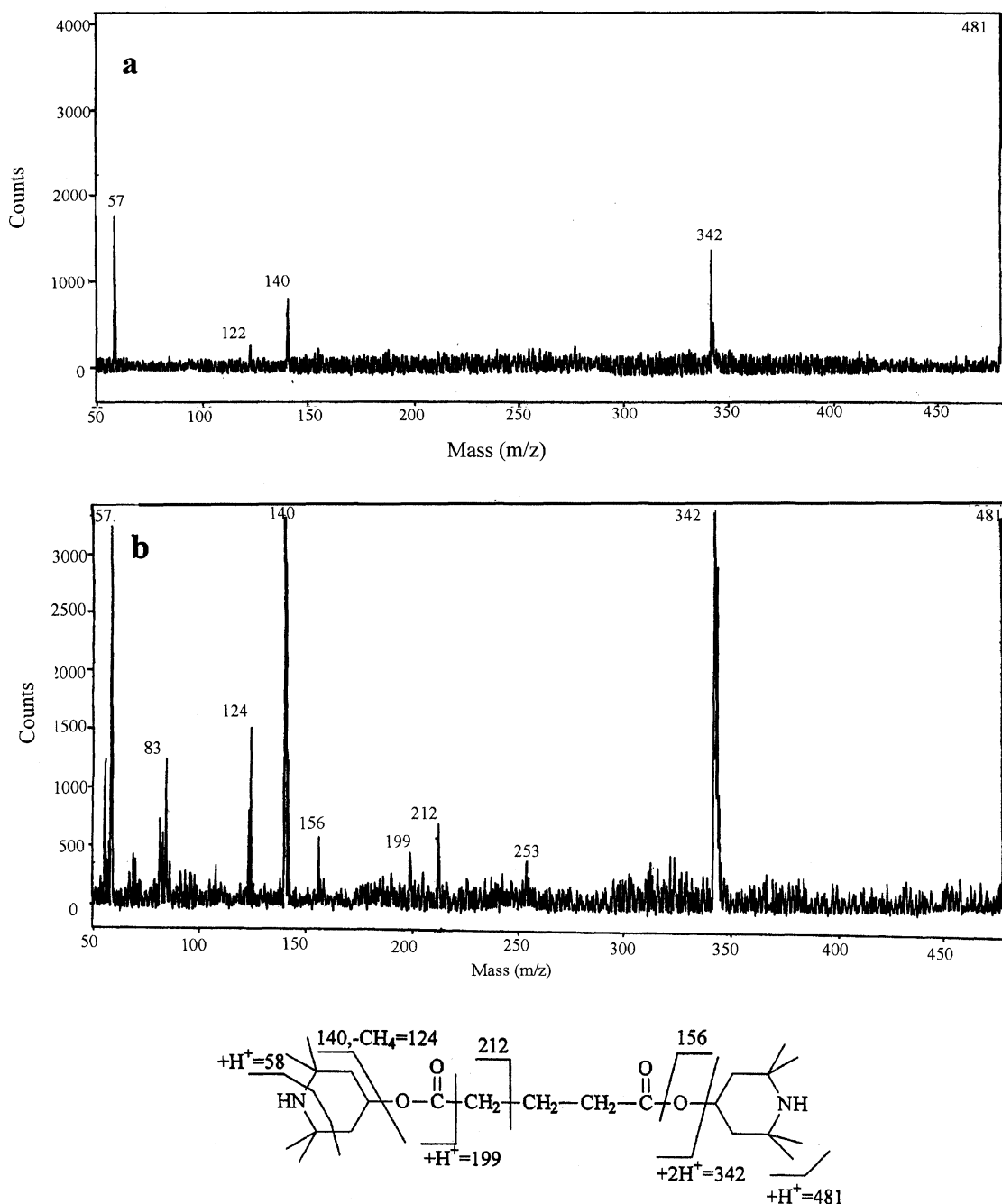


Fig. 4. (a) The PSD mass spectrum of Tinuvin 770 with continuous extraction. (b) The CID mass spectrum of Tinuvin 770 with delayed extraction.

essentially no fragmentation (Fig. 1a). Irganox antioxidants were characterized with $[M^{++}]/[M+Na]^+$ (Fig. 1b) or $[M+H]^+/[M+Na]^+$ ions (Fig. 1c) depending on the basicity of the molecules.

To reduce the loss during processing and services, there is an increase in the use of high molecular weight additives.

We have analyzed several high molecular weight additives by FAB, DCI, APCI, ESI and none of these techniques was capable of providing good quality mass spectrum. Because of its superb capability in the analysis of very large molecules, MALDI is expected to provide a better alternative to the analysis of these additives. The MALDI mass spectra of two high

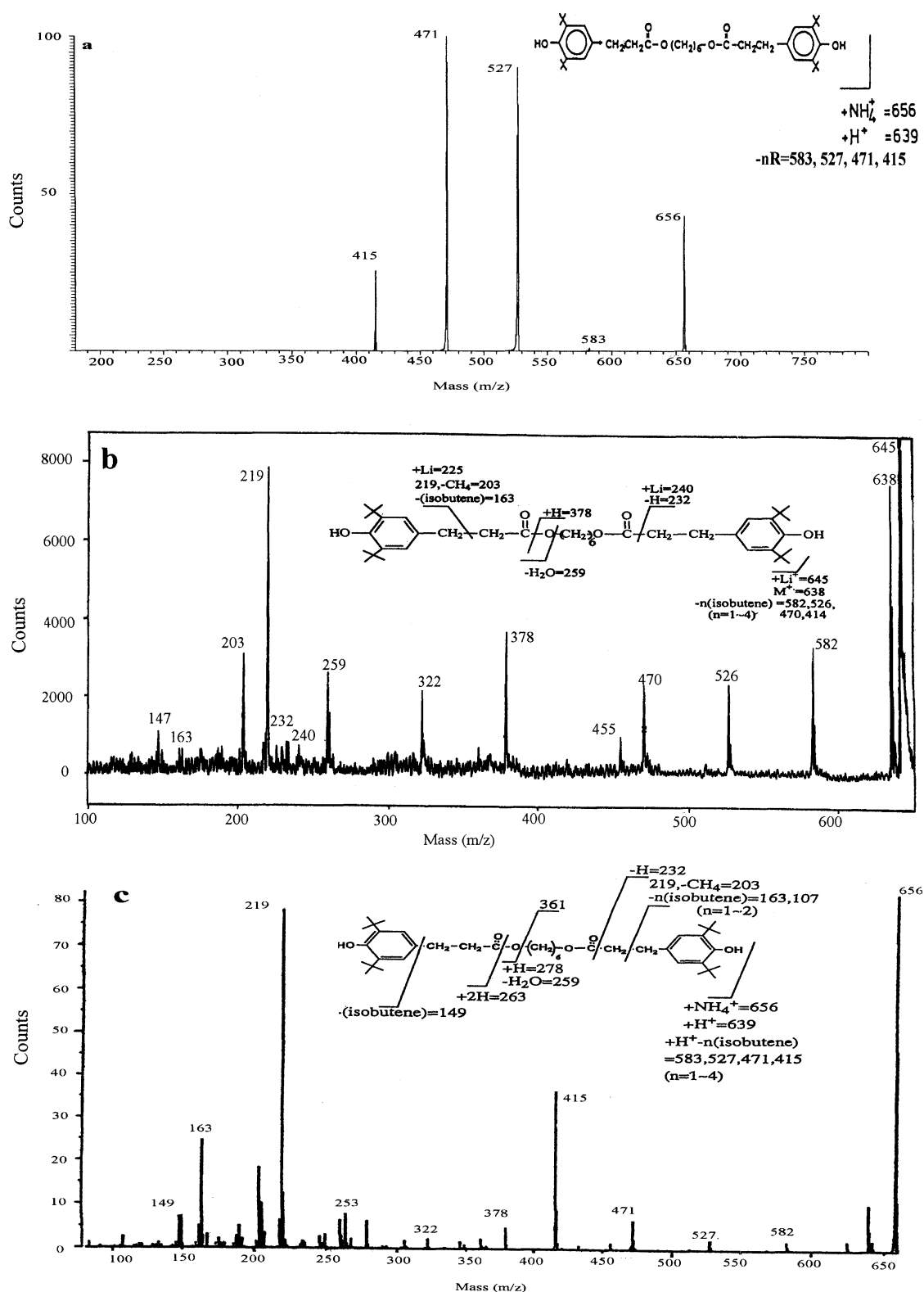


Fig. 5. (a) The CID mass spectrum of Irganox 259 $[M+NH_4]^+$ acquired with an ion trap mass spectrometer. (b) The CID mass spectrum of Irganox 259 $[M+Li]^+$ acquired with a MALDI-TOF mass spectrometer. (c) The CID mass spectrum of Irganox 259 $[M+NH_4]^+$ acquired with a sector instrument.

Table 2. The Product Ions of Polymer Additives

	Trade name (Formula)	Product ion
1	Irganox 245 C ₃₄ H ₅₀ O ₈	530, 474, 368, 263, 198, 190, 177, 161
2	Irganox 259 C ₄₀ H ₆₂ O ₆	582, 526, 470, 378, 322, 259, 219, 203
3	Irganox 1010 C ₇₃ H ₁₀₈ O ₁₂	1120, 1064, 1009, 952, 899, 259, 219, 203
4	Irganox 1024 C ₃₄ H ₅₂ N ₂ O ₄	497, 441, 385, 293, 259, 232, 237, 219, 203
5	Irganox 1076 C ₃₅ H ₆₂ O ₃	515, 475, 259, 232, 219, 203
6	Irganox 1098 C ₄₀ H ₆₄ N ₂ O ₄	581, 525, 469, 377, 321, 259, 219, 203, 163
7	Irganox 3114 C ₄₈ H ₆₉ N ₃ O ₆ Oxidized	565, 436, 354, 346, 260, 219, 203, 163
8	Naguard 524 C ₄₂ H ₆₃ O ₄ P	607, 551, 495, 459, 403
9	Tinuvin P C ₁₃ H ₁₁ N ₃ O	120, 107, 92
10	Tinuvin 144 C ₄₂ H ₇₂ N ₂ O ₅	532, 219, 154, 138, 123, 72
11	Tinuvin 320 C ₂₀ H ₂₅ N ₃ O	308, 268, 212, 57
12	Tinuvin 326 C ₁₇ H ₁₈ ClN ₃ O	300, 260, 57
13	Tinuvin 328 C ₂₂ H ₂₉ N ₃ O	336, 322, 282, 71
14	Tinuvin 440 C ₂₅ H ₄₅ N ₃ O ₃	418, 394, 377, 321, 110, 100, 58
15	Tinuvin 770 C ₂₈ H ₅₂ N ₂ O ₄	342, 212, 199, 140, 124, 58

molecular weight additives, Chimassorb 944 and Tinuvin 622, are shown in Fig. 2. In addition to the ions ($m/z = 599, 1198, 1797$) related to the intact molecules (repeating unit mass = 598 Da.), two major series of ions were also observed. The ions at m/z 994, 1592, 2191, 2789, 3388, 3986, 4585, 5183 corresponds to the loss of one triazine group from the oligomers. The ions at m/z 1454, 2052, 2650, 3249, 3848, 4446, and 5044 corresponds to the loss of one piperidyl group from the previous series. Tinuvin 622 stabilizer showed three major series of ions (Fig. 2b). The ions at m/z 316, 599, 882, 1165, 1448, 1731, 2014, 2297, 2580, 2863, 3146, 3429, and 3712 corresponds to the intact oligomers with repeating mass of 283 amu. The ions at m/z 485, 768, 1051, 1334, 1617, and 1900 correspond to the loss of a diester end group from the oligomers. The ions at m/z 713, 996, 1279, 1562, 1845 correspond to the loss of a piperidyl group from the linear oligomers.

MS/MS of polymer additives

The pseudomolecular ions in MALDI mass spectra provide tentative assignment of the additives present in polymer extracts. MS/MS of these pseudomolecular ions would provide much higher confidence in additive assignment. The study of additives by MS/MS indicated that, in comparison with $[M+H]^+$, it was rather difficult to fragment $[M+Na]^+$ ion (Fig. 3a). For many Irganox additives, $[M+Na]^+$ is either the only one or the major pseudomolecular ion, thus, it is difficult to obtain a useful product ion spectrum. It has recently been shown that lithium adduct fragments more extensively than sodium adduct.²⁸ Therefore, lithium salt was added to the matrix to promote the formation of $[M+Li]^+$ ion. As expected, many more fragments were observed in the CID of the $[M+Li]^+$ ion (Fig. 3b).

In our MALDI-TOF instrument, PSD and CID are the two means to acquire MS/MS spectra. Metastable decomposition during the flight through a MALDI-TOF instrument is referred to post-source decay. The resulting fragment ions possess different kinetic energies and therefore can be analyzed by stepwise decreasing the potential of the reflector. Although in some additives, such as Tinuvin 326, 328 and 440, the PSD spectra were essentially the same as the CID spectra; generally, CID produced more fragment ions than PSD as shown in Fig. 4. Accordingly, CID was used in the analysis of additives in polymer extract.

In the increasing of internal energy with CID, the acceleration voltage of the mass spectrometer plays an important role because the maximum energy, which can be converted into internal energy, is proportional to the translational energy of the precursor ion. The product ion spectra, obtained from three different instruments, of an antioxidant-Irganox 259 are shown in Fig. 5. As can be seen, many more product ions were detected in high-energy instruments (Fig. 5b, 5c) than in the low energy instrument (Fig. 5a). The CID spectra obtained from MALDI-TOF (for example, Fig. 5b) are similar but not quite the same as the spectra obtained from a sector instrument (for example, Fig. 5c). One possible reason is that the acceleration voltage of the TOF instrument (20 kV) is higher than that of the sector instrument (10 kV). Another possible explanation is the difference in the fragmentation pattern of $[M+Li]^+$ and $[M+H]^+$ ions. The major fragment ions of various UV-stabilizers and antioxidants are listed in Table 2.

The analysis of additives in polyethylene

The additives in polyethylenes were extracted with the procedures used in earlier DCI analysis¹⁴ and analyzed directly by MALDI. The MALDI mass spectrum of one sample



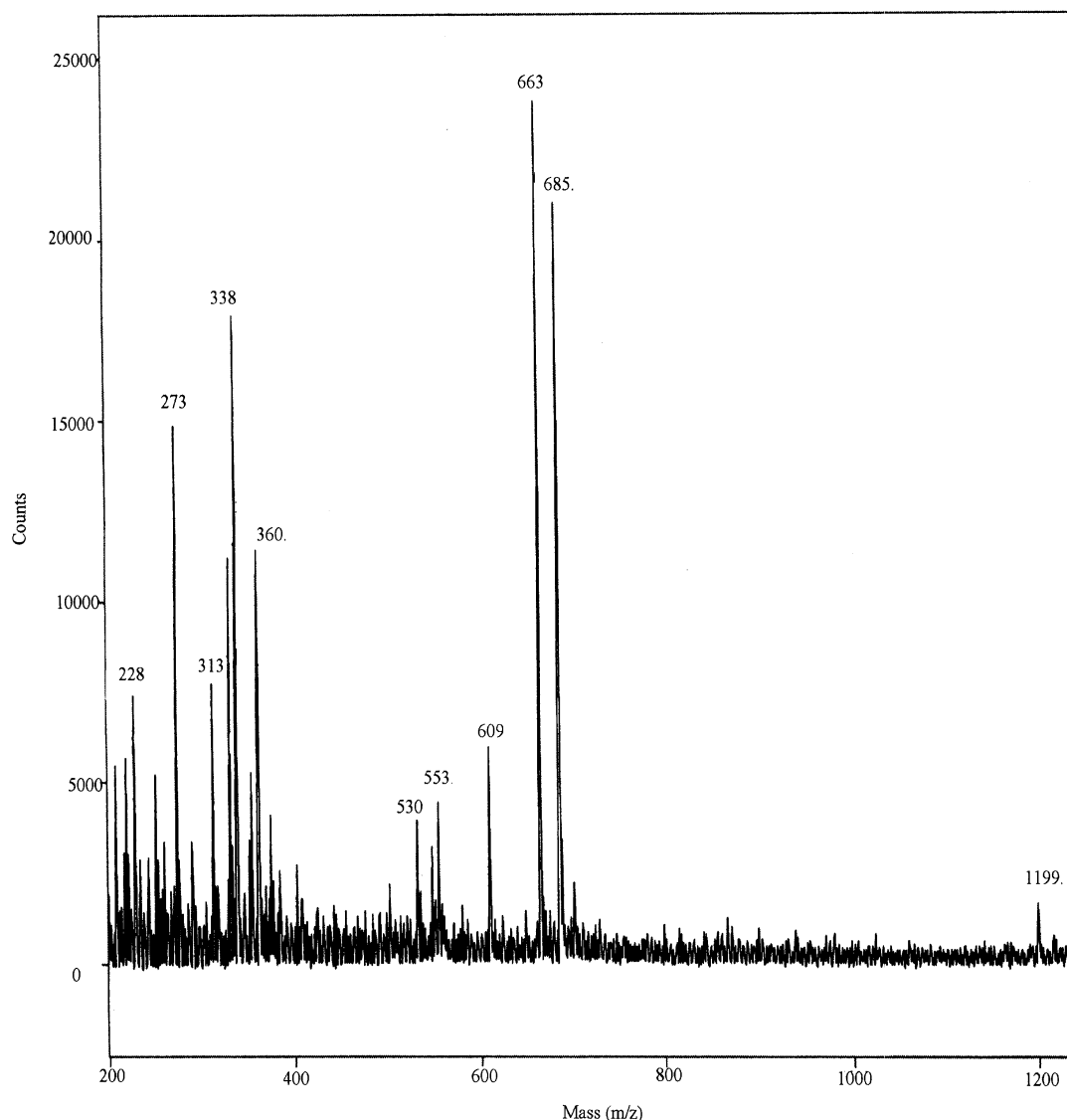


Fig. 6. MALDI mass spectrum of a polyethylene extract.

extract is shown in Fig. 6. Except the m/z 609 ion, the major ions at $m/z = 338/360$, $530/553$, $663/685$, and 1199 are tentatively as signed as $[M+H]^+/[M+Na]^+$ of Erucamide, $[M]^+/[M+Na]^+$ of Irganox 1076, $[M+H]^+/[M+Na]^+$ of oxidized Naugard 524 and $[M+Na]^+$ of Irganox 1010 respectively. In comparison with the analysis of additive standards, the spectrum of the extract was not very reproducible and only a few spots could produce the spec trum. One possible cause is that the low molecular weight polyethylene molecules are not precipitated completely when the low molecular weight polyethylenes in toluene are precipitated with methanol. Crystallization is believed to be critical for the quality of the MALDI spectrum and the existence of many low molecular

weight polyethylenes in the extract may interfere the process of crystallization. This problem was partially overcome with the use of acetone instead of methanol to precipitate the polyethylenes. The signals were found to be more reproducible if acetone was used in the precipitation of low molecular weight polyethylenes.

To improve the confidence of compound assignment, the pseudomolecular ions of the extract were subjected to CID analysis. Based on the CID spectra, the $m/z = 663$, 685 ions was as signed as the $[M+H]^+/[M+Na]^+$ ion of oxidized Naugard 524. The ion at $m/z = 338$, 360 was confirmed as the amide wax Erucamide. (Fig. 7) The signals at $530/553$ and 1199 were too weak to produce a useful product ion spec-

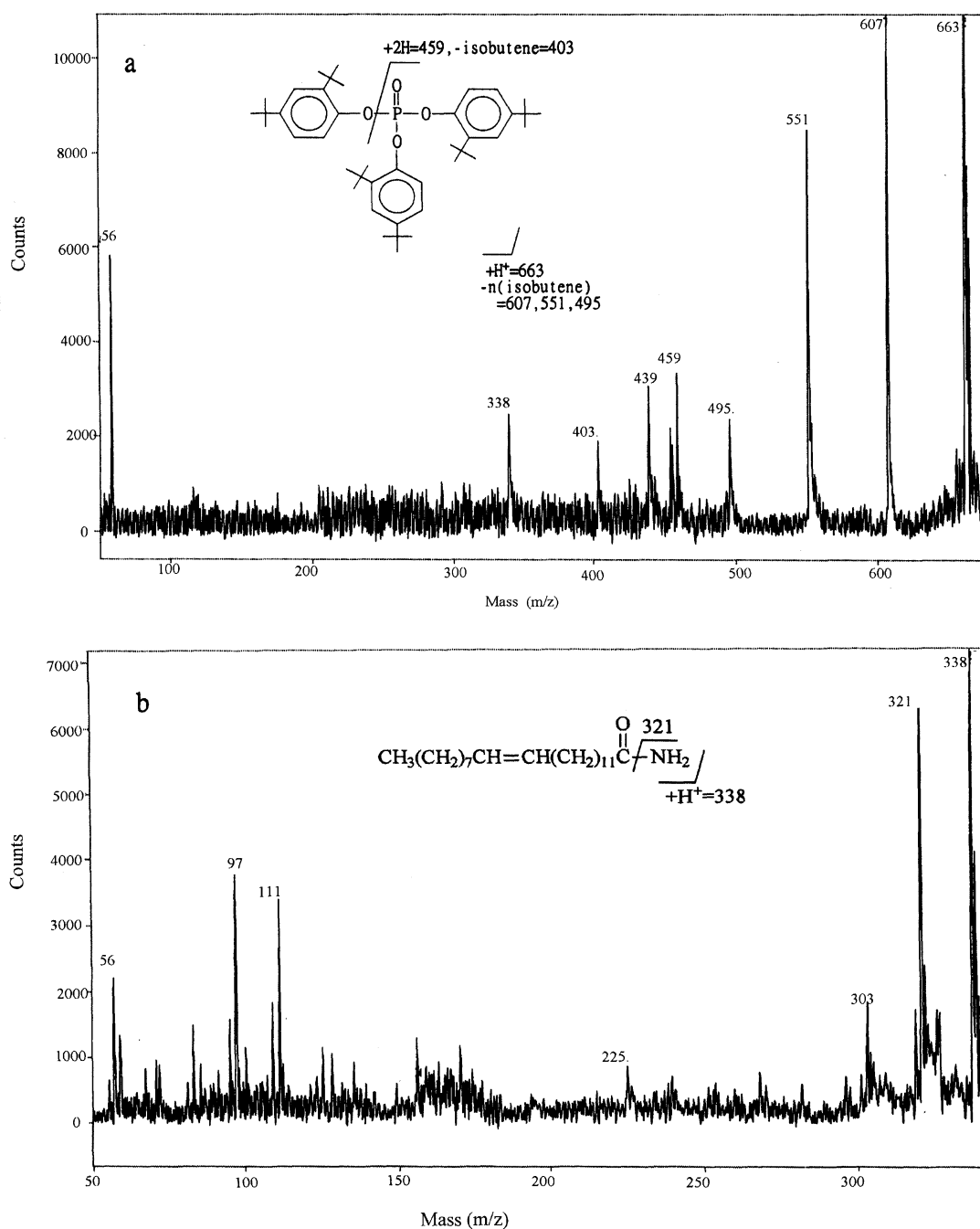


Fig. 7. The CID mass spectrum of (a) $m/z = 663$ ion and (b) $m/z = 338$ ion from the polymer extract.

trum.

CONCLUSION

MALDI was found to be a useful method for the identi-

fication of polymer additives in polyethylene. With the instrument operated under delayed extraction and using 2,5-dihydroxybenzoic acid (2,5-DHB) as the matrix, in general, only pseudomolecular ions were observed. Because of the observation of only pseudomolecular ions, tentative assignment can be easily made in the analysis of additives in

polymer extract. To confirm the tentative assignment, CID provides more information than PSD. While MALDI was found to be superior to other ionization techniques in the analysis of high molecular weight additives, sample preparation in MALDI was more critical in the analysis of real samples. Currently, we are looking for methods to separate the low molecular weight polyethylene molecules before the MALDI analysis.

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Key Words

Polyethylene; MALDI-TOF; Polymer additives; Collision induced dissociation (CID); Mass spectrometer.

REFERENCES

1. Vines, G. *New Sci.* **1995**, *147*, 22.
2. Floyd, T. R. *Chromatographia* **1988**, *25*, 791.
3. Carlson, D. W.; Hays, M. W.; Ransaw, H. C.; Mcfadden, R. S.; Altenau, A. G. *Anal. Chem.* **1971**, *43*, 1874.
4. Spell, H. L.; Eddy, R. D. *Anal. Chem.* **1960**, *32*, 1811.
5. Goe, G. R. *Soc. Plast. Eng. Tech.* **1997**, *23*, 496.
6. Floyd, T. R. *Chromatographia*. **1998**, *25*, 791.
7. Barcato, B.; Fantazzini, C.; Sevini, F. *J. Chromatogr.* **1991**, *553*, 415.
8. Munteanu, D.; Isfan, A.; Isfan, C.; Tincal, I. *Chromatographia* **1989**, *236*, 7.
9. Grosset, C.; Cantin, D.; Villet, A.; Alary, J. *Talanta*. **1990**, *37*, 301.
10. Schabron, J. F.; Fenska, L. E. *Anal. Chem.* **1990**, *52*, 1411.
11. Lay, J. O.; Barbara, Jr.; Miller, J. *Anal. Chem.* **1987**, *59*, 1323A.
12. Freeman, H. S.; Hao, Z.; Sokolowska-Gajda, J.; Breemen, R. B.; Le, J. C. *Dyes Pigments* **1991**, *16*, 317.
13. Chen, S. W.; Her, G. R. *Appl. Spectrosc.* **1993**, *47*, 844.
14. Juo, C. G.; Chen, S. W.; Her, G. R. *Anal. Chim. Acta* **1995**, *331*, 153.
15. Lattimer, R. P. *J. Anal. Appl. Pyrol.* **1993**, *26*, 65.
16. Asamoto, B.; Young, J. R.; Citerin, R. J. *Anal. Chem.* **1990**, *62*, 61.
17. Harad, K. I.; Masuda K.; Suzuki, M. *Biological Mass spectrom.* **1991**, *20*, 522.
18. Johlman, C. L.; Wilkin, C. L.; Hogan, J. D.; Donovan, T. L.; Youssefi, M. J. *Anal. Chem.* **1990**, *62*, 1167.
19. Zhan Qiao; Zenobi Renato; Wright, S. J.; Langridge-Smith, P. R. R. *Macromoleules* **1996**, *29*, 7865.
20. Wright, S. J.; Dale, M. J.; Langridge-Smith, P. R. R.; Zhan Qiao; Zenobi, Renato *Anal. Chem.* **1996**, *68*, 3585.
21. Jackson, A. T.; Buzy, Armelle; Jennings, K. R.; Scrivens, J. H. *Eur. Mass Spectrom.* **1996**, *2*, 115.
22. Carrott, M. J.; Jones, D. C.; Davidson, G. *Analyst* **1998**, *123*, 1827.
23. Yu, K.; Block, E.; Balogh, M. *LC-GC* **2000**, *18*, 162.
24. Jackson, A. T.; Jennings, K. R.; Scrivens, J. H. *Rapid Commun. Mass spectrometry* **1996**, *10*, 1449.
25. Kornfeld, R. A.; Trengove, R. D. *Proceeding of the 45th ASMS Conference on Mass Spectrometry and Allied Topics* **1997**, 419.
26. Bahr, U.; Deppe, A.; Karas, M.; Hillenkamp, F. *Anal. Chem.* **1992**, *64*, 2866.
27. Kim, S. H.; Shin, C. M.; Yoo, Y. S. *Proceeding of the 45th ASMS Conference on Mass Spectrometry and Allied Topics* **1997**, 1274.
28. Volmer, D. A.; Lock, C. M. *Rapid Commun. Mass Spectrom.* **1998**, *12*, 157.

