行政院國家科學委員會補助專題研究計畫成果報告

題目:尋找位於人類第十六號染色體上可能與肝癌發生有關抑癌基因的研究

計畫類別: 個別型計畫 整合型計畫 計畫編號:NSC89 - 2315 - B - 002 - 036 -

執行期間:八十九年八月一日至九十年七月三十一日

計畫主持人:許金川 共同主持人:

本成果報告包括以下應繳交之附件:

赴國外出差或研習心得報告一份 赴大陸地區出差或研習心得報告一份 出席國際學術會議心得報告及發表之論文各一份 國際合作研究計畫國外研究報告書一份

執行單位:台大醫學院內科

中華民國 90 年 10 月 17 日

關鍵詞:肝細胞癌,微小衛星,雜合子丟失

肝細胞癌是全世界最好發的癌症之一,在台灣也是國人癌症死因的第一位,目前真正 的致病機轉仍不清楚。肝癌的發生通常伴隨基因的變化,其中包括致癌基因的活化、或癌 抑制基因的不活化。目前已知有多種人類腫瘤均與癌抑制基因之變異有關。近年來利用染 色體中的微小衛星可以有系統而快速的分析比較每一染色體中基因的變化,更可進一步尋 找可能的癌抑制基因。

我們的研究結果顯示肝癌病患在 16q 常有雜合子丟失(LOH)的情形,且細微定位的結 果靠近 16q12.1,16q21-22 和 16q24.3。因此表示這些位置可能有癌抑制基因的存在。目前, 我們利用位於 16q12.1 的標記(D16S415, D16S419, D16S409, D16S3080 及 D16S3034) 開始 篩選人類之細菌人工染色體基因庫(Bacterial Artificial Chromosome Library, BAC Library),並配合表現序列捕捉系統(exon trapping system)找到表現序列的 clones。總共選

殖到 15 個可能的表現序列,與 NCBI (national center for Biotechnology Information)的資料 庫比對的結果,有二個序列與最近被發現的 KIAA1005 基因序列完全相同。而 KIAA 1005 基因在肝癌組織中有同源性丟失的現象,反轉錄-聚合 連鎖反應(RT-PCR)的結果也顯示 mRNA 的表現有差異,因此我們推測 KIAA1005 基因可能是位於 16q12.1 附近的假想抑癌 基因.

KIAA1005 基因目前功能不明,只能就所推測的氨基酸順序所含有的特殊結構來分析它可能有何種功能,由於 KIAA1005 含有 Leucine zipper 及 C2 domain,與 yeast hypothetical 蛋白及人類 desmoplakin 有 22%-30%的相同性,所以它可能是 transcription factor 或是與 desmosomal plaque 有關。KIAA1005 基因的 mRNA 表現也非常特殊,似乎 有三種型式,但究竟這兩種型式與原先 KIAA 1005 的功能是否相同,需再進一步研究。而 KIAA1005 基因是否真的在肝細胞癌的發生過程扮演重要角色則需要進行更多研究才能有正確的答案。

Abstract

Key words: hepatocellular carcinoma, micorsatellite analysis, tumor suppressor gene

Hepatocellular carcinoma (HCC) is one of the most common cancer in the world and is the leading cause of cancer death in Taiwan. The prognosis of this cancer is extremely poor with survival of only several months after symptoms occurred. Elucidation of the basic genetic changes of HCC is important for the understanding and treatment of this cancer. Cancer is usually accompanied with genetic alternations either through the activation of cellular oncogene or the inactivation of cancer suppressor gene. The recently identified short tandem repeat, the microsatellite, which is widely distributed throughout the whole human genome. Identification of disease genes as well as tumor suppressor genes by microsatellite polymorphism. have been published recently. In this study we use microsatellite marker to analyze 88 cases of HCCs, most of them are small in size, in order to study the genetic changes of HCC and to further narrow down the common LOH sites in 16q.

In this study, we have used 35 microsatellite markers for further fine mapping of LOH. We have confirmed the most frequent regions of LOH for HCC are 16q12.1, 16q22, and 16q24. After analyzing these information, we started to screen the human BAC(Bacterial Artificial chromosome)library by these markers at 16q12.1 and we identified 15 clones. Exon trapping system is used to search the putative exon sequences of BAC genomic clones. Two exon-like sequences are identical to the KIAA1005 gene. Homozygous deletion of KIAA1005 was found in 37%(10 /27) HCCs. These data suggested that the KIAA1005 might be the putative tumor suppressor genes at chromosome 16q12.1.

The function of KIAA is unknown. From the predicted sequences, we can predict that

the gene might be a transcription factor or be correlated with phosphorylation. Furthermore, it seems have different splicing form and play different role in HCC development. However, this need further investigation to elucidate KIAA function.

Introduction

Hepatocellular carcinoma (HCC) is one of the most common cancer in the world [1] and is the leading cause of cancer death in Taiwan [2-11]. Chronic hepatitis B and recently the hepatitis C viral infection is thought related to the development of HCC [2-13]. However, the basic molecular mechanism remained to be clarified. Cancer is usually accompanied with genetic alternations either through the activation of cellular oncogene or the inactivation of cancer suppressor gene [14-25]. Traditionally, cytogenetics was used to detect genetic change of cancer[26-32]. However, only major genetic changes resulting in chromosomal translocation, deletion or other changes can be identified by this method. Another method to detect the cancer suppressor gene is the use of RFLP (restriction fragment length polymorphism). However, this method is tedious, laborious and need a lot of DNA samples. Microsatellite, which is short tandem sequences is widely distributed throughout the whole human genome. The target DNA sequences can be amplified by polymerase chain reaction and the fragment was run on gel. Only small sample of starting DNA sample was needed. These markers were rather complete evenly distributed in the whole chromosome. Using the microsatellite as markers, we can study the loss of heterozygosity(LOH) of tumor[33-92]. Identification of disease gene including tumor suppressor gene by the linkage analysis of comparison of tumor and non-tumor part had been published recently[58,59].

In the past years, we have using 232 microsatellite markers to study the LOH of HCC. We included 30 cases of HCC and we have found the most common site of LOH for HCC is: 16q(43.3%), 13q(36.7%), 17p(30%), 5q(20%), 11p(20%), 11q(16.7%). These data provides as a basis for further fine mapping of the common site of LOH and to identify the putative tumor suppressor gene.

In this proposal, we will further study the LOH of HCC by narrowing down the common site of LOH and to identify the putative tumor suppresser gene. Since the 16q has the highest rate of LOH, we will start from this chromosome in the first year. Our preliminary study indicated that the most minimal commonly affected region of 16q was deduced to be between 16q12.1 and 16q12.2. Another small region was determined to be 16q24.1. Therefore, in this project, we used more micorsatellite markers in these 3 regions to narrow down the common site of LOH and to identify the putative tumor suppressor gene.

Materials and Methods

Methods

LOH analysis

Genomic DNA was extracted from tumor and non-tumor part with classical methods, then these samples subjected to study by using the microsatellite markers in the aforementioned 2 region. All the markers available from the literature will be used. Each reaction was amplified in a total volume 25 ul containing 25 ng of genomic DNA; 75 uM concentrations each of dGTP, dCTP, and dTTP; 2.5 uM dATP; 2 uCi[a-35S]dATP; 1x PCR buffer (10 mM Tris-Cl pH 8.3, 1.5 mM MgCl2, 0.01% gelatin); 0.2 uM for each primer; and 0.5 u of Taq polymerase. The PCR was performed in 94°C for 30 sec, 55°C for 75 sec, 72°C for 15 sec. A total of 27 cycles were performed. The product was further incubated in 72°C for another 6 min. After completion of PCR, 6 microliter products were subjected to electrophoresis in a 6% PAGE gel followed by

exposure of the gel to X ray film for 1 to 2 days.

The band pattern between tumor and non-tumor part was compared. The LOH was analyzed and the common site of LOH was deduced.

BAC library screening

After analyzing data, we started to screen the human BAC(Bacterial Artificial chromosome) by these markers at 16q12.1 and we identified 15clones.

Exon trapping system

We applied exon amplification strategy to isolate genes from complex BAC genomic DNA.

Results

LOH and fine mapping analysis of 16q in HCC

In this study we use microsatellite marker to analyze 88 cases of HCCs, most of them are small in size, in order to study the genetic changes of HCC and to further narrow down the common LOH sites in 16q.

A total of 36 microsatellite markers in chromosome 16q were used to study the LOH of HCC. The average number of microsatellite markers included in 16q was about 3-5 cM. Range of LOH for each primer was from 2.5 to 40.5% and the average was 17.5%. The highest percentage of LOH was shown for loci at D16S496(40.5%), D16S504(33.9%), D16S3034(30.2%), D16S3039(30%), D16S517(27%), D16S409(27%), D16S514(26%), D16S419(25%), D16S3080(24.7%), D16S3018(24%), D16S3137(23.4%), D16S415(22%), D16S390(22%), D16S515(21%), D16S3048(20.5%), D16S408(20%), D16S266(18%), and D16S422(18%) (Figure 1). The frequency of LOH seemed to have relationship with HBV carrier. The HBsAg positive group seemed to have higher LOH frequency at 16q12.1, and the difference was statistatically significant.

In the past year, we started to screen the human BAC(Bacterial Artificial chromosome)library by these markers at 16q12.1 and we identified 15 clones(Table 1). Exon trapping system is used to search the putative exon sequences of BAC genomic clones. Two exon-like sequences are identical to the KIAA1005 gene. Homozygous deletion of KIAA1005 was found in 37%(10/27) HCCs (figure 2). These data suggested that the KIAA1005 might be the putative tumor suppressor genes at chromosome 16q12.1.

Discussion

Our preliminary result shows that HCCs in Taiwan have frequent LOH in chromosome 16q and the common LOH regions were narrowed down to D16S409 (16q12.1), D16S419-D16S408 (16q12.1), D16S514-D16S496 (16q21-22), and D16S422-D16S402 (16q24.3). Furthermore, familial HCC seems to have common LOH sites at 16q12.1 and 16q21-22. These data suggested that the putative tumor suppressor genes might lie in these regions and we identified 15 BAC clones near these regions.

In addition to construct the physical map of 16q, we also intend to search the possible exon sequence near these regions. Exon trapping system is a good approach to search the possible exon sequences. Now we have applied the system to our project. Furthermore, we try to clone full-length cDNA using RACE system and search for the putative tumor suppressor genes.

Figure 2. Summary of the results of LOH analysis on 16q in 88 HCCs.

marker -D16S517(27.3%, q11.2) -D16S261(6.5%, q12.1) -D16S409(27.2%, q12.1), D16S3044(2.5%, q12.1), D16S3080(24.7%, q12.1) -D16S416(9.5%, q12.1) -D16S3136(7%,q12.1), D16S3120(11.5%,q12.1) -D16S419(25.4%, q12.1), D16S415(22.7%, q12.1), D16S390(22.4%, q12.1) -D16S3034(30.2%, q12.1-13), D16S3137(23.4%, q12.1-13), D16S3053(14.7%, q12.1-13) -D16S3112(17.5%, q13), D16S3039(30%, q13) -D16S408(20%, q21) -D16S494(15.7%, q21) -D16S514(26.2%, q21) -D16S496(40.5%, q22.1), D16S398(10.3%, q22.1) -D16S397(7.1%, q22.1) -D16S512(13.8%, q22.2) -D16S3018(24%, q22.2) -D16S515(20.8%, q22.3) -D16S266(18.6%, q23.1) -D16S516(13.8%, q24.1) -D16S504(33.95, q24.1) -D16S507(15%, q24.1) -D16S505(7%, q24.2) -D16S422(18.4%, q24.3) -D16S402(16.1%, q24.3) -D16s3037(11.1%, q24.3) -D16S520(8.9%, q24.3) -D16S3048(20.5%, q24.3) -D16S413(7%, q24.3)



Table 1. Summary of the results of BAC clones by screening human BAC library

Microsatellite loci used for	BAC clone
screening	
D16S409	106K13
D16S419	107F06
D16S415	29M12
	60F14
	72M15
D16S390	211D06
D16S3034	71E17
	35D12
	216N13
	250G9
D16S3044	25L24
	26L24
	39004
D16S3080	46B1
	58M17