Sesquiterpenoids and Norsesquiterpenoids from the Formosan Soft Coral Lemnalia laevis

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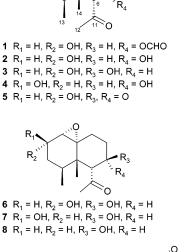
Eight new nornardosinane sequiterpenoids, laevinols A-H (1-8), a new neolemnane sesquiterpenoid, laevinone A (9), and the previously known 6β -acetyl- 4β , 5β -dimethyl- $1(10)\alpha$ -epoxy- 2β -hydroxy-7-oxodecalin (10) and 11,12-dihydroxy-6,10-eremophiladiene (11) were isolated from the methylene chloride solubles of the Formosan soft coral Lemnalia laevis. Their structures were elucidated by extensive spectroscopic analysis, and their cytotoxicity against selected cancer cells was measured in vitro.

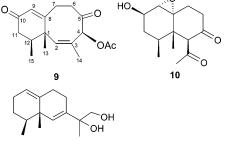
The genus Lemnalia has afforded a number of bioactive sesquiterpenes.¹⁻¹⁰ As part of our search for bioactive substances from marine organisms, the Formosan soft coral Lemnalia laevis Thomson and Dean (Nephtheidae) was studied because the CH₂Cl₂ extract showed significant cytotoxicity to A-549 (human lung adenocarcinoma), HT-29 (human colon adenocarcinoma), and P-388 (mouse lymphocytic leukemia) cell cultures, as determined by standard procedures.^{11,12} Bioassay-guided fractionation of the methylene chloride solubles of L. laevis resulted in the isolation and characterization of eight new nornardosinane sequiterpenoids, laevinols A-H (1-8), a new neolemnane sesquiterpenoid, laevinone A (9), and the previously known 6β -acetyl- 4β , 5β -dimethyl- $1(10)\alpha$ -epoxy- 2β -hydroxy-7-oxodecalin $(10)^2$ and 11,12-dihydroxy-6,10-eremophiladiene $(11).^{3}$

Results and Discussion

The molecular formula of laevinol A (1) was found to be $\mathrm{C_{15}H_{22}O_4}$ from its HREIMS and $^{13}\mathrm{C}$ NMR data. The DEPT spectrum showed signals for three methyls, three sp³ methylenes, four sp³ methines, one sp³ quaternary carbon, one sp² methine, and two sp² quaternary carbons. The ¹H and ¹³C NMR spectra indicated the presence of a secondary methyl at $\delta_{\rm H}$ 0.95 (3H, d, Me-13) and $\delta_{\rm C}$ 15.6 (CH₃, Me-13); a tertiary methyl at $\delta_{\rm H}$ 1.02 (3H, s, Me-14) and $\delta_{\rm C}$ 18.0 (CH₃, Me-14); a secondary hydroxyl at $\delta_{\rm H}$ 3.99 (1H, br s, H-2) and $\delta_{\rm C}$ 63.7 (CH, C-2); a secondary formyloxy group at $\delta_{\rm H}$ 5.44 (1H, dt, J = 12.0, 5.4 Hz, H-7), 8.04 (1H, s, OCHO) and $\delta_{\rm C}$ 71.6 (CH, C-7), 160.2 (CH, OCHO); a methyl ketone at $\delta_{\rm H}$ 2.19 (3H, s, H₃-12) and $\delta_{\rm C}$ 35.4 (CH₃, C-12), 210.5 (qC, C-11); and a trisubstituted olefin at $\delta_{\rm H}$ 5.69 (1H, d, J = 4.8 Hz, H-1) and $\delta_{\rm C}$ 123.9 (CH, C-1), 143.0 (qC, C-10). These spectroscopic data, coupled with the determined number of degrees of unsaturation (five), suggested that compound **1** is a bicyclic norsesquiterpenoid with secondary hydroxyl, secondary formyloxyl, and methyl ketone groups.

After assignments of all the direct ¹H-¹³C bondings were made on the basis of the HMQC spectrum, the gross structure of 1 was determined by ¹H-¹H COSY and HMBC NMR spectroscopic analysis (Figure 1). The ¹H-¹H COSY





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spectrum revealed two partial structures, **a** and **b** (Figure 1). The connectivity between C-11 and C-6 was indicated by the HMBC correlations from H-12 [$\delta_{\rm H}$ 0.88 (3H, s)] to C-11 [$\delta_{\rm C}$ 213.1 (qC)] and C-6 [60.8 (CH)] and from H-7 [$\delta_{\rm H}$ 5.44 (1H, dt, J = 11.4, 5.1 Hz)] to C-11. The HMBC correlation from H-13 to C-5 [$\delta_{\rm C}$ 35.9 (CH)] comfirmed the connectivity between C-4 and C-5. The HMBC correlations from the proton signal at δ 8.04 to C-7 and from H-7 to the carbon signal at δ 160.2 revealed the location of the secondary formyloxyl group.¹ The connectivity between partial structures **a** and **b** was exhibited by the HMBC correlations as shown in Figure 1.

The relative stereochemistry of 1 was deduced from a 2D NOESY NMR experiment (Table S1, Supporting Infor-

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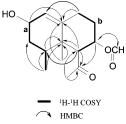


Figure 1. Key COSY and HMBC correlations of 1.

mation), which showed that Me-14 (axial), Me-13 (equatorial), H-6 (equatorial), and H-7 (axial) are on β face of the molecule, while H-4 (axial) and Me-12 are on the opposite, α face. The α -configuration of hydroxy at C-2 was determined by comparison with the $J_{1,2}$ value of lemnacarnol ($J_{1,2} = 0$ Hz) and 2-*epi*-lemnacarnol ($J_{1,2} = 4.5$ Hz).¹ From the aforementioned data, laevinol A could be formulated as 6α -acetyl- 4β , 5β -dimethyl-1(10)-ene- 2α -hydroxy- 7α -formyl-oxydecalin.

The HREIMS and ¹³C NMR data revealed laevinol B (2) to have a molecular formula of C₁₄H₂₂O₃. The DEPT spectrum showed resonances for three methyls, three sp³ methylenes, four sp³ methines, one sp³ quaternary carbon, one sp² methine, and two sp² quaternary carbons. The IR absorptions at 3520 and 1722 cm⁻¹ indicated the presence of hydroxyl and carbonyl groups. The ¹H and ¹³C NMR spectra showed the presence of a secondary methyl at $\delta_{\rm H}$ 0.95 (3H, d, J = 6.6 Hz, Me-13) and $\delta_{\rm C}$ 15.6 (CH₃, Me-13); a tertiary methyl at $\delta_{\rm H}$ 1.00 (3H, s, Me-14) and $\delta_{\rm C}$ 18.2 (CH₃, Me-14); two secondary hydroxyls at $\delta_{\rm H}$ 3.97 (1H, br s, H-2), 4.25 (1H, dt, J = 11.4, 5.1 Hz, H-7) and $\delta_{\rm C}$ 63.9 (CH, C-2), 69.0 (CH, C-7); a methyl ketone at $\delta_{\rm H}$ 2.34 (3H, s, H₃-12) and $\delta_{\rm C}$ 36.2 (CH₃, C-12), 213.1 (qC, C-11); and a trisubstituted olefin at $\delta_{\rm H}$ 5.66 (1H, d, J = 4.5 Hz, H-1) and $\delta_{\rm C}$ 123.3 (CH, C-1), 144.0 (qC, C-10). These spectroscopic data suggested that compound 2 is the deformylated derivative of 1. The COSY NMR correlations from H-7 to H-6 and H-8 revealed the location of the secondary hydroxyl group at C-7. The relative stereochemistry of 2 was established from a 2D NOESY experiment (Table S1, Supporting Information), which showed results similar to those determined from 1. Therefore, laevinol B was established as 6α -acetyl- 4β , 5β -dimethyl-1(10)-ene- 2α , 7α -dihydroxydecalin.

Laevinol C (3) was assigned a molecular formula of $C_{14}H_{22}O_3$, as indicated by its HREIMS and ¹³C NMR data. The ¹H and ¹³C NMR spectroscopic data resembled those of **2** except for the resonances and splitting patterns in the vicinity of C-7. The relative stereochemistry of **3** was deduced from a 2D NOESY experiment (Table S1, Supporting Information). NOESY correlations from Me-14 to H-6, H-9 β , H-8 β , and Me-13 indicated that Me-14 (axial), Me-13 (equatorial), and H-6 (equatorial) are on the β face of the molecule, while NOESY correlations from Me-12 to H-4 and from H-7 to H-8 α suggested that H-4 (axial), H-7 (equatorial), and Me-12 (axial) are on the opposite, α face. Therefore, laevinol C was determined as 6α -acetyl- 4β , 5β -dimethyl-1(10)-ene- 2α , 7β -dihydroxydecalin.

Laevinol D (4) was shown to have the molecular formula $C_{14}H_{22}O_3$ by HREIMS and from its ¹³C NMR data. The ¹H and ¹³C NMR spectra of 4 were quite similar to those of 2 with the exception of the resonances and splitting patterns in the vicinity of C-2. The configuration of the hydroxy group at C-2 was determined as β by comparison with the $J_{1,2}$ value of lemnacarnol ($J_{1,2} = 0$ Hz) and 2-*epi*-lemnacarnol ($J_{1,2} = 4.5$ Hz).¹ The relative stereochemistry of 4 was deduced from a 2D NOESY experiment (Table S1,

Supporting Information), which exhibited similar results as determined from 1. From the aforementioned data, laevinol D was formulated as 6α -acetyl- 4β , 5β -dimethyl-1(10)-ene- 2β , 7α -dihydroxydecalin.

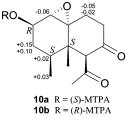
Laevinol E (5) gave a molecular formula of $C_{14}H_{20}O_3$ from the interpretation of its HREIMS and ¹³C NMR data. The ¹H and ¹³C NMR spectra of 5 were analogous to those of 2 and established that the secondary hydroxyl at C-7 in 2 was replaced by a ketone in 5. The HMBC correlations from H-6 to C-7 and from H-8 to C-7 helped position the ketone carbonyl at C-7. Therefore, laevinol E was determined as 6α -acetyl- 4β , 5β -dimethyl-1(10)-ene- 2α -hydroxy-7-oxodecalin.

The molecular formula of laevinol F (6) was found to be $C_{14}H_{22}O_4$ from its HREIMS and ¹³C NMR data. The DEPT spectrum showed the presence of signals for three methyls, three sp³ methylenes, five sp³ methines, two sp³ quaternary carbons, and one sp² quaternary carbon. The ¹H and ¹³C NMR spectra supported the presence of two secondary hydroxyls at $\delta_{\rm H}$ 3.98 (1H, t, J = 4.2 Hz, H-2), 4.18 (1H, br s, H-7) and $\delta_{\rm C}$ 63.6 (CH, C-2), 67.4 (CH, C-7); a methyl ketone at $\delta_{\rm H}$ 2.31 (3H, s, H_3-12) and $\delta_{\rm C}$ 31.8 (CH_3, C-12), 210.0 (qC, C-11); and a trisubstituted epoxide ring at $\delta_{\rm H}$ 3.14 (1H, d, J = 4.2 Hz, H-1) and $\delta_{\rm C}$ 61.1 (CH, C-1), 67.4 (qC, C-10). These data were similar to those of 6α -acetyl- 4β , 5β -dimethyl-1(10) α -epoxy- 2α -hydroxy-7-oxodecalin, but were consistent with the replacement of a ketone at C-7 by a secondary hydroxyl.⁶ The HMBC correlations from H-6 to C-7 and from H-8 to C-7 enabled the correct positioning of the secondary hydroxyl at C-7. The configurations of the epoxide ring and hydroxyl at C-2 were determined by comparison with the $J_{1,2}$ and $J_{2,3}$ values of 6α -acetyl- 4β , 5β dimethyl-1(10) α -epoxy-2 α -hydroxy-7-oxodecalin ($J_{1,2} = J_{2,3\beta}$ = 4.5 Hz, $J_{2,3\alpha} = 0$ Hz) and its 2-epimer.^{2,13} The relative stereochemistry of 6 was deduced from a 2D NOESY experiment (Table S1, Supporting Information), which exhibited results similar to those determined from 3. From these data, laevinol F could be formulated as 6α -acetyl- 4β , 5β -dimethyl-1(10) α -epoxy- 2α , 7β -dihydroxydecalin.

Laevinol G (7) was assigned a molecular formula of $C_{14}H_{22}O_4$, as indicated by its HREIMS and ¹³C NMR data. The ¹H and ¹³C NMR spectra of 7 were quite similar to those of **6** except for the resonances and splitting patterns in the vicinity of the secondary hydroxyl methine at C-2. The configurations of the epoxy and hydroxy group at C-2/C-3 were determined by comparison with $J_{1,2}$ and $J_{2,3}$ values of **6** and its 2-epimer.^{2,13} The relative stereochemistry of **7** was established from a 2D NOESY experiment (Table S1, Supporting Information), which showed results similar to those determined from **6**. Therefore, laevinol G was formulated as 6α -acetyl- 4β , 5β -dimethyl-1(10) α -epoxy- 2β , 7β -dihydroxydecalin.

Laevinol H (8) was shown to have a molecular formula of $C_{14}H_{22}O_3$ by HREIMS and from its ^{13}C NMR data. The ^{1}H and ^{13}C NMR spectroscopic data were similar to those of 7 except for the absence of the C-2 OH. HMBC correlations from H-1 to C-2/C-3 and from H-4 to C-2/C-3 confirmed the absence of the secondary hydroxyl at C-2. Therefore, laevinol H was assigned as 6α -acetyl- 4β , 5β -dimethyl-1(10) α -epoxy- 7β -hydroxydecalin.

Laevinone A (9) gave a molecular formula of $C_{17}H_{22}O_4$, from its HREIMS and ¹³C NMR data. The DEPT spectrum showed signals for four methyls, three sp³ methylenes, two sp³ methines, one sp³ quaternary carbon, two sp² methines, and five sp² quaternary carbons. Analysis of its ¹H, ¹³C, ¹H-¹H COSY, HMQC, and HMBC NMR spectral data revealed that **9** is a neolemnane sesquiterpene³ containing



 $\Delta \delta = \delta(S)$ -MTPA – $\delta(R)$ -MTPA (ppm)

Figure 2. ¹H NMR chemical shift differences [$\delta(S)$ -MTPA – $\delta(R)$ -MTPA] of the MTPA esters.

a secondary acetoxyl at $\delta_{\rm H}$ 5.98 (1H, s, H-4), 2.11 (3H, s) and $\delta_{\rm C}$ 76.1 (CH, C-4), 20.5 (CH₃), 170.0 (qC); a methylbearing trisubstituted olefin at $\delta_{\rm H}$ 5.55 (1H, s, H-2), 1.75 (3H, s, Me-14) and $\delta_C 136.9 (CH, C-2)$, 129.0 (qC, C-3), 18.1 (CH₃, C-14); a β , β -substituted enone at $\delta_{\rm H}$ 6.13 (1H, s, H-9) and $\delta_{\rm C}$ 128.7 (CH, C-9), 172.9 (qC, C-8), 197.6 (qC, C-10); and a secured ketone at $\delta_{\rm C}$ 200.6 (qC, C-5). HMBC NMR correlations from H-9 [$\delta_{\rm H}$ 6.13 (1H, s)] to C-10 [$\delta_{\rm C}$ 197.6 (qC)] and C-11 [$\delta_{\rm C}$ 39.1 (CH₂)] and from H₂-11 [$\delta_{\rm H}$ 2.38 (2H, m)] to C-10 were used to position the enone at C-8 through C-10. The location of the secondary acetoxy and ketone at C-4, C-5 was demonstrated by HMBC correlations from H-4 $[\delta_{\rm H} 5.98 \, (1H, s)]$ to C-5 $[\delta_{\rm C} 200.6 \, (qC)]$, from H-14 $[\delta_{\rm H} 1.75$ (3H, s)] to C-4 [δ_{C} 76.1 (CH)], and from H₂-6 [δ_{H} 2.73 (2H, m)] to C-5. The relative configuration of 9 was deduced from a 2D NOESY experiment. NOESY correlations between H-2 and Me-13, Me-14, and Me-15 indicated that Me-13, Me-14, Me-15, and H-2 are on the β face of the molecule. In turn, a NOESY correlation between H-4 and H-12 suggested that H-12 and H-4 are on the opposite side of the molecule. From the aforementioned data, laevinone A was established as $4(S^*)$ -acetoxy-5,10-dioxo, $1(S^*)$, $12(S^*)$ neolemna-2Z,8-diene.

The spectroscopic and physical data of 10 were identical with those of 6β -acetyl- 4β , 5β -dimethyl- $1(10)\alpha$ -epoxy- 2β hydroxy-7-oxodecalin² isolated from a soft coral, Lemnalia africana. To determine the absolute configuration, compound 10 was treated with (R)- or (S)- α -methoxy- α -trifluoromethylphenylacetyl chloride [(R)- or (S)-MTPA-Cl] in the presence of pyridine to yield the (R)- and (S)-MTPA esters (10b and 10a), respectively.¹⁴ The MTPA esters formed at C-2 were elucidated from the ¹H NMR chemical shifts and coupling constants of H-2 in 10a and 10b (10a, δ 5.33, 1H, t, J = 5.0 Hz, H-2; 10b, δ 5.32, 1H, t, J = 5.0Hz, H-2). Comparison of the ¹H NMR chemical shifts for **10a** and **10b** (Δ values shown in Figure 2) led to the assignment of the *R*-configuration at C-2. Therefore, the absolute structure of this compound was determined as shown in formula 10.

Compound **11** exhibited cytotoxicity against P-388 and HT-29 cell lines with ED_{50} values of 0.21 and 0.33 μ g/mL, respectively. The other isolates were not cytotoxic against P-388 and HT-29 cell lines ($ED_{50} > 5 \ \mu$ g/mL).

Experimental Section

General Experimental Procedures. Optical rotations were determined on a JASCO DIP-181 polarimeter. IR spectra were recorded on a Hitachi 26-30 spectrophotometer. The NMR spectra were recorded on a Bruker Avance 300 NMR spectrometer at 300 MHz for ¹H and 75 for ¹³C using CDCl₃ with TMS as internal standard. EIMS were obtained with a JEOL JMS-SX/SX 102A mass spectrometer at 70 eV. Silica gel 60 (Merck, 230–400 mesh) was used for column chromatography; precoated silica gel plates (Merck, Kieselgel 60 F₂₅₄, 0.25 mm) were used for TLC analysis.

Animal Material. The soft coral *L. laevis* was collected at Green Island, off Taiwan, in March 2003, at a depth of 3 m and was stored for 2 weeks in a freezer until extraction. A voucher specimen, NSUGN-062, was deposited in the Department of Marine Biotechnology and Resources, National Sun Yat-sen University, Taiwan.

Extraction and Isolation. The bodies of the soft coral L. *laevis* were freeze-dried to give 1.2 kg of a solid, which was extracted with CH_2Cl_2 (3.0 L \times 3, overnight for each cycle) at room temperature. After removal of solvent in vacuo, the residue (40 g) was chromatographed over a column containing silica gel 60, using n-hexane-EtOAc and MeOH-EtOAc mixtures as eluting solvents. Elution by n-hexane-EtOAc (65:35) afforded fractions containing 1 and 9. Elution by n-hexane-EtOAc (50:50) afforded fractions containing 2. Elution by n-hexane-EtOAc (30:70) afforded fractions containing 7, 10, and 11. Elution by n-hexane-EtOAc (1:3) afforded fractions containing 5 and 8. Elution by EtOAc afforded fractions containing 3, 4, and 6. Compound 1 (4 mg, 0.01%) was further purified using a RP-C₁₈ HPLC column, eluting with MeOH-H₂O (85:15). Compound 2 (5 mg, 0.01%) was further purified by silica gel column chromatography, eluting with MeOH-CH₂Cl₂ (98:2). Compound **3** (2 mg, 0.005%) was further purified by passage over a RP-C₁₈ HPLC column, eluting with MeOH-H₂O (52:48). Compound 4 (3 mg, 0.075%) was further purified by RP-C₁₈ HPLC separation, eluting with MeOH-H₂O (70:30). Compound **5** (1 mg, 0.0025%) was further purified using a RP-C₁₈ HPLC column, eluting with MeOH-H₂O (63:37). Compound 6 (3 mg, 0.0075%) was also purified further by RP-C₁₈ HPLC separation, eluting with MeOH-H₂O (52:48). Compound 7 (1 mg, 0.0025%) was further purified by silica gel column chromatography, eluting with MeOH-CH₂Cl₂ (95:5). Compound 8 (3 mg, 0.0075%) was further purified by RP-C₁₈ HPLC separation, eluting with MeOH $-H_2O$ (63:37). Compound 9 (5 mg, 0.01%) was further purified by silica gel column chromatography, eluting with $MeOH-CH_2Cl_2$ (85:15). Compound 10 (4 mg, 0.01%) was further purified by silica gel column chromatography, eluting with MeOH-CH₂Cl₂ (99:1). Finally, compound 11 (25 mg, 0.05%) was further purified by silica gel column chromatography, eluting with n-hexaneacetone (85:15).

Laevinol A (1): $[\alpha]^{25}_{D} -173$ (*c* 0.4, CHCl₃); IR (neat) ν_{max} 3449, 1720 cm⁻¹; ¹H NMR, see Table 1; ¹³C NMR, see Table 2; EIMS *m*/*z* 266 [M]⁺ (5), 248 (9), 202 (10), 176 (15), 232 (10), 201 (12), 173 (18), 160 (100); HREIMS *m*/*z* 266.1566 (calcd for C₁₅H₂₂O₄, 266.1569).

Laevinol B (2): $[\alpha]^{25}_{D} - 132$ (*c* 0.2, CHCl₃); IR (neat) ν_{max} 3520, 1722 cm⁻¹; ¹H NMR, see Table 1; ¹³C NMR, see Table 2; EIMS *m/z* 238 [M]⁺ (3), 220 (28), 202 (10), 160 (32), 120 (56), 55 (100); HREIMS *m/z* 238.1568 (calcd for C₁₄H₂₂O₃, 238.1563).

Laevinol C (3): $[\alpha]^{25}_{\rm D}$ -98 (c 0.1, CHCl₃); IR (neat) $\nu_{\rm max}$ 3480, 1718 cm⁻¹; ¹H NMR, see Table 1; ¹³C NMR, see Table 2; EIMS *m/z* 238 [M]⁺ (4), 220 (18), 202 (12); HREIMS *m/z* 238.1566 (calcd for C₁₄H₂₂O₃, 238.1563).

Laevinol D (4): $[\alpha]^{25}_{\rm D}$ –136 (*c* 0.3, CHCl₃); IR (neat) $\nu_{\rm max}$ 3448, 1723 cm⁻¹; ¹H NMR, see Table 1; ¹³C NMR, see Table 2; EIMS *m/z* 238 [M]⁺ (6), 220 (23), 202 (7), 160 (22); HREIMS *m/z* 238.1560 (calcd for C₁₄H₂₂O₃, 238.1563).

Laevinol E (5): $[\alpha]^{25}_{D} - 145$ (*c* 0.1, CHCl₃); IR (neat) ν_{max} 3520,1725 cm⁻¹; ¹H NMR, see Table 1; ¹³C NMR, see Table 2; EIMS *m/z* 236 [M]⁺ (8), 218 (12), 200 (10); HREIMS *m/z* 236.1403 (calcd for C₁₄H₂₀O₃, 236.1407).

Laevinol F (6): $[\alpha]^{25}_{D}$ -76 (*c* 0.2, CHCl₃); IR (neat) ν_{max} 3410, 1718 cm⁻¹; ¹H NMR, see Table 1; ¹³C NMR, see Table 2; EIMS *m*/*z* 254 [M]⁺ (6), 236 (10), 218 (8), 190 (30), 174 (52), 106 (100); HREIMS *m*/*z* 254.1515 (calcd for C₁₄H₂₂O₄, 254.1512).

Laevinol G (7): $[\alpha]^{25}_{D}$ –166 (*c* 0.3, CHCl₃); IR (neat) ν_{max} 3509, 1722 cm ⁻¹; ¹H NMR, see Table 1; ¹³C NMR, see Table 2; EIMS *m/z* 254 [M]⁺ (4), 236 (11), 218 (7), 190 (33), 106 (100); HREIMS *m/z* 254.1516 (calcd for C₁₄H₂₂O₄, 254.1512).

Laevinol H (8): $[\alpha]^{25}_{\rm D}$ -82 (*c* 0.1, CHCl₃); IR (neat) $\nu_{\rm max}$ 3430, 1725 cm ⁻¹; ¹H NMR, see Table 1; ¹³C NMR, see Table 2; EIMS *m/z* 238 [M]⁺ (8), 220 (12), 202 (9); HREIMS *m/z* 238.1569 (calcd for C₁₄H₂₂O₃, 238.1563).

Table 1. ¹H NMR Spectroscopic Data of Compounds 1-8 in CDCl₃^a

| position | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 |
|------------------------|--|----------------------------------|----------------------------------|----------------------------------|----------------------------------|--|----------------------------------|----------------------------------|
| 1 | $5.69 d (4.8)^b$ | 5.66 d (4.5) | $5.68 d (4.8)^b$ | 5.48 br s | 5.86 d (5.1) | 3.14 d (4.2) | 2.86 br s | 2.88 d (3.6) |
| 2 | 3.99 br s | 3.97 br s | 3.99 br s | $4.25 \mathrm{m}$ | $4.08 \mathrm{\ br\ s}$ | 3.98 t (4.2) | 4.11 t (7.8) | 1.88 m, 2.30 m |
| 3 | 1.52 m | 1.53 m | 1.60 m | 1.35 m, 1.74 m | 1.65 m | 1.36 m 1.58 m | 1.31 m 1.78 m | 1.28 m |
| 4 | 1.85 m | 1.84 m | 1.90 m | 1.65 m | 2.15 m | 1.99 m | 1.86 m | 1.97 m |
| 6 | 3.42 d (5.4) | 3.45 d (5.1) | 3.28 br s | 3.40 d (5.4) | $3.93 \mathrm{s}$ | $3.07 \mathrm{\ br\ s}$ | 3.00 br s | 2.97 br s |
| 7 | 5.44 dt (12.0, 5.4) | 4.25 dt (11.4, 5.1) | $4.11 \mathrm{ \ br \ s}$ | $4.25 \mathrm{m}$ | | 4.18 br s | 4.19 br s | 4.18 m |
| 8 | 1.89 m, 2.03 m | 1.77 m, 2.39 m | 1.72 m, 1.91 m | 1.77 m, 1.95 m | 2.48 m, 2.85 m | 1.79 m, 2.26 m | 1.83 m, 2.29 m | 1.75 m, 2.32 m |
| 9 | 2.29 m, 2.45 m | 1.90 m, 2.24 m | 2.08 m, 2.20 m | 2.30 m, 2.40 m | 2.65 m | 2.54 dt (13.5, 4.2) 1.05 dt (13.5, 1.5) | 2.53 dt (14.4, 4.8) 1.15 m | 1.01 m, 2.53 m |
| 12 13 14 OCHO | 2.19 s 0.95 d (6.9) 1.02 s 8.04 s | 2.34 s 0.95 d (6.6) 1.00 s | 2.24 s 0.92 d (6.9) 1.17 s | 2.21 s 0.96 d (6.6) 1.05 s | 2.22 s 0.93 d (6.9) 0.91 s | 2.31 s 0.75 d (6.9) 1.31 s | 2.31 s 0.78 d (6.9) 1.34 s | 2.33 s 0.73 d (6.6) 1.27 s |

^a Recorded at 300 MHz (assigned by COSY, HSQC, and HMBC experiments). ^b J values (in Hz) in parentheses.

Table 2. ¹³C NMR Spectroscopic Data of Compounds 1-9 in CDCl₃^{*a*}

| | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 |
|------|-------|-------|-------|-------|-------|-------|-------|-------|-------|
| 1 | 123.9 | 123.3 | 122.5 | 126.6 | 124.7 | 61.1 | 61.8 | 58.6 | 45.1 |
| 2 | 63.7 | 63.9 | 63.9 | 67.4 | 63.7 | 63.6 | 64.8 | 21.8 | 136.9 |
| 3 | 35.1 | 35.2 | 35.3 | 36.6 | 35.5 | 35.8 | 34.9 | 24.4 | 129. |
| 4 | 29.7 | 29.8 | 29.7 | 34.3 | 28.7 | 27.1 | 26.1 | 29.9 | 76. |
| 5 | 42.6 | 41.9 | 40.3 | 42.0 | 46.6 | 37.7 | 37.1 | 38.2 | 200. |
| 6 | 57.6 | 60.8 | 63.1 | 60.7 | 74.8 | 62.0 | 62.6 | 63.4 | 43. |
| 7 | 71.6 | 69.0 | 67.0 | 69.3 | 202.9 | 67.4 | 67.2 | 67.9 | 29. |
| 8 | 26.2 | 30.2 | 29.7 | 30.3 | 37.6 | 27.2 | 27.4 | 27.4 | 172. |
| 9 | 29.7 | 30.1 | 26.2 | 29.9 | 31.3 | 25.1 | 24.8 | 25.4 | 128. |
| 10 | 143.0 | 144.0 | 145.8 | 140.8 | 141.8 | 67.4 | 63.2 | 63.8 | 197. |
| 11 | 210.5 | 213.1 | 210.5 | 212.0 | 205.0 | 210.0 | 207.2 | 208.7 | 42. |
| 12 | 35.4 | 36.2 | 32.8 | 35.8 | 33.3 | 31.8 | 30.8 | 31.3 | 39. |
| 13 | 15.6 | 15.6 | 15.9 | 15.7 | 15.6 | 15.7 | 15.1 | 16.0 | 20. |
| 14 | 18.0 | 18.2 | 20.0 | 19.8 | 18.3 | 18.4 | 18.4 | 18.6 | 18. |
| 15 | | | | | | | | | 16. |
| OCHO | 160.2 | | | | | | | | |
| OAc | | | | | | | | | 170. |
| | | | | | | | | | 20. |

^a Recorded at 75 MHz (assigned by DEPT, COSY, HSQC, and HMBC experiments).

Laevinone A (9): $[\alpha]^{25}_{D}$ +165 (*c* 0.4, CHCl₃); UV λ_{max} (log $\epsilon)$ 232 (3.2) nm; IR (neat) $\nu_{\rm max}$ 1724, 1667 cm^-1; ¹H NMR (300 MHz, CDCl₃) δ 1.07 (3H, d, J = 6.6 Hz, Me-15), 1.20 (3H, s, Me-13), 1.75 (3H, s, Me-14), 2.11 (3H, s, OCOCH₃), 2.38 (2H, m, H₂-11), 2.40 (1H, m, H-7 β), 2.54 (1H, dq, J = 7.0, 2.0 Hz, H-12), 2.62 (1H, m, H-7a), 2.73 (2H, m, H₂-6), 5.55 (1H, s, H-2), 5.98 (1H, s, H-4), 6.13 (1H, s, H-9); ¹³C NMR, see Table 2; EIMS m/z 290 [M]⁺ (4), 272 (5), 230 (18), 188 (16), 91 (100); HREIMS m/z 290.1599 (calcd for $C_{17}H_{22}O_4$, 290.1596).

(R)- and (S)-MTPA Derivatives of 10. To a solution of compound 10 (1.0 mg, 4.0×10^{-3} mmol) in pyridine (0.5 mL) at room temperature was added (R)-MTPA-Cl (2.4 mg, $1.0 \times$ 10⁻² mmol), and the resultant mixture was stirred for 24 h at room temperature. The reaction mixture was worked up by adding 2 mL of water. Further purification was performed with a short silica gel column with CH₂Cl₂ to give **10b** (0.6 mg) as a colorless oil. The (S)-MTPA ester 10a (0.5 mg) was prepared in the same way. Selected $\Delta\delta$ values $[\delta (S) - \delta (R)]$ are as follows: H-1 = -0.06, $H-3\alpha = +0.10$, $H-3\beta = +0.15$, H-4 =+0.02, H-9 α = -0.02, H-9 β = -0.05, H₃-13 = +0.03.

(S)-MTPA ester of 10: ¹H NMR (CDCl₃, 300 MHz) δ 0.70 $(3H, d, J = 6.9 Hz, H_3-13), 1.24 (3H, s, H_3-14), 1.45 (1H, m, m)$ H-3a), 1.46 (1H, m, H-9a), 1.70 (1H, m, H-3b), 2.15 (1H, m, H-4), 2.22 (3H, s, H₃-12), 2.47 (1H, s, H-9β), 2.50 (1H, m, H-8β), 2.76 (1H, m, H-8a), 3.58 (3H, OMe), 3.57 (1H, s, H-1), 3.74 (1H, s, H-6), 5.33 (1H, t, J = 5.0 Hz, H-2), 7.40-7.65 (5H, t)aromatic H).

(**R**)-MTPA ester of 10: ¹H NMR (CDCl₃, 300 MHz) δ 0.67 $(3H, d, J = 6.9 Hz, H_3-13), 1.24 (3H, s, H_3-14), 1.35 (1H, m, m)$ H-3α), 1.48 (1H, m, H-9α), 1.60 (1H, m, H-3β), 2.13 (1H, m, H-4), 2.23 (3H, s, H₃-12), 2.52 (1H, s, H-9β), 2.53 (1H, m, H-8β), 2.72 (1H, m, H-8a), 3.63 (1H, s, H-1), 3.66 (3H, OMe), 3.74 (1H, s, H-6), 5.32 (1H, t, J = 5.0 Hz, H-2), 7.39–7.66 (5H, aromatic H).

Cytotoxicity Testing. P-388 (mouse lymphocytic leukemia) cells were kindly supplied by J. M. Pezzuto, Department of Medicinal Chemistry and Pharmacognosy, University of Illinois at Chicago; A-549 (human lung adenocarcinoma) and HT-29 (human colon adenocarcinoma) were purchased from the American Type Culture Collection. Cytotoxic assays were carried out according to the procedure described previously.³ Three concentrations (50, 5, and 0.5 μ g/mL) of the tested compounds were used in the cytotoxicity assays.

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Supporting Information Available: NOESY correlations of 1-9 are available free of charge via the Internet at http://pubs.acs.org.

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