

行政院國家科學委員會專題研究計畫第一年進度報告

狼尾草染色體組分子分析及孤雌生殖性狀遺傳定位

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一、中文摘要

狼尾草為本省目前主要之芻料作物，其栽培面積僅次於盤固草，但其單位產量卻高出盤固草一倍，雖營養價值欠佳，但許多文獻指出利用其近源種 - 珍珠粟與之雜交可組合出兼具狼尾草高產、生長旺盛、多年生性狀及珍珠粟優良品質之特性。因此自民國七十年起台灣省畜產試驗所即著手進行狼尾草與珍珠粟之雜交育種工作，釋出數個優良營養系，但因狼尾草有性生殖之特性使得以種子繁殖之後代產生性狀分離之現象，無法保持原基因型之優良組合，也因此使得狼尾草無法利用種子進行大面積之推廣。孤雌生殖此一以種子行無性繁殖之生殖現象，普遍存在狼尾草之近源種間，若能將此性狀導入優良狼尾草品系將有助於推廣工作之進行。惟孤雌生殖一直是牧草育種的障礙，主要是因為其鑑定不易且易受環境影響，因此利用分子標記輔助具孤雌生殖特徵之牧草進行育種為目前之趨勢，惟進展不多。大多數牧草所具有之多倍體性則為另一障礙，同時亦是遺傳研究的一大阻力。本計畫利用比較遺傳圖譜構建法，以近源種之現有遺傳圖譜為指引，以保守之 RFLP 分子標記為優先篩檢對象，配合狼尾草與珍珠粟雜交具孤雌生殖特徵之分離族群，將可縮短遺傳圖譜構建之時間，同時亦提供了將孤雌生殖性狀定位於遺傳圖譜上的機會。

關鍵詞：芻料作物、狼尾草、孤雌生殖、遺傳圖譜、比較遺傳圖譜構建法、多倍體性、牧草育種。

Abstract

Napier grass is one of the major forage crops in Taiwan. The acreage is second to the one of pangola grass, but the yielding ability is about twice of the one of pangola grass. Although the nutritive value is generally poor in napier grass, the inter-specific hybrids between napier grass and pearl millet, which considered to have better nutritive value, did show the ability to combine both species' desired characteristics. There were several clones released from such inter-specific crosses by the National Taiwan Animal Science Experiment Institution since 1981. Napier grass is a sexually reproduced perennial grass. In order to preserve the genetic composition of a genotype, napier grass used to be propagated through cloning of vegetative organ, which is the main problem in expanding its acreage. The introduction of apomixis, which is a process of cloning genotype through seed production, from related species within Pennisetum family may help the expansion of napier grass. The benefit of apomixis in fixing genotype is obvious, but the difficulties in identification and the instability of expression are also associated with apomixis. In order to overcome these problems, the techniques of utilizing DNA markers in breeding apomictic species are developing. The progress is hindered by the polyploidy nature of the most of the apomictic species. The purpose of this project is to simultaneously construct molecular genetic maps of napier grass, pearl millet and *Pennisetum squamulatum* from a apomictic/sexual segregating population of a three-way cross between (napier grass x pearl

millet) x *Pennisetum squamulatum*. The conservative RFLP markers will be chosen from related species with well established RFLP map and used in constructing the molecular genetic map. The principle of comparative mapping will be served as a guide line in choosing conservative markers, it will shorten the time for constructing map by reducing the time needed for surveying DNA probes. With the construction of molecular genetic map, the apomixis trait will be expected to be mapped.

Keywords: Forage crop, napier grass, *Pennisetum purpureum* L., apomixis, genetic map, comparative mapping, polyploidy, breeding

二、緣由與目的

配合台灣農業現階段及未來轉型趨勢，在酪農業、低投入農業、能源作物及生態保育方面，牧草之利用應有發展的空間。但由於大多數牧草物種具有多倍體性、多年生及異花受粉之特性，向為基礎遺傳研究避免之材料，再加上芻料作物並非人類直接利用之作物，向為各國政府忽略之目標因此與牧草相關之研究項目及成就也就遙遙落後於其他作物 (Sleper *et al.*, 1987)。此外孤雌生殖 (apomixis) 現象普遍存在於禾本科牧草物種，嚴重妨礙雜交分離族群之獲得，就遺傳變異而言，一度被認為是演化上的死胡同 (Darlington 1958, Stebbins 1950)，也影響了牧草育種工作的進展。雖然另外有一派學者認為孤雌生殖在演化上不僅不是死胡同，由分類上的研究，顯示孤雌生殖是一個極具彈性的遺傳系統，提供植物發展多倍體以適應環境的機制 (Bashaw *et al.* 1970, Gustafsson 1946)。但雙方均無法取得足夠的證據，以致至今無法獲得共同可認同的結論。

孤雌生殖造成子代的不分離性及多倍體性造成的遺傳複雜性是牧草遺傳育種研究的兩大難題。傳統石蠟切片利用連續二度空間切片，由人為想像連接而成的立體三度

空間結構來判定孤雌生殖性狀的窘境，雖因組織透明化技術 (clearing technique) 的發展 (Sheerwood *et al.* 1980) 可對子房或胚囊進行整器官製片 (whole mount)，配合相位差 (phase contrast microscopy) 或干涉顯微技術 (interference microscopy) 進行實體的立體三度空間觀察獲得舒解，但選取組織材料，製作標本及顯微鏡鏡檢仍然相當費時，同時也需要累積足夠的經驗才能減少誤判的比例。雖然如此，配合有性生殖與孤雌生殖植株雜交後代產生的分離族群，孤雌生殖性狀的遺傳行為研究已有不少成果 (Sheerwood *et al.* 1994)，但尚未見應用於大規模育種族群篩選的實例。雖然孤雌生殖性狀的遺傳行為研究已有不少成果，但如加入多倍體性所可能造成的複雜子代分離比例此一因子，則孤雌生殖性狀的遺傳行為的研究仍處於渾沌為清的狀態。研究孤雌生殖性狀的價值除了可瞭解物種演化分類及組織分化之學術意義外，就生態保育方面的意義而言，則可提供研究物種歧異的參考，就牧草育種而言，可提供複合品種或純系品種育種目標之選定，就作物育種而言，則可提供轉移時的參考。

多倍體性的遺傳育種研究及應用雖有不少成就，如六倍體小麥，三倍體西瓜的育種，但在牧草上則仍為處女地，除孤雌生殖性狀外，異花受粉植物常見的自交弱勢則為另一重要因素 (Wang, 1996)。因為自交弱勢而造成純質結合體之取得不易，而妨礙了遺傳育種之研究。

近年分子遺傳學的快速進步，提供了許多建立遺傳圖譜的新工具，如 RFLP, RAPD, AFLP, RLGS, micro-satellite 等，主要作物如水稻、玉米、高粱、棉花、甘蔗、小米、小麥、大麥之 RFLP 圖譜相繼發表 (Chittenden *et al.* 1994; Liu *et al.* 1994; Reighish *et al.*; Da Silva *et al.* 1993; Gardiner *et al.* 1993; Anderson *et al.* 1992; Graner *et al.* 1991; McCouch *et al.* 1988)。其中以甘蔗 RFLP 圖譜之建構最複雜，除了 RFLP 分子標記之連鎖分析外，還

需判定染色體組之組成以及同源染色體組之分群。牧草物種除了屬二倍體之珍珠粟(小米)已有圖譜發表外,同屬 *Pennisetum* 屬內之巴弗草 (buffelgrass, *Pennisetum ciliare* (L.) Link syn. *Cenchrus ciliaris* L.) 的遺傳圖譜之構建則剛起步 (Wang, 1996)。

狼尾草為本省目前主要芻料作物之一,其栽培面積僅次於盤固草,但其單位產量卻高出盤固草一倍,若能提高其品質,將可降低酪農戶生產成本,增加收益。狼尾草為四倍體、多年生之有性生殖禾草,台灣畜產試驗所利用其與珍珠粟雜交之種間後代選拔出數個有性生殖營養系,其缺點是無法利用種子繁殖,因此在期望利用孤雌生殖固定基因型而能利用種子繁殖的目的下,又進行有性狼尾草與珍珠粟雜交後代與孤雌生殖性狀供應親 (*Pennisetum squamulatum* L.) 之三向雜交 (成, 1994)。

本計畫之目的在構建狼尾草之 RFLP 圖譜以供育種之用。RFLP 圖譜應用在育種計畫中,除了可提供簡單遺傳性狀之連鎖分子標記以輔助篩選基因型外,更可提供數量性狀之連鎖分子標記以輔助數量性狀之改良,可大幅提高育種效率。DNA 分析技術的自動化將可進一步提高 RFLP 圖譜的應用價值,以本省精簡的牧草研究人員編制,構建 RFLP 圖譜應有其價值。

三、目前執行進度

本計畫第一年預定完成近源物種的 RFLP 片段庫的收集及 DNA 樣品的收集,並建立所有相關的實驗操作程序

目前經聯絡已經確定並將陸續於 87 年下半年寄到者,包括 University of Missouri RFLP laboratory 所建立之玉米 RFLP 核心探針組、The John Innes Centre, Norwich Research Park, Colney, NORWICH. NR4 7UH. UK. 所建立之小米 RFLP 核心探針組,及台大農藝系育種研究室所建立之高梁 cDNA library,此外 Cornell University 的水稻 RFLP 核心探針組以及

Rice Genome Research Program (RGP) at STAFF Institute in Tsukuba, Japan 的水稻 RFLP 核心探針組目前仍在接洽中。

目前已取得構建圖譜所需之分離族群,族群個體數為 87 個植株。

DNA 抽取步驟以仿自 Buffelgrass 之步驟,抽出率可達 3%。步驟如下:

Materials

Extraction buffer

100 mM Tris, pH 8.0
50 mM EDTA, pH 8.0
500 mM NaCl
1.25% SDS (W/V)
add 3.8 g of NaHSO₃ (Sodium bisulfite) per liter to extraction buffer before use.

5 M KOAc (Potassium Acetate)

Purifying buffer

70 % Ethanol with 0.3 M Sodium Acetate

70 % Ethanol

TE buffer

10 mM Tris.Cl, pH 8.0
1 mM EDTA, pH 8.0

Procedure

1. Add c.a. 2.5 g of ground tissue into 50 ml centrifuge tube.
2. Add 30 ml of extraction to each tube, mix well by vortexing. Incubate in 65 °C waterbath for 1 hour. Invert tube every 15 minutes during incubation.
3. Add 9 ml of 5M KOAc to each tube. Invert tube gently until thoroughly mixed. Leave tube on ice for 20 minutes. Keep sample cold, on ice or refrigerated, beyond this step.
4. Centrifuge 20 minutes at 2800 g, 4 °C.
5. Label a new tube and dispense 20 ml of cold (-20 °C) isopropanol to the tube. Pour supernatant to the new tube through a layer of miracloth. Do not mix the two phases and leave tube untouched at -20 °C for at least 1 hour.

6. Hook out DNA.
7. Add 25 ml of purifying buffer to soak the DNA pellet overnight in 4 °C.
8. Pour off the purifying buffer.
9. Add 5 ml of cold (-20 °C) 70 % ethanol soak DNA pellet for at least 5 minutes.
10. Pour off 70 % ethanol. Air dry pellet until no odor of ethanol can be detected.
11. Add 300-500 µl of TE buffer to each tube. Leave tube in 37 °C waterbath until DNA dissolves in the buffer. Transfer to 1.5 ml micro-centrifuge tube. Store at -20 °C.

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