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SESQUITERPENES FROM LEAVES OF CRYPTOMERIA JAPONICA

WEN-CHIUNG SU, JIM-MIN FANG and YU-SHIA CHENG*

Department of Chemistry, National Taiwan University, Taipei, Taiwan 106, Republic of China

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Abstract—Twenty-seven sesquiterpenes were isolated from leaves of *Cryptomeria japonica*. The new compounds included elem-1-en-4,11-diol, 11-acetoxyeudesman-4 α -ol, eudesmane-5 α ,11-diol, 3-eudesmene-1 β ,11-diol, 1 β -acetoxy-3-eudesmen-11-ol, 4-eudesmene-1 β ,11-diol, 1 β -acetoxy-4-eudesmen-11-ol, 7-epi- γ -eudesmol, 7-epi-4-eudesmene-1 β ,11-diol, 1 β -acetoxy-4(15)-eudesmen-11-ol. Their structures were determined by chemical and spectral methods.

INTRODUCTION

The Japanese cedar, Cryptomeria japonica D. Don., is a widely distributed conifer called 'sugi' in Japanese. We recently reported the isolation and structural determination of chamaecydin triterpene [1], abietane, kaurane and labdane diterpenes [2, 3] from the ethyl acetate-soluble part of the leaves of C. japonica. As a continuation of this study, we describe herein, 27 constituents of sesquiterpenes including 10 novel compounds 5, 8, 9, 11, 12, 14–17, and 23.

RESULTS AND DISCUSSION

The leaves of *C. japonica* were extracted with acetone. The ethyl acetate-soluble portion of the extract was subjected to chromatography to give sesquiterpenes **1–27**. The known compounds epitodomatuic acid (1) [4], epijuvabione (2) [4], 11-hydroxy-4,5-secoeudesmane-4,5-dione (3) [5], elemol (4) [6], cryptomeridiol (6) [7], 4-epicryptomeridiol (7) [8], α -eudesmol (10) [9, 10], γ -eudesmol (13) [9], 6-eudesmene-1 β ,4 β -diol (18) [11], oplodiol (19) [10, 12], β -eudesmol (20) [13], 4(15)-eudesmene-1 β ,11-diol (21) [14], 4(15)-eudesmene-1 β ,6 α -diol (22) [15, 16], α -cadinol (24) [17], T-cadinol (25) [18], oplopanone (26) [19] and cedrol (27) [20] were identified by comparison of their physical and spectral data (mp, [α], mass, IR, ¹H and ¹³C NMR) with literature.

The molecular formula, $C_{15}H_{28}O_2$, of 5 was inferred from its exact mass 240.209. The ¹³C NMR spectrum showed signals for a terminal double bond at $\delta 153.3$ (d) and 109.5 (t). The signals for two isopropanol moieties appeared at $\delta 1.17$ (Me), 1.18 (3 Me), 72.7 (s) and 75.4 (s). The proton resonance at $\delta 1.57$ was assigned to H-5 by means of C-H COSY and HMBC. As H-5 appeared as a double of doublet (J = 12, 3 Hz), it was in the axial orientation. Compound 5 had chemical shifts for C-7 and C-10 at δ 49.5 (d) and 40.3 (s), close to the values for those signals in elemol. The structure of 5 was determined to be elem-1-en-4,11-diol and its (5R,7R,10S)-configuration was tentatively assigned by analogy to that of elemol. Compound 5 was unstable in CDCl₃ solution. A product 5a, (1S)-elemane-1,4,11-triol, was obtained presumably by the acid-catalysed hydration of 5. The S-configuration was determined by Horeau's method [21]. Compound 5 can be also regarded as a hydration derivative of elemol (4).

From spectral analyses, **8** ($C_{17}H_{30}O_3$) was readily determined to be 11-acetoxyeudesman-4 α -ol. It showed the IR absorption at 1723 cm⁻¹ and carbon resonances at δ 22.4 and 170.5 for the acetoxy group. The structure of **8** was confirmed as it was saponified to yield cryptomeridiol (6). Two C-11 methyl groups in 6 appearing at δ 1.16 were deshielded to δ 1.40 and 1.43 in **8** as the C-11 hydroxyl group was converted to the acetoxy group.

An eudesmanediol (9) $(C_{15}H_{28}O_2)$ exhibited the parent peak in the mass spectrum at m/z 240.209. The C-H COSY and HMBC experiments led to the assignment of 9 as eudesmane-5 α ,11-diol. Irradiation of Me-10 (at δ 0.93) caused 7% NOE of Me-4 (at δ 0.81) and 10% NOE of H- 6β (at δ 1.28). The signal of H-6 α appeared at a relatively low field δ 1.72 presumably due to the deshielding effect of the 5 α -hydroxyl group. The large coupling constant 12.5 Hz between H-6 β and H-7 was in agreement with their axial positions.

Based on the spectral analyses, two isomers $(C_{15}H_{26}O_2)$ 11 and 14, were assigned as 3-eudesmene-1 β ,11-diol and 4-eudesmene-1 β ,11-diol, respectively. Two olefinic carbons in 11 occurred at δ 119.5 (d) and 135.4 (s), whereas those in 14 appeared at δ 123.8 (s) and 133.6 (s). The H-1 in 11 was axially oriented to exhibit its resonance as a double of doublet (at δ 3.50) with 10 and 6.5 Hz coupling constants. The resonance of H-1 in 14 also showed a similar pattern (dd, J = 9, 7 Hz). An allylic

^{*}Author to whom correspondence should be addressed.



proton H-5 appearing at $\delta 1.66$ (*dd*, J = 9, 3 Hz) also conformed to the *trans*-fused configuration of 11. The C-7 resonances of 11 and 14 occurred at $\delta 49.2$ and 49.7 close to the value of C-7 ($\delta 50.0$) in α -eudesmol (10).

Compounds 12 and 15 are 1β -acetoxy-3-eudesmen-11ol and 1β -acetoxy-4-eudesmen-11-ol, the acetates of 11 and 14, respectively. Due to the inductive effect of acetoxy groups, the H-1 resonances in 12 and 15 occurred at low fields $\delta 4.74 (dd, J = 9.5, 6.5 \text{ Hz})$ and 4.67 (dd, J = 8, 8 Hz). Saponification of 12 and 15 gave, respectively, the corresponding diol 11 and 14.

Compounds 16 ($C_{15}H_{26}O$) and 17 ($C_{15}H_{26}O_2$) were determined to be 7-epi- γ -eudesmol and 7-epi-4-eudesmene-1 β ,11-diol, respectively. The coupling constants between the C-7 and C-6 protons were small (2-3.5 Hz) as the C-7 protons were on equatorial positions. The C-7 signals in 16 and 17 appeared at δ 44.1, whereas those signals in their 7-epimers 13 and 14

			Table 1. ¹ HNMR	spectral data of new con	mpounds (CDCl ₃ solution	on, ô values in ppm, J val	lues in Hz)*+		
H	s.	6	11	12	14	15	16	17	23
-	$6.04 \ (dd, J = 10.5, 17.5)$	1.12 (m) 1 95 (m)	$3.50 \ (dd, J = 6.5, 10)$	$4.74 \ (dd, J = 6.5, 9.5)$	3.41 (dd, J = 7,9)	$4.67 \ (dd, J = 8,8)$		$3.50 \ (dd, J = 8,8)$	$4.64 \ (dd, J = 4.5, 11.5)$
7	4.92 (dd, J = 1,10.5) 5.00 (dd, J = 1,17.5)	1.35 (m) 1.60 (m)	1.86 (dd, J = 10,13.5)	$2.00 \ (dd, J = 9.5, 13.5)$	1.65 (m)				
ŝ	1.17 (s)	1.35 (m) 1.36 (m)	5.25 (brs)	5.24 (br s)	1.95 (ddd, J = 3,3,12.5)	$1.96 \ (ddd, J = 2,3,12)$			2.13 (ddd, J = 5,12,13.5)
Ś	1.57 (dd, J = 3.12)	(m) (m)	1.66 (dd. J = 3.9)	$1.85 \ (brd. J = 12)$		(71'6'7 = r 'boo) (77'7 (c			(2.21, 2, -2, -2, -2, -2, -2, -2, -2, -2, -2,
9	•	$1.28 \ (dd, J = 12.5, 12.5)$			$1.64 \ (ddd, J = 1.5,9,13.)$	5) 1.70 (dd , $J = 9,13.5$)	2.10 (dd, J = 2,15)	$2.01 \ (dd, J = 3.5, 15)$	
12	1.18 (s)	1.15 (au, $J = 2.3,12.1$) 1.15 (s)	1.18 (s)	1.18 (s)	.c1,c,c.z = r .aaa, J = 2.30	(C.C.1,C.7 = L , aaa, J = L.00 (c.C.1,C.7 = L , 1.18 (s)	(c1,2 = l, aa, bab, 20,2	$(c_1, c_2 = c, a_2) = (c_2, c_3)$	1,17 (s)
13	1.18 (s)	1.15 (s)	1.19 (s)	1.19 (s)	1.17 (s)	1.18 (s)	1.23 (s)	1.21 (s)	1.17 (s)
14	1.15 (s)	0.93 (s)	0.72 (s)	0.81 (s)	(s) (s)	1.05 (s)	1.06 (s)	1.02 (s)	0.72 (s)
15	1.18 (s)	0.81 (d, J = 7)	1.59 (s)	1.59 (s)	1.55 (s)	1.57 (s)	1.66 (s)	1.63 (s)	4.51 (d, J = 1)
									4.75(d, J = 1)
OAc				2.02 (s)		2.03 (s)			2.01 (s)
¦\$₽Ę	me assignable resonances to rest appeared at $\delta 1.0-2$.	for 8 appeared at $\delta 0.84$ (0 overlapping with other	(s, H-14), 1.08 (s, H-15), 1. r signals.	.40 (s, H-12), 1.43 (s, H-1	3), 1.94 (s, OAc) in addit	ion to others.			

appeared at lower fields ($\Delta \delta = 5$ ppm) [22]. In contrast, the C-11 signals of the isopropanol moieties in 16 and 17 occurred at lower fields than those in 13 and 14.

Compound 23 showed IR absorption at 3453 (broad) and 1711 cm⁻¹ attributable to hydroxyl and acetyl groups. The exact mass at m/z 280.202 indicated the molecular formula C17H28O3 and an intense signal at m/z 202 was attributable to the fragment derived by elimination of water and acetic acid $[M - H_2O]$ -HOAc]⁺.By analysis of the ¹H and ¹³C NMR spectra, the structure of 23 was determined to be 1β -acetoxy-4-(15)-eudesmen-11-ol. To confirm this structural assignment, 23 was saponified to give the eudesmenediol 21.

In summary, a series of known and new sesquiterpenes were isolated from the leaves of C. japonica. The structures of new sesquiterpenes were determined by analyses of their spectra. The acetates 8, 12, 15 and 23 were correlated with their corresponding alcohols by saponification.

EXPERIMENTAL

General. Merck silica gel 60F sheets were used for analyt. TLC. HPLC was carried out on a Hibar Lichrosorb Si 60 (7 μ m or 10 μ m) column (25 cm × 1 cm).

Plant material. The plant used in this study is introduced from Japan and cultivated in suburban Taipei. A voucher specimen has been deposited in our laboratory. The leaves (1.4 kg) of C. japonica D. Don. were exhaustively extracted with Me₂CO. The Me₂CO extract was passed through a pad of charcoal, concd and re-extracted with EtOAc. The EtOAc-soluble portion (45 g) was chromatographed on a silica gel column by elution with gradient of hexane and EtOAc. The appropriate frs were combined and purified by HPLC to give 16 (20 mg), 2 (16 mg), 24 (12 mg), 25 (8 mg), 27 (22 mg), 23 (27 mg), 4 (12 mg), 10 (8 mg), 20 (13 mg), 8 (27 mg), 15 (10 mg), 12 (11 mg), 13 (15 mg), 26 (20 mg), 9 (21 mg), 1 (15 mg), 19 (8 mg), 17 (3 mg), 22 (12 mg), 11 (14 mg), 14 (50 mg), 3 (3 mg), 21 (15 mg), 18 (5 mg), 7 (35 mg), 5 (5 mg) and 6 (25 mg), in order of increasing polarity.

Epitodomatuic acid (1). Oil, $[\alpha]_{D}^{20} + 71^{\circ}$ (CHCl₃; c 1.5), lit. [4], Oil, $[\alpha]_D^{25} + 71.2^{\circ}$ (CHCl₃; c 1.07). Epijuvabione (2). Oil, $[\alpha]_D^{25} + 60^{\circ}$ (CHCl₃; c 1.6), lit.

[4], Oil, $[\alpha]_D^{25} + 60^\circ$ (CHCl₃; c 1.18).

11-Hydroxy-4,5-secoeudesmane-4,5-dione (3). Oil, $[\alpha]_{D}^{15}$ + 50° (CHCl₃; c 0.3), lit. [5], Oil, $[\alpha]_D^{24}$ + 46° (CHCl₃; c 0.39). ¹³C NMR (CDCl₃, 75 MHz); δ18.3 (C-2), 21.9 (C-8), 23.1 (C-14), 27.2 (C-12), 27.4 (C-13), 29.9 (C-15), 36.3 (C-9), 37.3 (C-1), 39.8 (C-6), 44.3 (C-3), 47.1 (C-10), 49.6 (C-7), 72.0 (C-11), 209.1 (C-4), 215.8 (C-5).

Elemol (4). Oil, $[\alpha]_D^{25} - 6^\circ$ (CHCl₃; c 1.8), lit. [6], mp 52–53°; $[\alpha]_D - 5.82^\circ$ (CHCl₃; c 3.4). ¹³C NMR (CDCl₃, 75 MHz): δ16.6 (C-14), 22.5 (C-8), 24.7 (C-15), 27.1 (C-12, 13), 28.5 (C-6), 39.7 (C-10), 39.9 (C-9), 49.3 (C-7), 52.7 (C-5), 72.7 (C-11), 109.9 (C-2), 112.0 (C-3), 147.9 (C-4), 150.2 (C-1).

Elem-1-en-4,11-*diol* (5). Oil, $[\alpha]_{\mathbf{D}}^{15} - 14^{\circ}$ (CHCl₃; c 0.5). TLC (50% EtOAc in hexane) R_{f} 0.44. IRv_{max}^{neat} cm⁻¹: 3388, 3081, 1625, 914. EIMS (70 eV) m/z (rel. int.): 240 [M]⁺

С	5	8	9	11	12	14	15	16	17	23
1	153.3	41.0	33.0	76.3	78.2	78.3	80.3	39.4	75.4	80.7
2	109.5	20.2	22.2	32.3	29.1	27.1	23.9	18.9	26.9	28.0
3	31.5	43.6	30.5	119.5	119.0	31.9	31.5	32.7	31.3	33.8
4	75.4	72.2	32.4	135.4	135.3	123.8	124.0	126.0	125.1	148.2
5	55.7	54.8	75.2	46.5	46.4	133.6	133.4	135.0	133.6	48.7
6	23.0	21.2	32.1	23.9	23.7	26.5	26.3	25.4	25.2	24.2
7	49.5	47.2	45.2	49.2	49.1	49.7	49.7	44.1	44.1	47.6
8	25.5	22.1	21.3	21.9	21.8	22.8	22.7	22.6	21.8	21.9
9	43.8	44.4	36.3	35.0	34.9	38.8	38.6	38.1	33.5	36.6
10	40.3	34.6	37.6	37.4	36.3	39.4	38.4	34.4	39.5	39.1
11	72.7	85.1	72.5	72.9	72.8	72.7	72.6	74.6	74.1	72.7
12	27.5	23.5	26.9	26.7	26.7	26.7	26.7	27.8	27.4	27.0
13	28.2	23.7	27.0	27.6	27.6	27.1	27.2	29.8	29.4	27.2
14	17.1	18.6	22.7	9.5	10.6	17.3	18.4	25.9	19.2	11.2
15	26.6	22.5	14.9	20.9	21.2	18.9	18.9	19.6	19.3	107.2
OAc		170.5			171.0		171.0			170.9
		22.4			20.7		21.3			21.2

Table 2. ¹³CNMR spectral data of new compounds (75 MHz, CDCl₃ solution, δ values in ppm)

(10), 239 (95), 221 (7), 154 (20), 134 (55), 98 (65), 43 (100). HRMS for $C_{15}H_{28}O_2$ requires 240.2091; found 240.2085.

(1S)-Elemane-1,4,11-triol (5a). This was obtained when 5 stood in CDCl₃ soln for 2 weeks. White crystals from CHCl₃-hexane (1:1), mp 93-94°, $[\alpha]_{D}^{30} - 3.3^{\circ}$ (CHCl₃; c 0.3). TLC (30% EtOAc in hexane) R_f 0.47. IR v_{max}^{KBr} cm⁻¹: 3435. ¹H NMR (CDCl₃, 300 MHz): $\delta 0.82$ (s, H-14), 1.05 (s, H-15), 1.07 (d, J = 6.5 Hz, H-2), 1.18 (s, H-3), 1.20 (s, H-12), 1.21 (s, H-13), 1.50 (dd, J = 3, 12 Hz, H-5), 3.45 (q, J = 6.5 Hz, H-1). ¹³C NMR (CDCl₃, 75 MHz): δ13.5 (C-2), 14.0 (C-14), 21.8 (C-6), 21.9 (C-8), 24.0 (C-15), 27.1 (C-12), 27.8 (C-13), 30.4 (C-3), 36.0 (C-9), 43.5 (C-10), 49.5 (C-7), 58.1 (C-5), 72.9 (C-11), 78.8 (C-4), 81.6 (C-1). EIMS (70 eV) m/z (rel. int.): 225 $[M - H_2O - Me]^+$ (42), 207 (3), 178 (70), 163 (50), 135 (100), 121 (18), 95 (15). HRMS for $[C_{15}H_{30}O_3 - H_2O - Me]$ requires 225.1856; found 225.1855. A sample of 5a was treated with (\pm) -2-phenylbutanoic anhydride in pyridine at 25° for 1 hr. The recovered 2-phenylbutanoic acid after work up showed levorotation, 5a was thus assigned to have (1S)-configuration [21].

Cryptomeridiol (6). Mp 136–137°. $[\alpha]_D^{25} - 33°$ (CHCl₃; c 1.5), lit. [7], mp 137.5°; $[\alpha]_D^{20} - 21.7°$ (CHCl₃; c 2.5).

4-Epicryptomeridiol (7). Mp $81-82^{\circ}$. $[\alpha]_D^{25} + 4^{\circ}$ (CHCl₃; c 2.5), lit. [8], mp $81-82^{\circ}$; $[\alpha]_D + 3.8^{\circ}$ (CHCl₃; c 0.22).

11-Acetoxyeudesman-4α-ol (8). Oil, $[\alpha]_{D}^{25} - 13^{\circ}$ (CHCl₃; c 2.7). TLC (9% EtOAc in CH₂Cl₂) R_f 0.33. IR ν_{max}^{neat} cm⁻¹: 3423, 1723. EIMS (70 eV) m/z (rel. int.): 282 [M]⁺ (2), 281 (M - H)⁺ (7), 222 [M - MeCOOH]⁺ (35), 204 (52), 189 (21), 161 (20), 149 (35), 109 (25), 81 (30), 43 (100). HRMS for C₁₇H₃₀O₃ requires 282.2196; found 282.2175.

Saponification of 8. A soln of 8 (20 mg) in EtOH (5 ml) was treated with 10% KOH in EtOH (2 ml) at 25° for 16 hr. The mixt. was extracted with Et_2O and sepd by

HPLC (30% EtOAc in hexane) to give 8 (10 mg) and 6 (8 mg).

Eudesmane- 5α -11-*diol* (9). Solid, mp 66–67°. $[\alpha]_D^{25}$ + 38° (CHCl₃; c 2.1). TLC (15% EtOAc in CHCl₃) R_f 0.32. IR ν_{max}^{KBr} cm⁻¹: 3465. EIMS (70 eV) *m/z* (rel. int.): 240 [M]⁺ (13), 222 (12), 207 (22), 181 (20), 164 (18), 149 (42), 126 (100), 112 (82). HRMS for C₁₅H₂₈O₂ requires 240.2090; found 240.2092.

 α -Eudesmol (10). Oil, $[\alpha]_D^{25} + 28^\circ$ (CHCl₃; c 0.8), lit. [9], mp 75°; $[\alpha]_D + 28.6^\circ$ (CHCl₃; c 1.86).

3-Eudesmene-1 β -11-diol (11). Needles from CHCl₃hexane (7:3), mp 144–145°. $[\alpha]_D^{25} - 4^\circ$ (CHCl₃; c1.4), TLC (33% EtOAc in CH₂Cl₂) R_f 0.52. IR ν_{max}^{KBr} cm⁻¹: 3333. EIMS (70 eV) m/z (rek int.): 238 [M]⁺ (7), 220 (15), 202 (5), 177 (15), 121 (25), 93 (38), 59 (100). HRMS for C₁₅H₂₆O₂ requires 238.1934; found 238.1939.

1β-Acetoxy-3-eudesmen-11-ol (12). Oil, $[\alpha]_D^{28} + 15.5^{\circ}$ (CHCl₃; c 1.1). TLC (20% EtOAc in hexane) R_f 0.4. IR ν_{max}^{neat} cm⁻¹: 3443, 1730. EIMS (70 eV) m/z (rel. int.): 280 [M]⁺ (1), 262 (3), 235 (13), 220 (20), 203 (85), 187 (30), 159 (35), 145 (50), 43 (100). HRMS for C₁₇H₂₈O₃ requires 280.2039; found 280.2058. Saponification of 12 (10 mg) by a procedure similar to that for 8 gave 11 (8 mg).

 γ -Eudesmol (13). Mp 72–73°. $[\alpha]_D^{25} + 21°$ (CHCl₃; c 1.5), lit. [9], mp 73–74°; $[\alpha]_D + 18.7°$ (CHCl₃; c 0.7).

4-Eudesmene-1β,11-diol (14). Crystals from CH₂Cl₂-hexane (6:4), mp 137-138°. $[\alpha]_D^{15}$ + 61° (CHCl₃; c 5.0). TLC (33% EtOAc in CH₂Cl₂) R_f 0.38. IR ν_{max}^{KBr} cm⁻¹: 3436. EIMS (70 eV) m/z (rel. int.): 238 [M]⁺ (15), 220 (100), 203 (40), 187 (35), 159 (48), 133 (35), 119 (25). HRMS for C₁₅H₂₆O₂ requires 238.1934; found 238.1932.

 1β -Acetoxy-4-eudesmen-11-ol (15). Oil, $[\alpha]_D^{28} + 60^{\circ}$ (CHCl₃; c 1.0). TLC (20% EtOAc in hexane) R_f 0.44. IR $\nu_{\text{max}}^{\text{neat}}$ cm⁻¹: 3451, 1731. EIMS (70 eV) m/z (rel. int.): 280 [M]⁺ (8), 262 (30), 220 (12), 202 (75), 187 (70), 159 (100), 145 (40), 131 (70). HRMS for $C_{17}H_{28}O_3$ requires 280.2039; found 280.2065. Saponification of 15 (10 mg) by a procedure similar to that for 8 gave 14 (8 mg).

7-Epi- γ -eudesmol (16). Oil, $[\alpha]_D^{25} - 45^\circ$ (CHCl₃; c 2.0). TLC (50% CHCl₃ in hexane) R_f 0.3. IR ν_{max}^{neat} cm⁻¹: 3419. EIMS (70 eV) m/z (rel. int.): 222 [M]⁺ (22), 204 (95), 189 (80), 161 (100), 149 (25), 133 (35), 119 (18). HRMS for C₁₅H₂₆O requires 222.1985; found 222.1988.

7-Epi-4-eudesmene-1 β ,11-diol (17). Oil, $[\alpha]_D^{25} - 15^\circ$ (CHCl₃; c 0.2). TLC (30% EtOAc in hexane) R_f 0.25. IR ν_{max}^{neat} cm⁻¹: 3395, 1647. EIMS (70 eV) m/z (rel. int.): 238 $[M]^+$ (4), 220 (15), 202 (10), 187 (11), 159 (17), 105 (20), 59 (60), 43 (100). HRMS for C₁₅H₂₆O₂ requires 238.1934; found 238.1938.

6-Eudesmene-1β,4β-diol (18). Mp $137-139^{\circ}$. $[\alpha]_D^{25} - 25^{\circ}$ (CHCl₃; c 0.5), lit. [11], oil; $[\alpha]_{4.36nm}^{24} - 35.4^{\circ}$ (CHCl₃; c 1.0). ¹³C NMR (CDCl₃, 75 MHz): δ11.6 (C-14), 21.4 (C-12), 21.7 (C-13), 22.7 (C-8), 27.0 (C-2), 29.4 (C-15), 35.1 (C-9), 35.3 (C-11), 38.1 (C-10), 38.7 (C-3), 49.8 (C-5), 71.1 (C-4), 78.4 (C-1), 115.4 (C-6), 136.0 (C-7).

Oplodiol (19). Mp 106–107°, $[\alpha]_D^{25} - 52^\circ$ (CHCl₃; c 0.8), lit. [10], mp 107–108°; $[\alpha]_D^{27} - 58.0^\circ$ (CHCl₃; c 1.0).

β-Eudesmol (20). Mp 79–80°, $[\alpha]_D^{25}$ + 56° (CHCl₃; c 1.3), lit. [13], mp 79–80°; $[\alpha]_D$ + 56.6° (CHCl₃; c 2.0). 4(15)-Eudesmene-1β,11-diol (21). Mp 156–157°, $[\alpha]_D^{30}$

+ 56° (CHCl₃; c1.5), lit. [14], mp 156–157°; $[\alpha]_D^{31}$ + 56.4° (CHCl₃; c1.5).

4(15)-Eudesmene-1 β ,6 α -diol (22). Oil, $[\alpha]_D^{25} + 7.5^{\circ}$ (CHCl₃; c 1.2), lit. [15], gum; $[\alpha]_{436nm}^{24} + 16^{\circ}$ (CHCl₃; c 0.1). ¹³C NMR (CDCl₃, 75 MHz): δ 11.6 (C-14), 18.2 (C-8), 16.2 (C-12), 21.1 (C-13), 26.0 (C-11), 31.9 (C-2), 35.1 (C-3), 36.3 (C-9), 41.7 (C-10), 49.3 (C-7), 55.9 (C-5), 67.0 (C-6), 79.0 (C-1), 107.8 (C-15), 146.2 (C-4).

1β-Acetoxy-4(15)-eudesmen-11-ol (23). Oil, $[α]_{D}^{30} + 29^{\circ}$ (CHCl₃; c 2.2). TLC (EtOAc-CHCl₃-hexane, 5:50:45) R_f 0.27. IR v_{max}^{neat} cm⁻¹: 3453, 3079, 1711, 888. EIMS (70 eV) m/z (rel. int.): 202 [M - H₂O - HOAc]⁺ (15), 162 (100), 147 (65), 133 (20), 119 (18), 106 (16), 59 (18). HRMS for C₁₇H₂₈O₃ requires 280.2039; found 280.2021. Saponification of 23 (20 mg) by a procedure similar to that for 8 gave 21 (16 mg).

 α -Cadinol (24). Mp 70–71°, $[\alpha]_D^{25} - 45°$ (CHCl₃; c 0.8), lit. [17], mp 72.5°; $[\alpha]_D - 39.4°$ (CHCl₃; c 1.32).

T-Cadinol (25). Oil, $[\alpha]_D^{25} - 5^\circ$ (CHCl₃; c 1.2), lit. [18], oil; $[\alpha]_D^{30} - 4.7^\circ$ (CHCl₃; c 4.4).

Oplopanone (26). Mp 96–97°, $[\alpha]_D^{32} - 16°$ (CHCl₃; c 2.0), lit. [19], mp 96–97°; $[\alpha]_D^{25.5} - 20.0°$ (dioxane; c 0.571). ¹³C NMR (CDCl₃, 75 MHz): δ 15.6 (C-12), 20.2 (C-14), 21.9 (C-13), 23.0 (C-8), 25.3 (C-2), 28.6 (C-3), 29.4 (C-15), 29.5 (C-11), 42.0 (C-9), 46.7 (C-7), 49.4 (C-1), 55.7 (C-6), 57.0 (C-5), 73.0 (C-10), 211.4 (C-4).

Cedrol (27). Mp 80-81°, $[\alpha]_D^{25} + 3^\circ$ (CHCl₃; c 2.2), lit. [20], mp 86-87°; $[\alpha]_D^{28} + 9.9^\circ$ (CHCl₃; c 5). Acknowledgement—We thank the National Science Council for financial support (NSC83-0208-M002-095).

REFERENCES

- 1. Su, W.-C., Fang, J.-M. and Cheng, Y.-S. (1993) *Phytochemistry* 34, 779.
- Su, W.-C., Fang, J.-M. and Cheng, Y.-S. (1994) Phytochemistry 35, 1279.
- Su, W.-C., Fang, J.-M. and Cheng, Y.-S. (1994) Phytochemistry 37, 1109.
- Barrero, A. F., Sanchez, J. F., Alvarez-Manzaneda, R. E. J. and Munoz Dorado, M. (1989) *Phytochemistry* 28, 2617.
- 5. Zdero, C., Bohlmann, F. and Muller, M. (1987) *Phytochemistry* **26**, 2763.
- Wagh, A. D., Paknikar, S. K. and Bhattacharyya, S. C. (1964) Tetrahedron 20, 2647.
- Dolejs, L. and Herout, V. (1961) Colln Czech. Chem. Commun. 26, 2045.
- Nanayakkara, N. P. D., Kinghorn, A. D. and Farnsworth, N. R. (1986) J. Chem. Res. (S) 454.
- McQuillin, F. J. and Parrack, J. D. (1956) J. Chem. Soc. 2973.
- Takahashi, K. and Takani, M. (1976) Chem. Pharm. Bull. 24, 2000.
- Bohlmann, F., Knoll, K. H., Zdero, C., Mahanta, P. K., Grenz, M., Suwita, A. Ehlers, D., Van, N. L., Abraham, W. R. and Natu, A. A. (1977) *Phyto*chemistry 16, 965.
- 12. Minato, H. and Ishikawa, M. (1967) J. Chem. Soc. C. 423.
 - Humber, D. C., Pinder, A. R. and Williams, R. A. (1967) J. Org. Chem. 32, 2335.
 - 14. Adinarayana, D. and Syamasundar, K. V. (1982) Phytochemistry 21, 1083.
 - 15. Bohlmann, F., Ates(Goren), N., King, R. M. and Robinsin, H. (1983) Phytochemistry 22, 1675.
 - 16. Itokawa, H., Matsumoto, H. and Mihashi, S. (1983) Chem. Letters 1253.
 - 17. Herout, V. and Sykora, V. (1958) Tetrahedron 4, 246.
 - Cheng, Y.-S., Kuo, Y.-H. and Lin, Y.-T. (1967) J. Chem. Soc., Chem. Commun. 565.
 - Takeda, K., Minato, H. and Ishikawa, M. (1966) Tetrahedron Suppl. 7, 219.
 - Stork, G. and Clarke Jr, F. H. (1961) J. Am. Chem. Soc. 83, 3114.
 - Fiaud, J. C., Horeau, A. and Kagan, H. B. (1977) Stereochemistry, Fundamentals and Methods, Vol. 3, p. 52. Georg Thieme Publishers, Stuttgart.
- 22. van Beek, T. A., Kleis, R., Posthumus, M. A. and van Veldhuizen, A. (1989) Phytochemistry 28, 1909.