



## Ammonium in relation to proline accumulation in detached rice leaves

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### Abstract

Ammonium accumulation in relation to proline accumulation in detached rice leaves under stress conditions was investigated. Ammonium accumulation in dark-treated detached rice leaves preceded proline accumulation. Ammonium accumulation caused by water stress coincided closely with proline accumulation in detached rice leaves. Exogenous  $\text{NH}_4\text{Cl}$  and methionine sulfoximine (MSO), which caused an accumulation of ammonium in detached rice leaves, increased proline content. It was found that proline in  $\text{NH}_4\text{Cl}$ - or MSO-treated rice leaves is less utilized than in water-treated rice leaves (controls). These results are in agreement with the observation that a decrease in proline utilization contributes to the accumulation of proline in dark-treated and water stressed rice leaves. Although ammonium content increased in Cd- and Cu-treated rice leaves, the increase in ammonium content was only observed after the increase in proline content.

*Abbreviations:* MSO – methionine sulfoximine; WS – water stress

### 1. Introduction

Proline has been shown to accumulate in plants subjected to water stress and exposed to excess Cd and excess Cu [1, 2, 9, 12, 26, 27, 29, 32]. Previously, we have reported that proline accumulated in detached rice leaves during dark-induced senescence [10, 31].

The ammonium ion is a central intermediate in the metabolism of nitrogen in plants. Glutamine synthetase plays a crucial role in the assimilation of ammonium [16]. It has been shown that water stress results in a decline in the activity of glutamine synthetase in leaves [4, 18]. Glutamine synthetase activity is also known to decrease during dark-induced senescence of leaves [7, 13, 20, 28]. We have demonstrated that excess Cu decreased the activity of glutamine synthetase in detached rice leaves [7]. A decline in glutamine synthetase activity in leaves during water stress, dark-induced senescence or when exposed to heavy metals may result, at least in part, in an accumulation of ammonium in leaves. In fact, ammonium has been shown to accumulate in leaves subjected to

water stress, when exposed to excess Cu, and during dark-induced senescence [7, 8, 18, 22, 30].

Accumulation of putrescine has been reported in response to environmental stresses [6]. Several lines of evidence suggest that putrescine biosynthesis is linked to the assimilation of ammonium nutrition in both stressed and nonstressed plant tissues. Le Rudulier and Goas [15] showed that ammonium nutrition produced higher tissue putrescine content than did nitrate feeding. Similarly, Klein et al. [14] reported that  $\text{K}^+$ -deficient peas utilizing ammonium as a nitrogen source accumulated putrescine to a greater extent than those maintained on nitrate. Rabe and Lovatt [24] have demonstrated that increased ammonium content in  $\text{PO}_4$ -deficient citrus are accompanied by increased arginine biosynthesis. Rabe [23] suggested that putrescine and arginine accumulating during environmental stress conditions might serve as an ammonium detoxification mechanism. By extension, it seems likely that stress-induced accumulation of proline could represent a manifestation of such an ammonium detoxification mechanism. To understand the metabolic

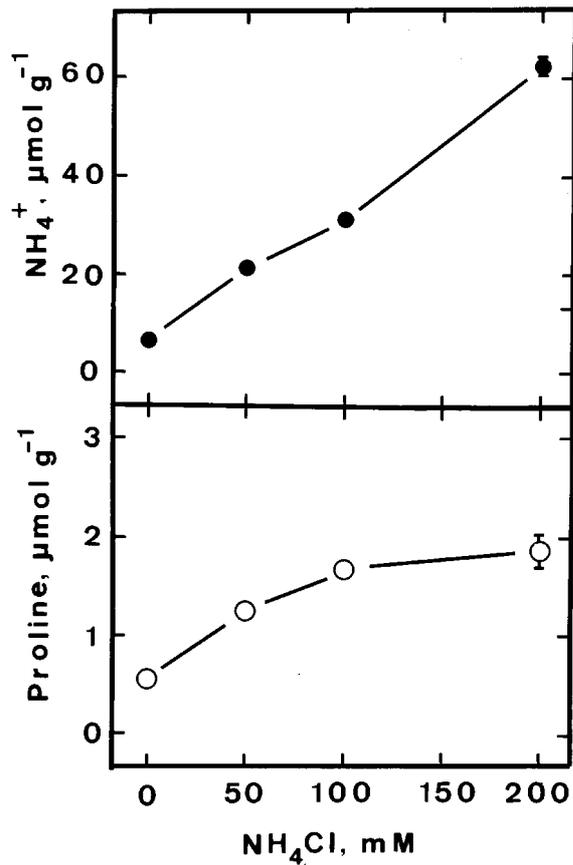


Figure 1. Influence of  $\text{NH}_4\text{Cl}$  on ammonium and proline contents in detached rice leaves. All measurements were made 12 h after treatment in the light. Vertical bars represent standard errors ( $n = 4$ ).

and cellular phenomena that lead to proline accumulation, in detached rice leaves induced by water stress, heavy metals and during dark-induced senescence, we have tried to determine the relation, if any, between ammonium and proline accumulations. This paper reports more detailed time-course measurements along with some dose-response experiments, designed to describe the relation between ammonium and proline accumulations in detached rice leaves.

## 2. Materials and methods

Rice (*Oryza sativa* cv. Taichung Native 1) was cultured on a stainless net floating on half-strength Johnson's modified nutrient solution (pH 4.2) in a 500 ml beaker [11]. The nutrient solution was replaced every three days. Rice seedlings were grown for 12 d in a greenhouse, where natural light was provided and

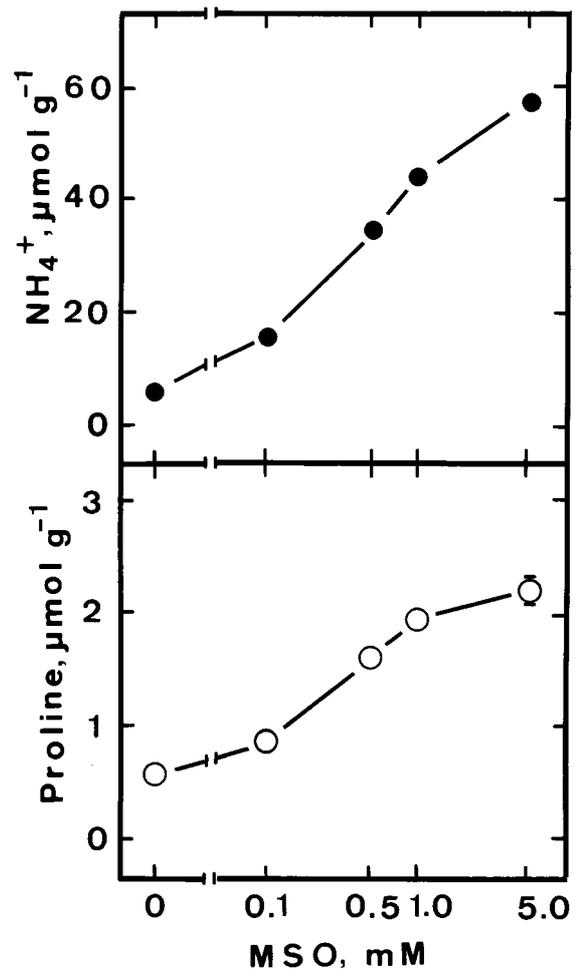


Figure 2. Influence of MSO on ammonium and proline contents in detached rice leaves. All measurements were made 12 h after treatment in the light. Vertical bars represent standard errors ( $n = 4$ ).

the temperature was controlled at  $30^\circ\text{C}$  during the day and at  $25^\circ\text{C}$  at night. The apical 3 cm segments excised from the third leaves of 12-d-old seedlings were used. A group of 10 segments was floated in a Petri dish containing 10 ml of test solutions or water (controls). Incubation was carried out at  $27^\circ\text{C}$  in light ( $40 \mu\text{mol quanta m}^{-2} \text{s}^{-1}$ ) or in darkness. Water stress was applied by a previously described method [12]. Briefly, leaf segments were exposed to the vapour above a solution of 0.5 M NaCl to decrease air humidity.

Proline was extracted and its concentration determined by the method of Bates et al. [3]. Leaf segments were homogenized with 3% sulfosalicylic acid and the homogenate was centrifuged. The supernatant was treated with acetic acid and acid ninhydrin, boiled for

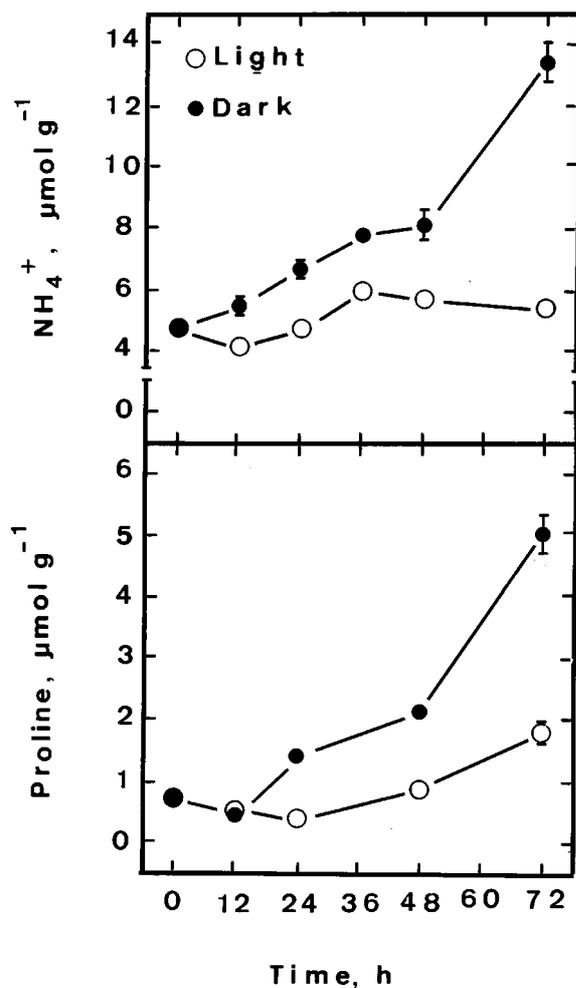


Figure 3. Time courses of proline and ammonium contents in detached rice leaves incubated in distilled water under dark and light conditions. Vertical bars represent standard errors ( $n = 4$ ).

1 h and then absorbance at 520 nm was determined. Amounts of proline are expressed as  $\mu\text{mol g}^{-1}$  initial fresh weight.

Ammonium was extracted by homogenizing leaf segments in 0.3 mM sulphuric acid (pH 3.5). The homogenate was centrifuged for 10 min at 39,000 g and the supernatant was used for determination of ammonium as described previously [17]. Ammonium content was expressed as  $\mu\text{mol g}^{-1}$  initial fresh weight.

### 3. Results and discussion

Figure 1 shows the effect of  $\text{NH}_4\text{Cl}$  on ammonium and proline contents in detached rice leaves in the

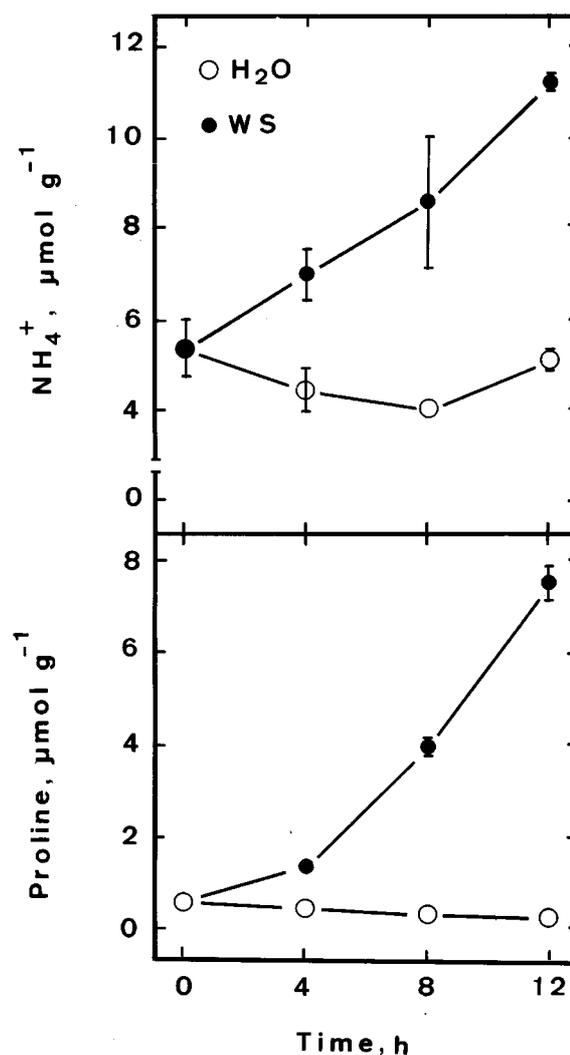


Figure 4. Time courses of proline and ammonium contents in detached rice leaves incubated in distilled water and water stress (WS) conditions. Vertical bars represent standard errors ( $n = 4$ ).

light. It is clear that the increase in the ammonium content in  $\text{NH}_4\text{Cl}$ -treated detached rice leaves was associated with the increase in proline content. MSO is a structural analogue of glutamate, and serves as an irreversible inhibitor of glutamine synthetase [25]. There is evidence that the addition of MSO results in an accumulation of ammonium [5, 21]. To characterize further the relationship between ammonium and proline contents in detached rice leaves, detached rice leaves were incubated in the presence of various concentrations of MSO. As indicated in Figure 2, MSO increased endogenous ammonium and proline contents.

Table 1. Proline content in ornithine-pretreated detached rice leaves incubated in distilled water,  $\text{NH}_4\text{Cl}$  and MSO

Treatment	Proline, $\mu\text{mol g}^{-1}$
Orn, 3h	$18.7 \pm 1.3$
Orn, 3h $\rightarrow$ $\text{H}_2\text{O}$ , 8h	$5.7 \pm 0.4$
Orn, 3h $\rightarrow$ $\text{NH}_4\text{Cl}$ , 8h	$10.6 \pm 0.7$
Orn, 3h $\rightarrow$ MSO, 8h	$9.1 \pm 0.7$

Detached rice leaves were pretreated with 50 mM ornithine (Orn) for 3 h in the light and then incubated in distilled water,  $\text{NH}_4\text{Cl}$  (200 mM), or MSO (1 mM) for 8 h in the light. Means  $\pm$  SE (n = 4).

Figure 3 shows the change of proline content in detached rice leaves incubated in distilled water in darkness and in the light. In darkness, the amount of proline in detached rice leaves did not increase until 12 h after incubation, but increased about 2-, 2.5- and 6-fold at 12, 48, and 72 h, respectively, which confirmed our previous results [10, 31]. However, the amount of proline in detached rice leaves, incubated in the light remained relatively unchanged during 48 h, but subsequently increased slightly. In detached rice leaves, it is apparent that proline accumulates more in darkness than in the light. It is expected that the ammonium content is greater in darkness than in the light. As indicated in Figure 3, this is indeed the case. An increase in the ammonium content of dark-treated detached rice leaves was found to precede the increase in proline contents.

It has long been recognized that water stress is effective in increasing proline content in detached rice leaves [12]. The effect of water stress on proline content in relation to ammonium content is presented in Figure 4. Water stress effectively increased proline and ammonium content.

Recently, we have shown that a decrease in proline utilization contributes to the accumulation of proline in dark-treated and water stressed rice leaves [33]. If ammonium accumulation is indeed important in regulating dark- and water stress-induced proline accumulation in detached rice leaves,  $\text{NH}_4\text{Cl}$  or MSO is expected to decrease proline utilization. To test this possibility, detached rice leaves were pretreated with 50 mM ornithine for 3 h to increase the endogenous proline content and then transferred to  $\text{NH}_4\text{Cl}$  or MSO for 8 h. Proline content was then determined. As indicated in Table 1, proline content in  $\text{NH}_4\text{Cl}$ - or MSO-treated rice leaves was greater than in water-treated control rice leaves, suggesting that proline in  $\text{NH}_4\text{Cl}$ - or MSO-treated rice leaves is less utilized than in con-

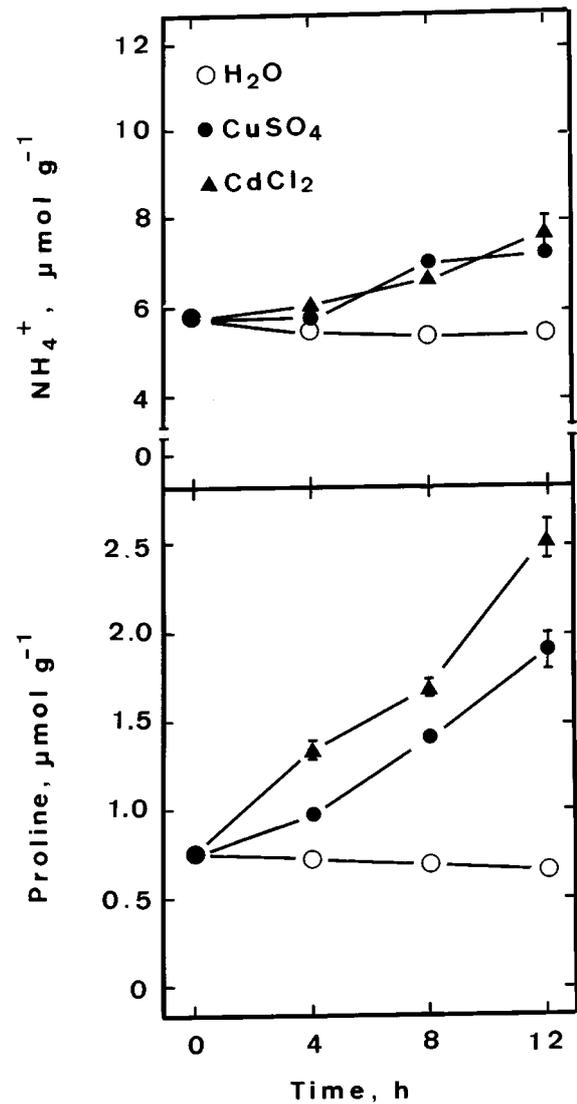


Figure 5. Time courses of proline and ammonium contents in detached rice leaves incubated in distilled water,  $\text{CdCl}_2$  (5 mM) and  $\text{CuSO}_4$  (10 mM). Vertical bars represent standard errors (n = 4).

trols. These results supported further that ammonium accumulation in dark-treated and water stressed rice leaves is associated with proline accumulation.

Proline has been shown to accumulate in plants exposed to excess Cd or Cu [2, 9, 26, 27, 32]. In detached rice leaves, treatment of Cd or Cu also resulted in an accumulation of proline (Figure 5). However, proline accumulation in Cd- and Cu-treated detached rice leaves preceded ammonium accumulation (Figure 5). It is apparent that ammonium accumulation is

not involved in Cd- and Cu-induced proline accumulation in detached rice leaves.

Results from studies conducted in varieties of citrus which were tolerant or susceptible to phosphorus deficiency and cold treatment showed increased ammonium content and increased rate of arginine biosynthesis in leaves exposed to stress [19, 23]. Susceptible plants had lower rates of arginine biosynthesis and higher ammonium content; tolerant plants had greater rates of arginine biosynthesis and lower ammonium content. These data suggest that there is a positive correlation between detoxification of ammonium, arginine biosynthesis and tolerance to several types of environmental stress. Similar findings were reported for studies conducted on alfalfa suspension cultures subjected to osmotic shock as well as cucurbits exposed to salinity stress [19]. In the present investigation, we are able to show that ammonium accumulation in dark-treated and water stressed rice leaves is associated with proline accumulation. Future work should be focused on the correlation between conversion of ammonium into proline and tolerance to dark-induced senescence and water stressed in rice leaves.

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### References

- Aspinall D and Paleg LG (1981) Proline accumulation: physiological aspects. In: Paleg LG and Aspinall D (eds) *Physiology and Biochemistry of Drought Resistance in Plants*, 205 pp. Sydney: Academic Press
- Bassi R and Sharma SS (1993) Changes in proline content accompanying the uptake of zinc and copper by *Lemna minor*. *Ann Bot* 72: 151–154
- Bates LS, Waldren RP and Teare ID (1973) Rapid determination of free proline for water stress studies. *Plant Soil* 39: 205–207
- Becker TW, Hoppe M and Foch PH (1986) Evidence for the participation of dissimilatory processes in maintaining high carbon fluxes through the photosynthetic carbon reduction and oxidation cycle in water-stressed bean leaves. *Photosynthetica* 20: 153–157
- Berger MG and Fock HP (1983) Effect of methionine sulfoximine and glycine on nitrogen metabolism of maize leaves in the light. *Aust J Plant Physiol* 10: 187–194
- Bouchereau A, Aziz A, Larher F and Martin-Tanguy J (1999) Polyamines and environmental challenges: recent development. *Plant Sci* 140: 103–125
- Chen L-M, and Kao CH (1998) Relationship between ammonium accumulation and senescence of detached rice leaves caused by excess copper. *Plant Soil* 200: 169–173
- Chen SJ, Hung KT and Kao CH (1997) Ammonium accumulation is associated with senescence of rice leaves. *Plant Growth Regul* 21: 195–201
- Chen SL and Kao CH (1995) Cd induced changes in proline content and peroxidase activity in roots of rice seedlings. *Plant Growth Regul* 17: 67–71
- Chou IT, Chen CT and Kao CH (1990) Regulation of proline accumulation in detached rice leaves. *Plant Sci* 70: 43–48
- Kao CH (1980) Senescence of rice leaves IV. Influence of benzyladenine on chlorophyll degradation. *Plant Cell Physiol* 21: 1255–1262
- Kao CH (1981) Senescence of rice leaves VI. Comparative study of the metabolic changes of senescing and turgid and water-stressed excised leaves. *Plant Cell Physiol* 22: 683–688
- Kar M and Feierabend J (1984) Changes in the activities of enzymes involved in amino acid metabolism during the senescence of detached wheat leaves. *Physiol Plant* 62: 39–44
- Klein H, Priebe A and Jager HJ (1979) Putrescine and spermidine in peas: effects of nitrogen source and potassium supply. *Physiol Plant* 45: 497–499
- Le Ruduller D and Goas G (1971) Mise en evidence et dosage de guelgues amines dans les plantules de *Soja hispida* Moench. Privees de lenrs cotyledons et cultivees en presence de nitrates, d'uree et de chlorure d'ammonium. *CR Acad Sci (Paris) Ser D* 273: 1108–1111
- Lea DJ and Mifflin BJ (1974) Alternative route for nitrogen assimilation in higher plants. *Nature* 251: 614–616
- Lin CC and Kao CH (1996) Disturbed ammonium assimilation is associated with growth inhibition of roots in rice seedlings caused by NaCl. *Plant Growth Regul* 18: 233–238
- Lin J-N and Kao CH (1998) Water stress, ammonium, and leaf senescence in detached rice leaves. *Plant Growth Regul* 26: 165–169
- Lovatt CJ (1991) Stress alters ammonia and arginine metabolism. In: Flores HE, Arteca RN and Shannon JC (eds) *Polyamines and Ethylene: Biochemistry, Physiology, and Interactions*, pp 166–179. Maryland: American Society of Plant Physiologists
- Peeters KMU and Van Laere AJ (1992) Ammonium and amino acid metabolism in excised leaves of wheat (*Triticum aestivum*) senescing in the dark. *Physiol Plant* 84: 243–249
- Platt SG and Anthon GE (1981) Ammonium accumulation and inhibition of photosynthesis in methionine sulfoximine treated spinach. *Plant Physiol* 67: 509–513
- Postitus C and Jaccobi G (1976) Dark starvation and plant metabolism. VI. Biosynthesis of glutamic acid dehydrogenase in detached leaves of *Cucurbita maxima*. *Z Pflanzenphysiol* 78: 133–140
- Rabe E (1990) Stress physiology: the functional significance of the accumulation of nitrogen containing compounds. *J Hort Sci* 65: 231–243
- Rabe E and Lovatt CJ (1986) Increased arginine biosynthesis during phosphorus deficiency. *Plant Physiol* 81: 774–779
- Ronzio RA, Rowe WB and Meister A (1969) Studied on the mechanism of inhibitor of glutamine synthetase by methionine sulfoximine. *Biochemistry* 8: 1061–1075
- Saradhi A and Saradhi PP (1981) Proline accumulation under heavy metal stress. *J Plant Physiol* 138: 554–558

27. Schat H, Sharma SS and Vooijs R (1997) Heavy metal-induced accumulation of free proline in a metal-tolerant and a nontolerant ecotype of *Silene vulgaris*. *Physiol Plant* 101: 477–482
28. Simpson RJ and Dalling MJ (1981) Nitrogen redistribution during grain growth in wheat (*Triticum aestivum* L.). III. Enzymology and transport of amino acids from senescing flag leaves. *Planta* 151: 447–456
29. Singh TN, Paleg LG and Aspinall D (1973) Stress metabolism I. Nitrogen metabolism and growth in the barley plant during water stress. *Aust J Biol Sci* 26: 45–56
30. Thomas H (1978) Enzymes of nitrogen mobilization in detached leaves of *Lolium temulentum* during senescence. *Planta* 142: 161–169
31. Wang CY, Cheng SH and Kao CH (1982) Senescence of rice leaves VII. Proline accumulation in senescing excised leaves. *Plant Physiol* 69: 1348–1349
32. Wu J-T, Chang SJ and Chou T-L (1995) Intracellular proline accumulation in some algae exposed to copper and cadmium. *Bot Bull Acad Sin* 36: 69–93
33. Yang C-W, Lin CC and Kao CH (1999) Endogenous ornithine and arginine contents and dark-induced proline accumulation in detached rice leaves. *J Plant Physiol* (in press)