

1 **Microencapsulation of probiotics and chiu-niang extract in**
2 **Kou Woan Lao (an oriental style dairy product) by**
3 **spray-drying**

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13 Key words: probiotics, lao-chao, Kou Woan Lao, microencapsulation
14 spray-drying,

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ABSTRACT

26
27 This objective was to combine the physiological functionality of
28 probiotics and the unique flavor and milk-clotting activity of culture
29 filtrate from lao-chao for the development of a new dairy product so
30 different from the commercial yogurt. Two probiotic strains,
31 *Lactobacillus acidophilus* and *Bifidobacterium longum* were applied to
32 skim milk before acid-curd formation, and then culture filtrates were
33 added for the formation of milk curd. The higher percentage of SNF, the
34 stronger curd firmness ($P < 0.05$). The percentage of syneresis decreases
35 significantly when the level of SNF rising ($P < 0.05$). As the SNF
36 increasing from 10% to 18%, the percentage of syneresis decreases from
37 12.9% to 4.3%. The pH value and probiotic counts of 1% *L. acidophilus*
38 group did not significantly decrease during 14 days storage at 4 °C. The
39 counts of probiotics decrease about 1 log CFU/ml after spray-drying
40 process at 50 °C outlet temperature. The final total probiotic counts are
41 about 6.58 log CFU/ml. Regarding resuspension methods, it is found that
42 there are no significant difference of using stomacher or vortex to release
43 probiotics ($P > 0.05$). The counts of probiotics significantly decrease when
44 the outlet temperature getting higher. Therefore, when the spray-drying
45 outlet temperature cools down, the survival percentage of probiotics

46 increases. In order to increase the probiotic counts of Kou Woan Lao, it is
47 suggested to choose the lower temperature of microencapsulation. In the
48 same time, the product quality and storage period could be effected due to
49 the spray-drying process. It will be a challengeable task to adjust different
50 production procedure in order to meet market needs.

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INTRODUCTION

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66 The use of *Lactobacillus acidophilus* and/or *Bifidobacterium* spp. in
67 fermented or culture-containing dairy products became popular by the
68 end of the 1970s as a result of the increase in knowledge encompassing
69 the properties of bifidobacteria. The nutritional benefits of probiotics
70 have been mostly studied in milk-based products fermented with
71 lactobacilli and bifidobacteria. They are characterized by a lower level of
72 residual lactose and higher levels of free amino acids and certain vitamins
73 than non-fermented milks. Furthermore, they preferentially contain
74 L(+)-lactic acid (that is more easily metabolized by human beings than
75 D(-)-lactic acid) produced by bifidobacteria in addition to acetic acid.
76 Moreover, the L(+)-lactic acid absorbed in the intestine is used as energy
77 source, with an energy yield of 15 kJ/g, that compares well with 16 kJ/g
78 for lactose(Gurr, 1987). During the past years, numerous healthy and
79 nutritional benefits have been claimed including maintenance of normal
80 intestinal microflora, alleviation of lactose intolerance, reduction of
81 serum cholesterol levels, potential antitumor activity, and some
82 therapeutic effects on intestinal disturbances and intestinal infections. The
83 intact intestinal epithelium, together with the normal intestinal microflora
84 represents a barrier to movement of pathogenic bacteria, antigens and

85 other noxious substances from the gut lumen to the blood. In healthy
86 subjects this barrier is stable, thus protecting the host and assuring normal
87 intestinal function. When either the normal microflora or the epithelial
88 cells are disturbed, as triggered by dietary antigens, pathogens, chemicals
89 or radiation, defects in the barrier mechanisms become apparent; altered
90 permeability facilitates blood invasion by pathogens, foreign antigens and
91 other harmful substances (Gomes and Malcata, 1999). Experiments
92 performed in animal models showed that a few strains of *L. acidophilus*
93 and *Bifidobacterium* spp. are able to decrease the levels of enzymes
94 responsible for activation of some procarcinogens, and consequently
95 decrease the risk of tumor development. (Mital and Garg, 1992). A
96 number of studies indicate that administration of bifidobacteria or
97 lactobacilli alone or with fermentable carbohydrate (defined as a prebiotic)
98 can alter colonic microflora populations and decrease the development of
99 early preneoplastic lesions and tumors. Many of the antitumor activities
100 attributed to lactic cultures have been suggested to involve an enhanced
101 function of the immune response (Hirayama and Rafter, 2000). The
102 anticarcinogenic, antitumorigenic, and antimutagenic activities of these
103 probiotic bacteria have been proposed to occur via some modification of
104 mutagens *in vitro* (Vorobeva *et al.*, 1995), reduction in

105 carcinogen-generating fecal enzymes *in vivo* (Kulkarni and Reddy, 1994),
106 and suppression of precancerous lesion or tumor formation (Reddy and
107 Rivenson, 1993).

108 A new oriental-style dairy product coagulated with culture filtrates
109 from lao-chao (chiu-niang), a fermented rice product well known in
110 China, has been developed to meet consumer preferences for a low-acid
111 or non-sour tasting yogurt-like product (Kuo *et al.*, 1996 ; Lin and Chen,
112 1996). Culture filtrates from lao-chao, which is produced through
113 fermentation by inoculating steamed glutinous rice with commercial
114 starter (chiu-yao) or fungal cultures, have been used as both milk-clotting
115 agents and flavoring agents. The resultant yogurt-like products with soft
116 curd appearance are characterized by a sweet, fruity and slightly alcoholic
117 flavor. Therefore, this objective was to combine the physiological
118 functionality of probiotics and the unique flavor and milk-clotting activity
119 of culture filtrate from lao-chao for the development of a new dairy
120 product so different from the commercial yogurt.

121 Microencapsulation is a technique in which a membrane encloses
122 small particles of solid, liquid, or gas, with the objective of offering
123 protection to the sensitive core material from adverse environmental
124 conditions such as undesirable effects of light, moisture, and oxygen, thus

125 contributing to an increase in the shelf life of the product and promoting a
126 controlled liberation of the encapsulate (Shahidi and Han, 1993). Many
127 methods for preparing microcapsules have been developed and improved
128 significantly. They can briefly classify as mechanical and chemical
129 processes (Thies, 1996). Spray-drying is the microencapsulation
130 technique most widely used in the chemical, pharmaceutical, and food
131 industries (Gibbs, 1999). Spray-drying emulsions is a particularly
132 effective means of microencapsulating chemically reactive oils, volatile
133 oils, and flavor compounds. Spray-dried powder particles contain the
134 encapsulated material as minute droplets embedded within its wall
135 (McNamee *et al.*, 1998).

136 Gum arabic (gum *Acacia*) is a hydrocolloid produced by the natural
137 exudation of arabic trees and is an effective encapsulation agent due to
138 its high water solubility, the low viscosity of concentrated solutions
139 relative to other hydrocolloid gums, and its ability to act as an
140 oil-in-water emulsifier. Gum arabic is composed of a highly branched
141 arrangement of the simple sugars galactose, arabinose, rhamnose, and
142 glucuronic acids (Street and Anderson, 1983) and also contains a protein
143 component (~2%w/w) covalently bound within its molecular arrangement
144 (Anderson *et al.*, 1985). The protein fraction plays a crucial role in
145 determining the functional properties of gum arabic (Randall *et al.*, 1988).

146 Maltodextrins and chemically modified starches have been investigated
147 as replacers for gum arabic in spray-dried emulsions (Amandaraman and
148 Reineccius, 1987). To wholly or partially replace gum arabic as an
149 encapsulating agent.

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MATERIALS & METHODS

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167 **Preparation of inocula**

168 The two strains of fungal cultures, *Rhizopus javanicus* (CCRC 30288)
169 and *Saccharomyces cerevisiae* (CCRC 21685), from the Culture
170 Collection and Research Center, Taiwan, R. O. C., were used to inoculate
171 steamed glutinous rice. Before use, *R. javanicus* and *S. cerevisiae* were
172 transferred to slants of YM agar and those of potato dextrose agar,
173 respectively, and incubated at 30±1 for 6-8 days. In addition, the
174 probiotics strains, *Lactobacillus acidophilus* (CCRC 14079) and
175 *Bifidobacterium longum* (CCRC 14605) were transferred to Lactobacilli
176 MRS (deMan, Rogosa and Sharpe) and MGL (modified Garche's
177 lithium-chloride) media. Fungal spore suspensions and yeast cell
178 suspensions for inoculation were prepared by adding sterilized distilled
179 water containing 0.1 gL⁻¹ Tween 80 to slants and shaking the cultures
180 vigorously for 1 min.

181 **Preparation of fermented rice (lao-chao) and culture filtrates**

182 Glutinous rice was purchased from a local market (Taipei, Taiwan,
183 R.O.C.). One hundred grams of glutinous rice, which had been washed
184 with distilled water and drained, was soaked in 75 ml distilled water at
185 room temperature (20-25) for 12 hr, sterilized at 121 for 15 min, and
186 then cooled to 35 . The steamed glutinous rice was inoculated with 5 ml
187 suspensions containing 10⁷ cells of *S. cerevisiae* and 10⁷ spores of *R.*

188 *javanicus* and mixed completely, followed by static incubation at 30±1
189 for the designated period. After fermentation, the culture filtrates were
190 obtained by filtering the fermented rice through 4 layers of cheesecloth.
191 The sediment in the culture filtrate was eliminated by centrifuging at
192 1,480×g for 30 min (Kubota, KR-20000T, Japan).The culture filtrates
193 were stored at 4 for further analysis.

194 **Preparation of Kou Woan Loa and determination of viable probiotics**

195 Pasteurized skimmilk contains various ratios of SNF was inoculated
196 with probiotics and then incubated at 37 . The viable probiotics were
197 determined during the periods of incubation and storage (4) by
198 according to the method of Teraguchi *et al.*(1982).

199 **Determination of the firmness of product**

200 The firmness, as measured by the breaking force of the Kou Woan
201 Loa product, was determined by using a rheometer (Fudoh
202 NRM-2010J-CW, Tokyo 141, Japan) with a rheoplotter (Rikadenki Kigyo,
203 FR 801, Tokyo 113, Japan). Adaptor No.4 (20 mm diam.) of the
204 rheometer was used and table speed was 50 mm min⁻¹.

205 **Viscosity of Kou Woan Loa**

206 The apparent viscosity was determined by using a Viscometer
207 (Brookfield, LVDV-II + Viscometer, U. S. A.) with a No. 3 disc spindle at
208 a speed of 0.3rpm.

209 **Encapsulation procedure**

210 **Preparation of probiotics and culture filtrates emulsion**

211 All glassware, and solution used in the protocols were sterilized at
212 121 for 15 min. Gum arabic obtained from Hayashi Pure Chemical
213 Industries, Ltd. (Osaka, Japan). Maltodextrins and chemically modified
214 starches obtained from Gemfont Co. (Taipei, Taiwan, R.O.C.). Emulsion
215 with different gum arabic ratios were prepared and agitated with magnetic
216 stirrer bars for 30 min.

217 **Spray-Drying the Emulsion**

218 The emulsions were spray-dried using a laboratory mini spray drier
219 (model B-190, Buchi Co., Flawil, Switzerland) by control inlet air
220 temperature, compressed air for spray-flow, compressed air consumption
221 and feed rate to obtain outlet temperatures ranging from 50 to 60 . The
222 residence time of the product was very low. The dried powders were
223 collected and analysis immediately or subsequently stored in sealed glass
224 bottles at 4 .

225 **Determination of probiotic viability in spray-dried powders**

226 We assessed the viability of the probiotics in the inoculated
227 emulsions preparations before spray drying and in the resulting powders
228 by examining MRS and MGL pour plates after 72 hrs of incubation at
229 37°C (aerobically or anaerobically). To 1g of powder, 9ml of diluents

230 were added (1:10 dilution); the powders were allowed to rehydrate by
231 stomacher (Stomacher 400 Lab Blender, Sewad Medical, London, UK) at
232 normal speed (230 rpm 65%) for 1 min or by mixing with a vortex (Maxi
233 Mix II, Barnstead Thermolyne, Iowa, U.S.A.) for 1 min to release
234 microencapsulated probiotics and then diluted further with diluent, and
235 appropriate dilutions were pour plated.

236 **Statistical analysis**

237 Data were analyzed using the general linear model procedure of the
238 SAS software package (SAS/STAT, 1987), and Duncan's multiple range
239 test (Montgomery, 1991) were used to detect differences between
240 treatment means, statistical significance was tested at the 5% level. All
241 experiments were replicated three times.

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RESULTS & DISCUSSIONS

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251 Two probiotic strains, *Lactobacillus acidophilus* (CCRC 14079) and
252 *Bifidobacterium longum* (CCRC 14605) were applied to skim milk before
253 acid-curd formation. Different culture ratios on total probiotic counts
254 during incubation period examined are shown in Fig. 1. In the beginning
255 of incubation period, the counts of 1% *L. acidophilus* was lower than the
256 other three treatments significantly ($P < 0.05$). When the single starter
257 treatment compares with the mix of 1% *L. acidophilus* and 1% *B. longum*
258 treatments, the counts of *L. acidophilus* was lower than the counts of *B.*
259 *longum* significantly ($P < 0.05$). Synergistic growth-promoting effects
260 between *L. acidophilus* and *B. longum* are occur. *L. acidophilus* is
261 beneficial to *B. longum* as the former organism acts as an oxygen
262 scavenger.

263 According preliminary screening tests for suitable fungal and yeast
264 cultures, *Rhizopus javanicus* (CCRC 30288) and *Saccharomyces*
265 *cerevisiae* (CCRC 21685) were selected as starters of lao-chao and the
266 resultant culture filtrates were used as the milk-clotting agents and
267 flavoring agents. Effects of various ratios of culture filtrate on pH value
268 and titratable acidity in Kou Woan Lao during incubation period are
269 shown in Fig. 2. As the incubation period increasing, the pH value
270 decreases and titratable acidity rising gradually since the number of

271 probiotics increase. However, when the percentage of culture filtrate
272 increases, the growth of probiotics is inhibited. As long as the percentage
273 of culture filtrate getting higher, the increasing level of the titratable
274 acidity decreases. If the percentage of culture filtrate was controlled in
275 40%, the titratable acidity was keeping in the same level. The acidity
276 increases significantly during the 30th to 36th incubation hours.

277 During the 18th –36th incubation hour, there are no significant
278 differences among the four SNF treatments on probiotic counts of Kou
279 Woan Lao (Fig. 3).

280 Fig. 4 shows the higher percentage of SNF, the stronger curd
281 firmness ($P < 0.05$). When the SNF of skim milk increasing from 10% to
282 16%, the curd firmness grows from 18.9g to 44.6g. In the mean time, the
283 viscosity significantly increases from 12200cps to 29800cps ($P < 0.05$). In
284 Fig. 5, the percentage of syneresis decreases significantly when the level
285 of SNF rising ($P < 0.05$). As the SNF increasing from 10% to 18%, the
286 percentage of syneresis decreases from 12.9% to 4.3%.

287 These results are similar to the finding of Parnell-clunies *et al.*
288 (1988). According to Parnell-clunies *et al.* (1988), the curd firmness and
289 viscosity depend on if the structure of casein micelle is complete or not.
290 They also indicate the curd firmness has positive relationship with the
291 viscosity.

292 In order to produce better quality of Kou Woan Lao, the increase of
293 SNF will help to improve the curd firmness and viscosity. It is also
294 helpful to decrease the percentage of syneresis. However, in order to meet
295 consumer flavor expectation and lower the production cost, it is
296 suggested to keep SNF between 12 and 14%.

297 After adding culture filtrate, the total probiotics counts in other three
298 treatment groups decrease significantly ($P < 0.05$), except the 1% *L.*
299 *acidophilus* group (Fig. 6). The counts of 1% *B. longum* group decrease
300 from 6.3 log CFU/ml to 6.0 log CFU/ml. The 1% *B. longum* group has
301 dramatically decreased when compared with other groups. The pH value
302 and probiotic counts of 1% *L. acidophilus* group did not significantly
303 decrease during 14 days storage at 4 °C.

304 In Table 1, the counts of probiotics decrease about 1 log CFU/ml
305 after spray-drying process at 50 °C outlet temperature. The final total
306 probiotic counts are about 6.58 log CFU/ml. Regarding resuspension
307 methods, it is found that there are no significant difference of using
308 stomacher or vortex to release probiotics ($P > 0.05$). It is attribute to the
309 high solubility of gum arabic (about 50 %).

310 The viability of probiotics differs in various spray-drying outlet
311 temperature showed as Fig. 7. The counts of probiotics significantly
312 decrease when the outlet temperature getting higher. Therefore, when the

313 spray-drying outlet temperature cools down, the survival percentage of
314 probiotics increases. However, it may cause the poor condition of
315 spray-drying process, which will decrease the efficiency of
316 microencapsulation and the stability of storage.

317 There are four separate phases of spray-drying methods : 1.
318 Nebulization of the solution in the form of an aerosol. 2. Contact of the
319 nebulized solution with the warm air. 3. Drying of the aerosol. 4.
320 Separation of the dried product and the air charged with the solvent.
321 During these process, the probiotics are easily vanished. In order to
322 increase the probiotic counts of Kou Woan Lao, it is suggested to choose
323 the lower temperature of microencapsulation. In the same time, the
324 product quality and storage period could be effected due to the
325 spray-drying process. It will be a challengeable task to adjust different
326 production procedure in order to meet market needs.

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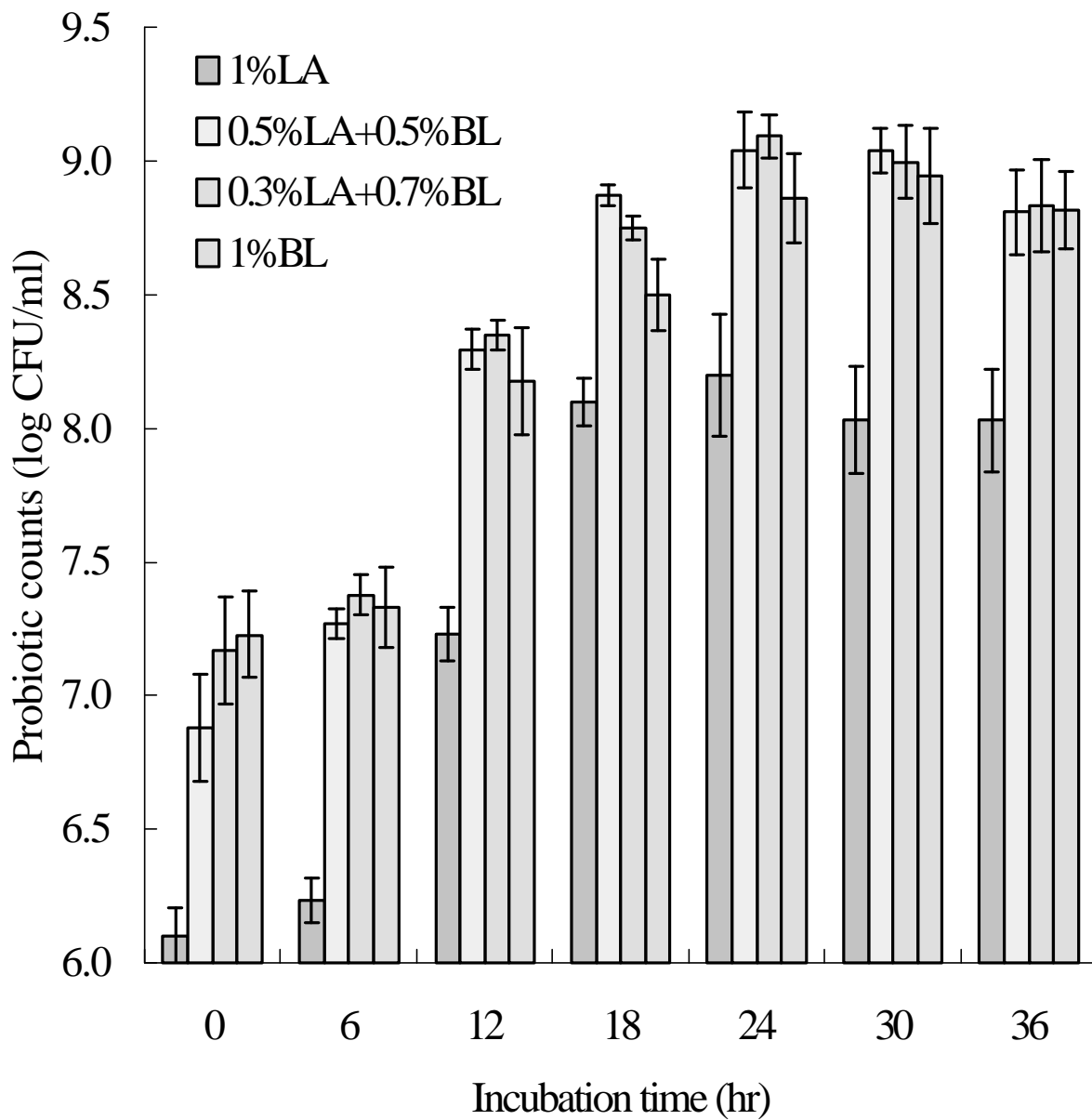


Fig. 1. Effects of different culture ratios on total probiotic counts in skim milk during incubation period (n = 9 ; error bars indicate 95% confidence intervals).
 LA : *L. acidophilus*. BL : *B. longum*.

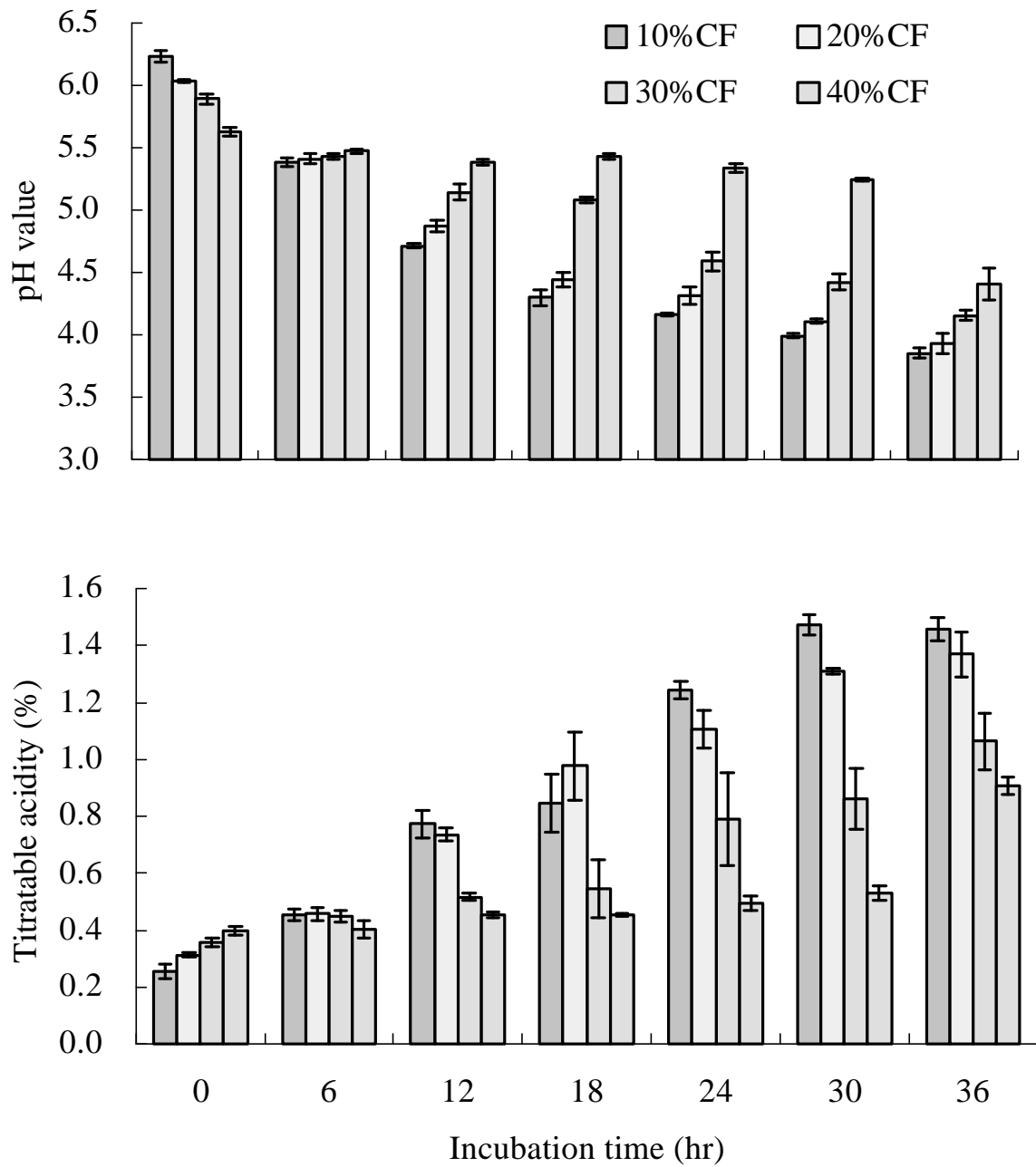


Fig. 2. Effects of culture filtrate on pH value and titratable acidity in Kou Woan Lao during incubation period (n = 9 ; error bars indicate 95% confidence intervals).
*CF : culture filtrate.

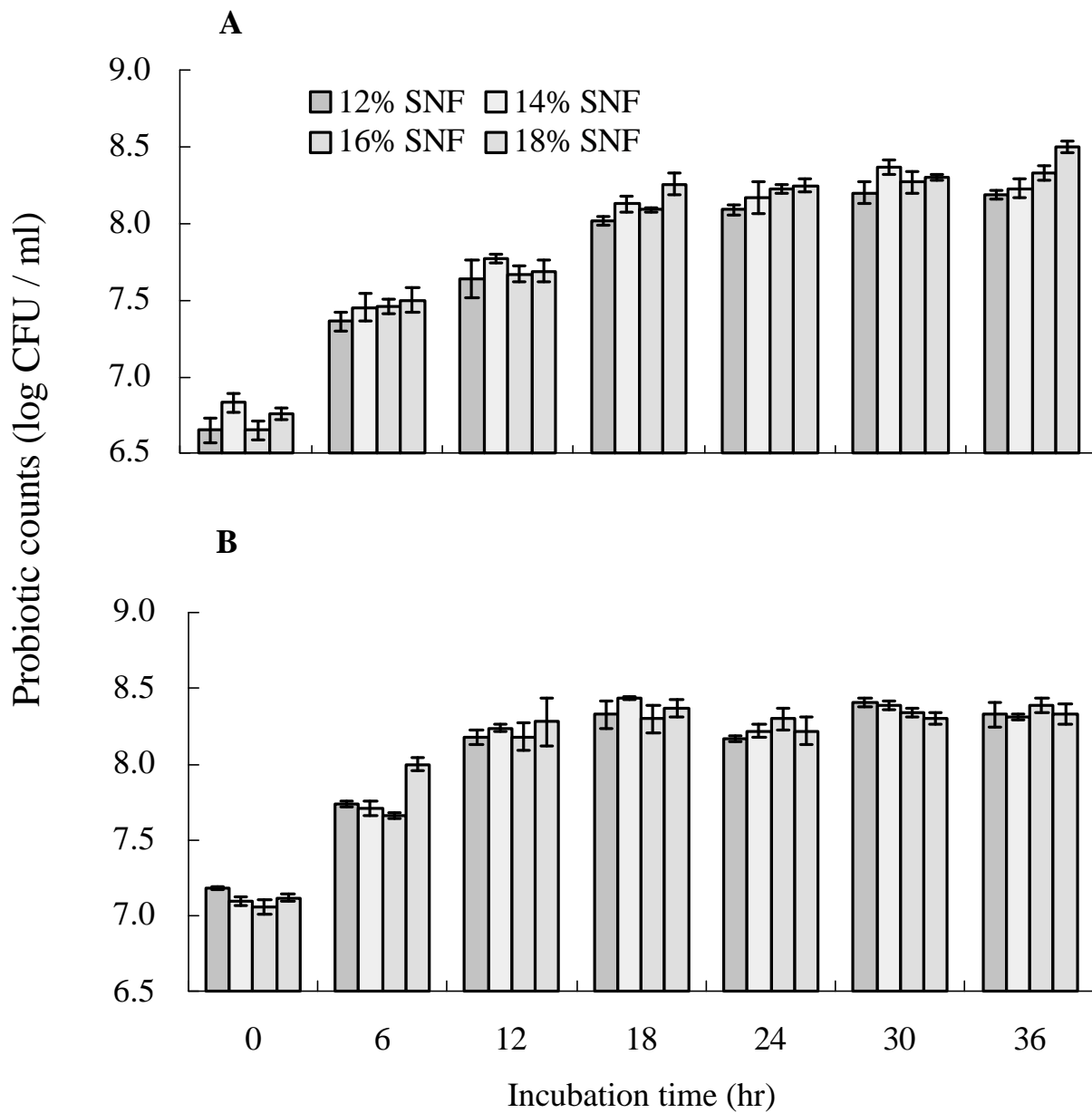


Fig. 3. Effects of solids-not-fat on probiotic counts in Kou Woan Lao during incubation period (n = 9 ; error bars indicate 95% confidence intervals).

*A : *L. acidophilus* .

B : *B. longum* .

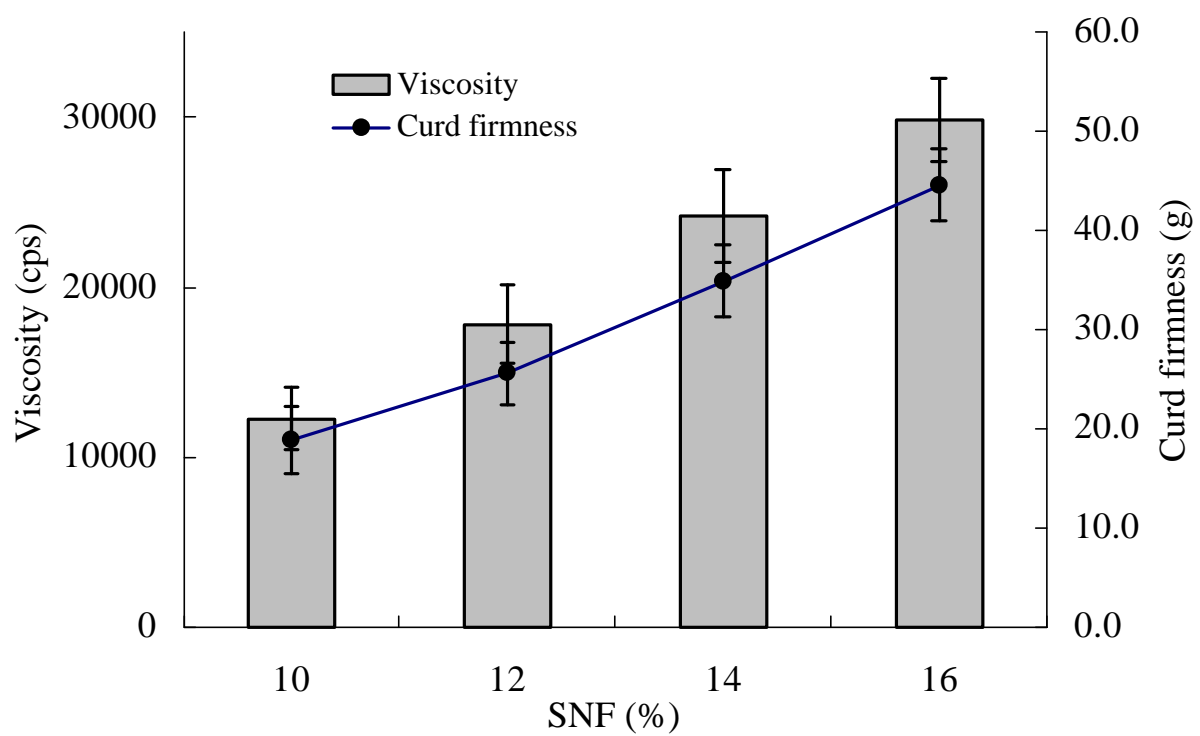


Fig. 4. Effect of SNF content on curd firmness and viscosity of curd (n = 5 ; error bars indicate 95% confidence intervals).

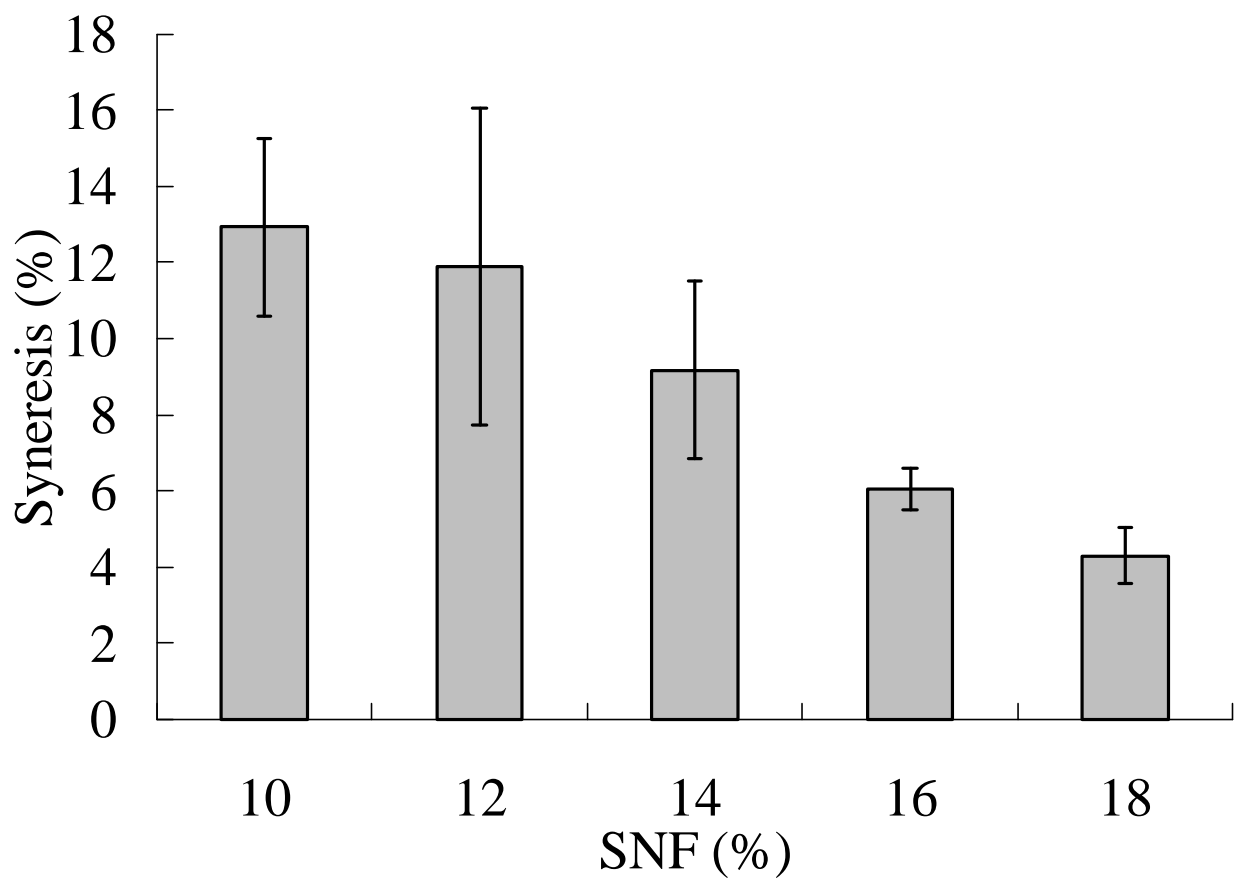


Fig. 5. Effect of SNF content on syneresis of curd (n = 5 ; error bars indicate 95% confidence intervals).

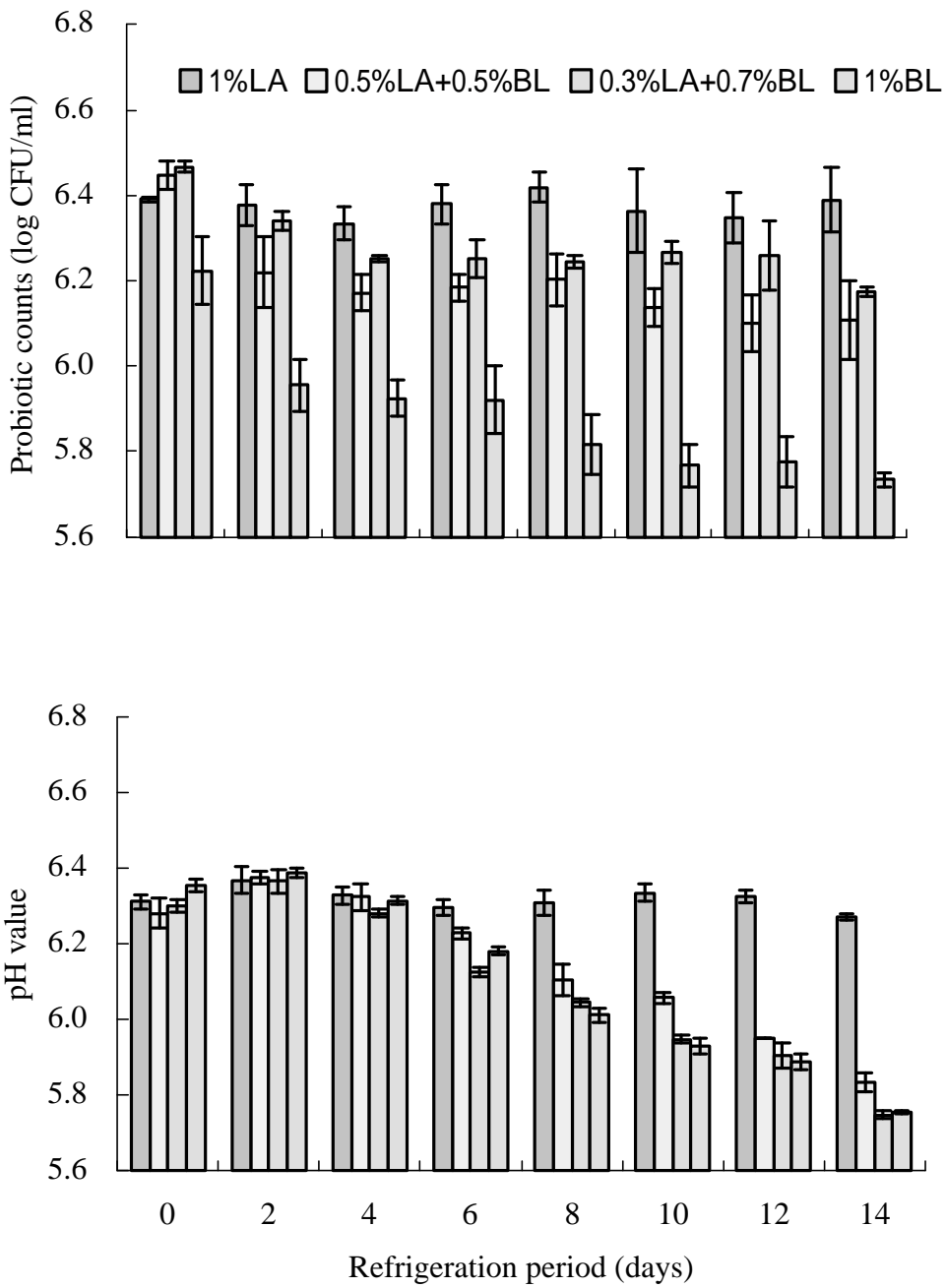


Fig. 6. Effects of different culture ratios on total probiotic counts and pH value in Kou Woan Lao during refrigeration period (n = 9 ; error bars indicate 95% confidence intervals).

Table 1. The viability of probiotics during the spray-drying process (outlet temperature : 50)

	Viability		
	<i>L. acidophilus</i>	<i>B. longum</i>	Total probiotic counts
	Log (CFU / ml)		
Emulsions (before spray-drying)	7.46 ^a ±0.05	6.99 ^a ±0.06	7.59 ^a ±0.04
stomacher	6.45 ^b ±0.03	5.99 ^b ±0.01	6.58 ^b ±0.02
After spray-drying			
vortex	6.46 ^b ±0.02	5.98 ^b ±0.03	6.58 ^b ±0.02

*Values in the same column with different letters were significantly different. (P<0.05) (n=4)

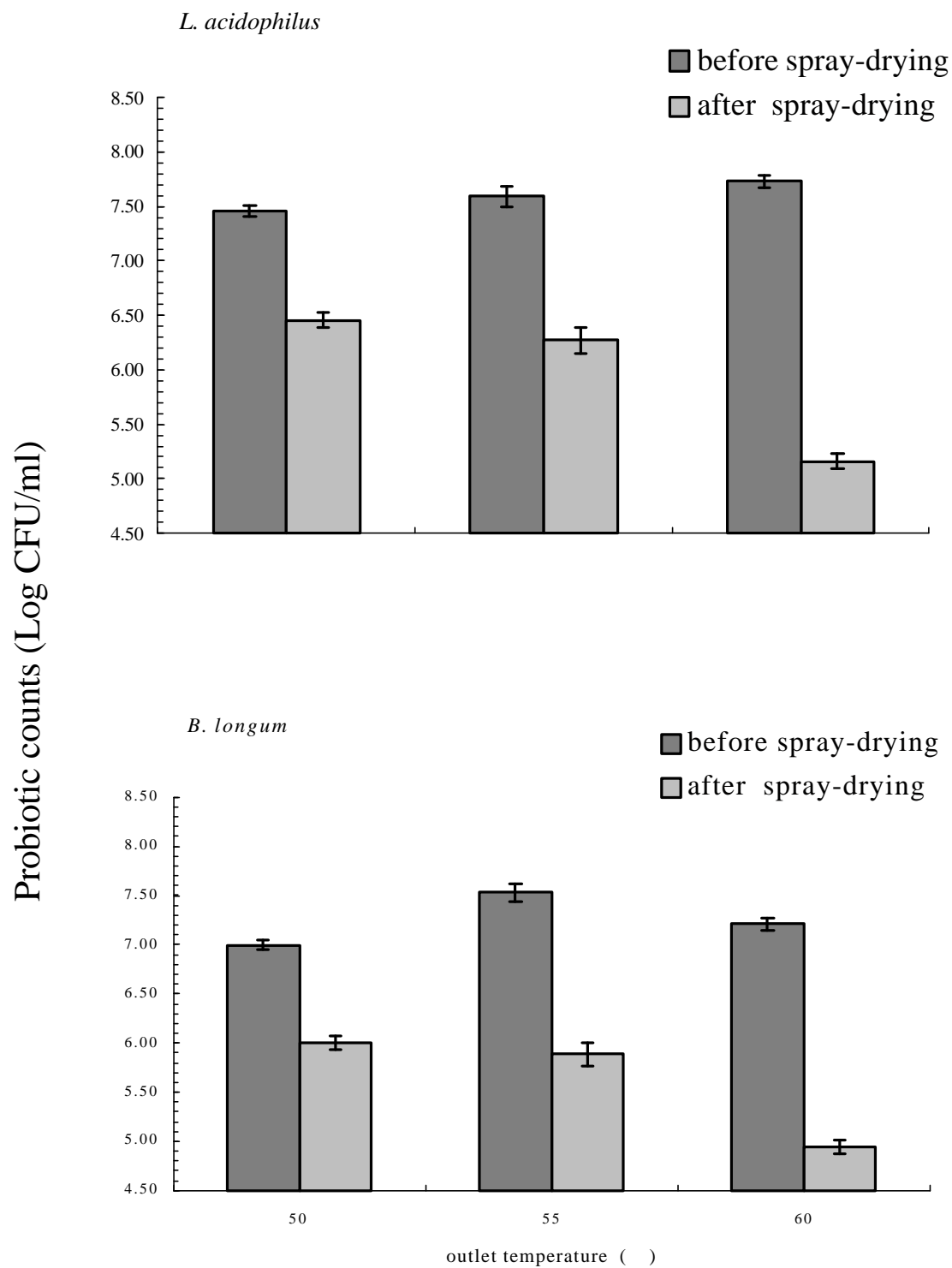


Fig. 7. Effect of outlet temperature on probiotic counts during the spray drying (n = 4 ; error bars indicate 95% confidence intervals).